

## Characterization of an Alcohol Dehydrogenase from the Cyanobacterium *Synechocystis* sp. Strain PCC 6803 That Responds to Environmental Stress Conditions via the Hik34-Rre1 Two-Component System<sup>∇†</sup>

Rebeca Vidal, Luis López-Maury,<sup>‡</sup> Miguel G. Guerrero, and Francisco J. Florencio\*

*Instituto de Bioquímica Vegetal y Fotosíntesis, CSIC—Universidad de Sevilla, Av. Américo Vespucio 49, E-41092 Sevilla, Spain*

Received 11 February 2009/Accepted 22 April 2009

**The slr1192 (*adhA*) gene from *Synechocystis* sp. strain PCC 6803 encodes a member of the medium-chain alcohol dehydrogenase/reductase family. The gene product AdhA exhibits NADP-dependent alcohol dehydrogenase activity, acting on a broad variety of aromatic and aliphatic primary alcohols and aldehydes but not on secondary alcohols or ketones. It exhibits superior catalytic efficiency for aldehyde reduction compared to that for alcohol oxidation. The enzyme is a cytosolic protein present in photoautotrophically grown *Synechocystis* cells. The expression of AdhA is enhanced upon the exposure of cells to different environmental stresses, although it is not essential for survival even under such stress conditions. The induction of the expression of the *adhA* gene is dependent on the Hik34-Rre1 two-component system, as it is severely impaired in mutant strains lacking either the histidine kinase Hik34 or the response regulator Rre1. In vitro DNA-protein interaction analysis reveals that the response regulator Rre1 binds specifically to the promoter region of the *adhA* gene.**

Medium-chain dehydrogenases/reductases (MDR) constitute a superfamily of alcohol dehydrogenases that catalyze the reversible NAD(P)-dependent oxidation of alcohols to aldehydes or ketones. It includes a large number of structurally related proteins, which catalyze several types of enzymatic activity (23, 41, 44). Screening of complete genome sequences has revealed that this family is widespread, complex, and of ancient origin (22, 44). MDR alcohol dehydrogenases are found in mammals, plants, fungi, and bacteria (52). The alcohol dehydrogenases fulfill an astonishing variety of functions in cell metabolism (21), also being a key enzyme in ethanol generation by *Saccharomyces cerevisiae* (6) and bacteria (10). Furthermore, the generation of biofuels by photoautotrophic microorganisms is of great biotechnological interest (43). Complementation of a cyanobacterium's enzyme machinery with a specific exogenous gene(s) can result in the ability to generate bioethanol from photosynthetically fixed CO<sub>2</sub> (11). Notwithstanding, current knowledge of cyanobacterial alcohol dehydrogenases is rather limited. In the cyanobacterium *Synechocystis* sp. strain PCC 6803 (referred here as *Synechocystis*), the slr1192 gene encodes a putative MDR alcohol dehydrogenase. According to in silico analyses (38, 44), the slr1192 protein has similarity with two subfamilies of MDRs: the yeast ADH family (Y-ADH) and the cinnamyl ADH family (CADH). Y-ADH-related enzymes have catabolic functions

and are involved mainly in the metabolism of ethanol or short-chain alcohols for which they exhibit broad substrate specificity. CADH and related enzymes, on the other hand, perform anabolic functions and participate in biosynthetic pathways in plants and bacteria (5, 25, 44).

In *Synechocystis*, the expression of slr1192 is induced by osmotic (35) or salt (48) stress. In higher plants, alcohol dehydrogenase activity appears to be involved in aerobic metabolism under certain stress conditions (26, 56) such as low temperature, water deficit, or ozone exposure, but its function remains unknown. A temperature decrease seems to induce the accumulation of alcohol dehydrogenase mRNA in *Arabidopsis thaliana* (20), corn, and rice (9).

In general, cyanobacteria perceive and respond to environmental changes by means of two-component regulatory systems, a ubiquitous signal transduction pathway that represents a prevalent signaling mechanism in bacteria (8, 61). Two-component systems consist of a histidine kinase (Hik) and a response regulator (Rre) and generally induce or repress the expression of specific genes in response to environmental stimuli. The histidine kinase autophosphorylates a conserved histidine residue in response to the environmental signal and then transfers the phosphate group to a conserved aspartate residue of the response regulator, which mediates the transfer of the signal. In *Synechocystis*, different Hik-Rre systems have been identified as being regulators of the response to different environmental stresses (37). A membrane-bound histidine kinase, Hik33, is involved in the perception of cold, salt, and osmotic stress (33, 35, 39, 48, 54). A cytosolic histidine kinase, Hik34, has been shown to be involved in the perception of salt and hyperosmotic stress (33, 39, 48) as well as heat shock (53). Specifically, the couples Hik33-Rre31, Hik10-Rre13, Hik16-Hik41-Rre17, Hik34-Rre1, and a putative Hik2-Rre1 have been identified as being elements involved in the perception

\* Corresponding author. Mailing address: Instituto de Bioquímica Vegetal y Fotosíntesis, Universidad de Sevilla—CSIC, Av. Américo Vespucio 49, E-41092 Sevilla, Spain. Phone: 34 954489509. Fax: 34 954460065. E-mail: floren@us.es.

<sup>†</sup> Supplemental material for this article may be found at <http://jlb.asm.org/>.

<sup>‡</sup> Present address: Fission Yeast Functional Genomics, Cancer Research UK, Wellcome Trust Sanger Institute, Morgan Building, Hinxton, Cambridge CB10 1HH, United Kingdom.

<sup>∇</sup> Published ahead of print on 1 May 2009.

TABLE 1. Primers used in this work

Oligonucleotide	Sequence (5'→3')
ADH1	ACTCTATTACATATGATTAAGCC
ADH2	TGACCATTCCACGTCGACAGAAGC
ADP1	TATTCATCGGATCCGATAACA
ADP2	GGCAGCGTAGGCTTGATCCTGG
PEX1	TTCCGTTGGCTTCCAGGGCAGCG
PEX2	CCTCTGCCTTGTCTTCATCATCGT
RR1	CAAGGATCCGTGGGGTTGAGTTTGGCTG
RR2	TAATTGCTGCAGACTGTGCATAGTTAT
S061	CATGGAGCCTAGTAATCACC
S062	GCGCTGGCATAAACCAGACC
S071	GATCGTTAGGATCATTGCG
S072	CGTCTAGAAGTGTCTGCGGGC

and transduction of signals promoted by hyperosmotic and salt stress (39, 48).

In the present work, a biochemical characterization of the *slr1192* protein (designated AdhA here) from *Synechocystis* has been performed, revealing that the tetrameric 140-kDa enzyme is active toward linear and aromatic primary alcohols and that it preferentially reduces aldehydes rather than oxidizing alcohols. In addition, an extensive analysis of the expression of the *adhA* gene has verified its induction in response to heat shock, hyperosmotic stress, salt stress, and the addition of benzyl alcohol (BA). The Hik34-Rre1 two-component system has been shown to play a relevant role in the regulation of the expression of the *adhA* gene under these stress conditions. A specific interaction of Rre1 with the promoter region of the *adhA* gene has also been demonstrated. In light of this finding and additional information presented here, the physiological role of AdhA in *Synechocystis* is discussed.

#### MATERIALS AND METHODS

**Strains and culture conditions.** *Synechocystis* sp. strain PCC 6803 cells were grown photoautotrophically on BG-11c medium (42) at 30°C under conditions of continuous illumination (50  $\mu\text{E m}^{-2} \text{s}^{-1}$ ) and bubbled with 1%  $\text{CO}_2$  in air. Heterotrophic growth was performed as described previously (4). The  $\Delta\text{hik34}$  and  $\Delta\text{rre1}$  mutant strains were kindly provided by Iwane Suzuki (Graduate School of Life and Environmental Sciences, University of Tsukuba, Japan) (55). For precultures of the  $\Delta\text{adhA}$  or  $\Delta\text{rre1}$  strain, kanamycin was added to a final concentration of 100  $\mu\text{g ml}^{-1}$ , and for precultures of the  $\Delta\text{hik34}$  strain, spectinomycin was added to a final concentration of 5  $\mu\text{g ml}^{-1}$ . Experiments were performed using cultures from the mid-logarithmic phase (optical density at 730 nm of 0.6 to 0.7 and 3 to 5  $\mu\text{g chlorophyll ml}^{-1}$ ). For high-light treatment, cell suspensions were diluted with fresh medium to an optical density at 730 nm of 0.35, and the cultures were placed in a temperature-controlled chamber at 30°C and exposed to an irradiance of 500  $\mu\text{E m}^{-2} \text{s}^{-1}$ . Under heat shock conditions, cultures were incubated in a water bath at 45°C. For salt or hyperosmotic shock, NaCl or sorbitol, respectively, was added to the cell suspension to a final concentration of 0.5 M. BA was added to a final concentration of 30 mM. For oxidative stress,  $\text{H}_2\text{O}_2$  was added to a final concentration of 1 mM. Glucose and 3-(3P,4P-dichlorophenyl)-1,1P-dimethylurea were added to final concentrations of 5 mM and 10  $\mu\text{M}$ , respectively.

*Escherichia coli* DH5 $\alpha$  and *E. coli* BL21(DE3) cells were grown in LB broth medium as described previously (46) and supplemented with 100  $\mu\text{g ml}^{-1}$  of ampicillin or 50  $\mu\text{g ml}^{-1}$  of kanamycin when required. M9 minimal medium (46) was used for the metal dependence analysis of His-AdhA expression.

**Construction of the *adhA* disruption mutant.** To inactivate the *adhA* gene, the entire coding region was amplified by PCR to yield a fragment of 1,011 bp using primer pairs ADH1 and ADH2. All the primers used in this work are listed in Table 1. The PCR product was cloned into the pGEM-T vector to generate plasmid pADH1. A DNA fragment containing the *npt* gene (neomycin phosphotransferase) under the control of the *Synechocystis* sp. strain PCC 6803 *psbA* promoter (13) was introduced into the HindIII site of the coding region of *adhA* (712 bp downstream

of the start codon), generating a construct that was used to transform *Synechocystis* sp. strain PCC 6803 (14). Transformants were selected by screening for resistance to 50  $\mu\text{g ml}^{-1}$  of kanamycin in BG-11 medium plates. Segregation of the inactivated *adhA* gene was monitored by Southern blotting using standard procedures (46). Total DNA from cyanobacteria was isolated as previously described (7).

**Expression and purification of the AdhA protein.** To generate a plasmid for the expression of recombinant N-terminal His-tagged AdhA, the PCR fragment obtained using primers ADH1 and ADH2 was digested with NdeI and SalI and cloned into the same restriction sites of the pET-28a(+) vector (Novagen) to generate plasmid pETAdhA. The DNA fragments cloned were totally sequenced to ensure that no modifications in the nucleotide sequence occurred during cloning. For the expression of the AdhA protein, *E. coli* BL21(DE3) was transformed with plasmid pETAdhA. For large-scale protein production, the transformant cells were grown at 37°C in LB broth to an optical density at 580 nm of 0.5, 1 mM isopropyl- $\beta$ -D-thiogalactopyranoside (IPTG) was added, and the culture was incubated for 2.5 h. Thereafter, cells were harvested by centrifugation, resuspended in 20 mM Tris-HCl (pH 7.9) supplemented with 50 mM NaCl and 1 mM phenylmethylsulfonyl fluoride, and broken by sonication. The insoluble debris was pelleted by centrifugation at 18,000  $\times g$  for 30 min, and the supernatant was used immediately for protein purification. AdhA was purified by anion-exchange chromatography using DEAE-cellulose resin equilibrated with 20 mM Tris-HCl (pH 7.5) and applying a 0 to 0.5 M NaCl gradient. The eluted fractions containing the protein were loaded onto Ni-nitrilotriacetic acid (NTA) resin (Novagen) and eluted with 0.4 M imidazole. Protein samples were examined by 12% sodium dodecyl sulfate-polyacrylamide gel electrophoresis (SDS-PAGE) (46).

**Gel filtration.** A 200- $\mu\text{l}$  aliquot of native AdhA (30  $\mu\text{M}$ ) was applied to a Superose 6 10/300 GL (GE Healthcare) column connected to a fast-performance liquid chromatograph (Äkta; Amersham Biosciences) at 0.8 ml  $\text{min}^{-1}$ . The column was previously equilibrated with 30 mM Tris-HCl (pH 8.0) supplemented with 0.5 M NaCl. The molecular mass standards used were bovine thyroglobulin (670,000 Da), bovine  $\gamma$ -globulin (158,000 Da), chicken ovalbumin (44,000 Da), horse myoglobin (17,000 Da), and vitamin B<sub>12</sub> (1,350 Da).

**Expression and purification of the Rre1 protein.** A DNA fragment covering from +1 to +846 bp of *slr1783* (Cyanobase) was amplified by PCR using primers RR1 and RR2, digested with BamHI and PstI, and cloned into the same restriction sites of vector pQE-80L to generate plasmid pQR1, which was used to produce a recombinant N-terminal His-tagged Rre1 protein. The DNA fragment cloned was totally sequenced to ensure that no modifications in the nucleotide sequence occurred during cloning. For the expression of the Rre1 protein, IPTG-inducible *E. coli* strain DH5 $\alpha$  transformed with plasmid pQR1 was grown at 37°C in LB broth to an optical density at 580 nm of 0.5, 1 mM IPTG was then added, and the culture was incubated for 2.5 h. Thereafter, cells were harvested by centrifugation, resuspended in 20 mM Tris-HCl (pH 7.9) supplemented with 50 mM NaCl and 1 mM phenylmethylsulfonyl fluoride, and broken by sonication. The insoluble debris was pelleted by centrifugation at 18,000  $\times g$  for 30 min, and the supernatant was used immediately for protein purification. Rre1 was purified by affinity chromatography using Ni-NTA resin (Novagen).

**Alcohol dehydrogenase assay.** Enzyme activity was assayed at 25°C by monitoring changes in the absorbance of NAD(P)H at 340 nm in a Pharmacia LKB Ultrospec Plus spectrophotometer with a 1-cm-light-path cuvette. Reactions were performed using 30 mM Tris-HCl buffer (pH 8.0) containing 0.2 mM NAD(P)H for the reduction of aldehydes or 0.5 mM NAD(P)<sup>+</sup> for the oxidation of alcohols. The kinetic constants were determined by fitting the initial rates, calculated in the linear range of protein concentration and reaction time, to the Michaelis-Menten equation, and the corresponding values are expressed as the means  $\pm$  standard deviations for three independent determinations.

**RNA isolation and Northern blot analysis.** Total RNA was isolated from 30-ml samples of *Synechocystis* sp. strain PCC 6803 cultures in the mid-exponential growth phase (3 to 5  $\mu\text{g chlorophyll ml}^{-1}$ ) as previously described (17). For Northern blotting, 15  $\mu\text{g}$  of total RNA was loaded per lane in 1% (wt/vol) agarose denaturing formaldehyde gels and transferred onto nylon membranes (Hybond N<sup>+</sup>; Amersham Biosciences). Hybridization was performed at 65°C. The *adhA*, *slr1106*, and *slr1107* probes were synthesized by PCR using primer pairs ADH1/ADH2, S061/S062, and S071/S072, respectively, and <sup>32</sup>P labeled with a random-primer kit (Amersham Biosciences) with [ $\alpha$ -<sup>32</sup>P]dCTP (3,000 Ci  $\text{mmol}^{-1}$ ). All filters were stripped and rehybridized with the constitutively expressed *mipB* gene from *Synechocystis* sp. strain PCC 6803 (59).

**Preparation of AdhA polyclonal antibodies.** A homogeneous sample of the protein was used for the production of antibodies at the Animal Production and Experimentation Service of Seville University (Seville, Spain).

**Preparation of cytosolic extracts from *Synechocystis* and Western blot analysis.** Cytosolic protein extracts from *Synechocystis* were obtained as described previously (40). Cytosolic proteins (5  $\mu\text{g}$ ) from *Synechocystis* cultures exposed to

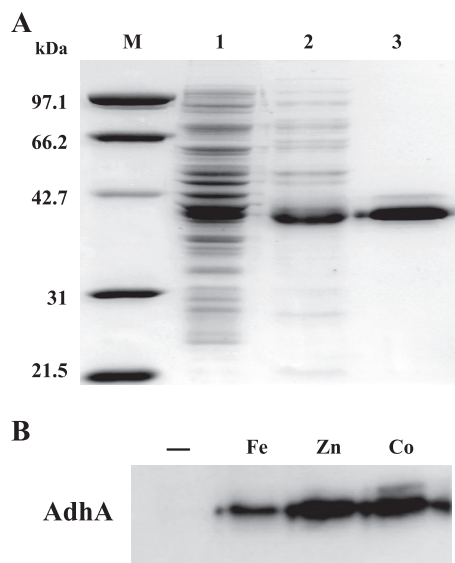


FIG. 1. Purification and metal dependence of AdhA. (A) Coomassie-stained SDS-PAGE gel of different fractions corresponding to steps of AdhA purification (0.5  $\mu$ g total protein per lane). M, molecular mass markers. Lane 1, crude extract from *E. coli* cells that overexpress His-tagged AdhA; lane 2, pooled DEAE-cellulose chromatography fractions containing His-tagged AdhA; lane 3, pooled affinity chromatography (Ni-NTA) fractions containing His-tagged AdhA. (B) Western blotting of soluble extracts of AdhA-overexpressing *E. coli* cells grown on M9 medium. —, no metal added; Fe, 5  $\mu$ M  $(\text{NH}_4)_2\text{Fe}(\text{SO}_4)_2$ ; Zn, 5  $\mu$ M  $\text{ZnSO}_4$ ; Co, 5  $\mu$ M  $\text{CoCl}_2$ .

different conditions were separated by SDS-PAGE and transferred onto nitrocellulose membranes. Membranes were incubated with diluted anti-AdhA (1:1,000) overnight at 4°C. Detection was performed using ECL Plus Western blotting detection reagents (Amersham Biosciences) according to instructions provided by the manufacturer.

**Gel mobility shift assay.** A probe was synthesized by PCR using oligonucleotide pair ADP1 and ADP2, both containing a BamHI restriction site. The PCR product was digested with BamHI, generating a fragment from positions –215 to +22 with respect to the transcription start point of *adhA*, which was end labeled with  $[\alpha\text{-}^{32}\text{P}]\text{dCTP}$  (3,000 Ci  $\text{mmol}^{-1}$ ) by using the Klenow fragment of DNA polymerase. The binding reaction was carried out using a final volume of 30  $\mu$ l containing 0.1 ng of labeled DNA (2 pM) and 3  $\mu$ g of poly(dI-dC) in 50 mM Tris-HCl (pH 8.0), 100 mM KCl, 15 mM dithiothreitol, 10% glycerol, and different amounts of protein. Reaction mixtures were incubated for 30 min at room temperature, and samples were loaded onto a nondenaturing 6% polyacrylamide gel. Electrophoresis was carried out at 4°C and 180 V in 22 mM Tris-borate–0.5 mM EDTA. The gel was transferred onto a Whatman 3-mm filter and dried prior to autoradiography. For competition assays, a 222-bp DNA fragment from pGEM-T digested with SspI was used as nonspecific competitor DNA.

**Primer extension analysis.** Oligonucleotides PEX1 and PEX2 were end labeled with T4 polynucleotide kinase and  $[\gamma\text{-}^{32}\text{P}]\text{ATP}$  (3,000 Ci  $\text{mmol}^{-1}$ ) and used for primer extension analysis of the *adhA* and *sll1106* genes, respectively. Primer extension reactions were carried out as previously described (31). One-half of the reaction mixture was subjected to electrophoresis on a 6% polyacrylamide sequencing gel, and the *adhA* and *sll1106* promoter regions were subjected to a sequencing reaction using the oligonucleotides described above and plasmid pEXAdh as a template.

## RESULTS

**AdhA is a tetrameric medium-chain Zn-containing alcohol dehydrogenase.** In order to gain knowledge on the activity and properties of the AdhA protein, the *adhA* gene was cloned, and a His<sub>6</sub>-tagged version of the protein was expressed in *E.*

*coli* cells. Homogeneous preparations of AdhA were obtained by DEAE-cellulose ion-exchange chromatography followed by affinity chromatography on Ni-NTA (Fig. 1A). Purification attempts using extracts of *E. coli* cells grown on M9 chemically defined medium revealed that the presence of divalent metals in the medium was a requisite for the availability of the enzyme in a soluble form (Fig. 1B). Thus, when metals were absent from the growth medium, expressed AdhA did not appear in the soluble fraction but appeared there, however, if the medium was supplemented with zinc, cobalt, or ferrous ions (Fig. 1B). Differences in the specific activities of the enzyme purified from cultures grown on media with different metal contents showed that AdhA can use either zinc or cobalt with analogous effectivenesses ( $2 \pm 0.2$  or  $1.9 \pm 0.12$  mU  $\cdot$  mg total protein<sup>–1</sup>, respectively), while ferrous ion was less efficient ( $0.3 \pm 0.05$  mU  $\cdot$  mg total protein<sup>–1</sup>). Analysis of the amino acid sequences (see Fig. S1 in the supplemental material) revealed the presence of conserved residues involved in the binding of both catalytic (Cys<sub>55</sub>, His<sub>78</sub>, Cys<sub>176</sub>, Asp<sub>58</sub>, and Glu<sub>79</sub>) and structural (Cys<sub>111</sub>, Cys<sub>114</sub>, Cys<sub>117</sub>, and Cys<sub>125</sub>) zinc ions (23, 30, 52).

To establish the quaternary structure of the protein, gel filtration on a Superose 6 10/300 GL column were performed using purified preparations of recombinant AdhA, and an apparent molecular mass of 140 kDa was determined. Taking into account the polypeptide mass (36.7 kDa) of the recombinant protein calculated from migration on SDS-PAGE gels (Fig. 1A), a tetrameric structure is suggested for the AdhA active enzyme.

A thorough substrate specificity analysis as well as a kinetic characterization of AdhA have been performed. As shown in Table 2, the enzyme was active toward a wide variety of primary alcohols and their corresponding aldehydes, but neither ketones nor secondary alcohols were effective substrates. Aliphatic compounds were efficiently processed by the enzyme, with the activity increasing with the length of the chain up to

TABLE 2. Substrate specificity of AdhA from *Synechocystis*<sup>c</sup>

Reduction <sup>a</sup>	Relative activity (%)		Oxidation <sup>b</sup>	Relative activity (%)	
	0.1 mM	1 mM		1 mM	10 mM
Acetaldehyde	1	23	Ethanol	19	91
Propanal	26	87	Propanol	21	84
Butanal	73	120	Butanol	23	70
Pentanal	100	107	Pentanol	31	52
Hexanal	53	100	Hexanol	27	40
Octanal	42	80	Octanol	8	17
Decanal	21	22	Decanol	3	6
Benzaldehyde	30	14	Benzyl alcohol	42	79
Cinnamaldehyde	61	7	Cinnamyl alcohol	100	116
Acetone	ND	ND	Phenylethanol	15	24
3-Pentanone	ND	ND	2-Methylbutanol	24	22
d-Glucose	ND	ND	3-Pentanol	ND	ND

<sup>a</sup> The reduction activities were measured using 30 mM Tris-HCl (pH 8.0) containing 0.1 mM or 1 mM substrate in the presence of 0.2 mM NADPH. The activity toward 0.1 mM pentanal (67.7 U  $\text{mg}^{-1}$  protein) was considered to be 100%.

<sup>b</sup> The oxidation activities were measured using 30 mM Tris-HCl (pH 8.0) containing 1 mM or 10 mM substrate in the presence of 0.5 mM NADP<sup>+</sup>. The activity toward 1 mM cinnamyl alcohol (26.1 U  $\text{mg}^{-1}$  protein) was considered to be 100%.

<sup>c</sup> ND, not detected.

TABLE 3. Kinetic parameters of AdhA from *Synechocystis*<sup>a</sup>

Substrate	Mean $K_m$ (mM) $\pm$ SD <sup>a</sup>	Mean $k_{cat}$ (min <sup>-1</sup> ) $\pm$ SD	$k_{cat}/K_m$ ratio (min <sup>-1</sup> mM <sup>-1</sup> )
Acetaldehyde	0.510 $\pm$ 0.050	9,458 $\pm$ 138	18,545
Butanal	0.110 $\pm$ 0.008	9,398 $\pm$ 181	84,664
Pentanal	0.075 $\pm$ 0.002	9,547 $\pm$ 185	123,295
Cinnamaldehyde	0.017 $\pm$ 0.001	6,475 $\pm$ 131	380,882
Ethanol	18.800 $\pm$ 3.300	1,641 $\pm$ 33	87
Butanol	3.280 $\pm$ 0.100	1,023 $\pm$ 21	312
Pentanol	1.540 $\pm$ 0.060	789 $\pm$ 15	512
Cinnamyl alcohol	0.270 $\pm$ 0.010	3,647 $\pm$ 73	13,507
NADPH	0.025 $\pm$ 0.007		
NADP	0.050 $\pm$ 0.007		
NADH	1.310 $\pm$ 0.060		
NAD	9.700 $\pm$ 0.800		

<sup>a</sup> Reactions were performed using 30 mM Tris-HCl (pH 8.0). For the reduction of aldehydes and oxidation of alcohols, 0.15 mM NAD(P)H and 0.4 mM NAD(P)<sup>+</sup> were used, respectively. Kinetic constants for NAD(P)<sup>+</sup> and NAD(P)H were determined with 100 mM ethanol and 25 mM acetaldehyde, respectively.

five carbon atoms and decreasing for longer chains. Aromatic alcohols and aldehydes were also substrates for the enzyme, as was the case for the BA/benzaldehyde or cinnamyl alcohol/cinnamaldehyde. Branched-chain primary alcohols, such as 2-methyl-butanol, were processed less efficiently than the linear substrate (Table 2). The  $K_m$  and  $k_{cat}$  values were calculated for several substrate pairs of alcohols and their corresponding aldehydes, and they are shown in Table 3. The values of  $k_{cat}$  were significantly higher for aldehydes than for alcohols. Cinnamaldehyde and pentanal behaved as the substrates most efficiently processed by the AdhA protein, with the highest  $k_{cat}/K_m$  ratios. The enzyme could use both NADPH and NADH as electron donors, although NADPH ( $K_m = 0.025$  mM) was preferred to NADH ( $K_m = 1.3$  mM) as the reductant for aldehydes. Equally, that affinity for NADP<sup>+</sup> was higher than that for NAD<sup>+</sup> as an alcohol oxidant. Thus, it is likely that AdhA works to reduce aldehydes to alcohols.

**Expression of *adhA* is induced under various stress conditions.** The *adhA* gene was previously identified by DNA microarray analysis to be a hyperosmotic-stress- and salt stress-inducible gene (35, 48). Using Northern blot hybridization

(Fig. 2), we have confirmed that the level of transcription of *adhA* increases in response to either hyperosmotic stress (0.5 M sorbitol) or salt stress (0.5 M NaCl). In order to determine whether *adhA* expression is specifically enhanced in response to the osmotic stress induced by the addition of sorbitol or NaCl or as a part of a general stress response, *Synechocystis* cells were exposed to different stress conditions such as heat shock, high-light conditions, and oxidative stress, and changes in levels of *adhA* transcripts were monitored. The exposure of cells to a sudden increase in the ambient temperature (45°C) also resulted in enhanced *adhA* transcript levels (Fig. 2). However, neither high-light treatment nor oxidative stress (1 mM H<sub>2</sub>O<sub>2</sub>) had an influence on the transcription of the *adhA* gene (data not shown). It was previously reported (19) that many of the genes that respond to heat shock are also induced when BA, a membrane-fluidizing agent (18), is added to the culture medium. As shown in Fig. 2, we have confirmed that BA also induces *adhA* gene expression, although the kinetics of *adhA* transcript induction were different. Thus, the induction of *adhA* expression was transient under conditions of heat shock exposure, with the maximum transcript level being reached in less than 1 h and returning to the steady-state level in 2 h (Fig. 2). However, for the response to BA, the maximum transcript level was reached at 5 h. On the other hand, the maximum level of *adhA* transcript was reached in less than 1 h and then decreased at 2 h under conditions of salt stress. In response to hyperosmotic shock, the kinetics were very similar to those of salt stress, but the decrease in the transcript level at 2 h was followed by another increase to reach the maximum level at 5 h. In both cases, the transcription of the *adhA* gene remained induced at least 8 h after treatment and returned to basal levels after 24 h (data not shown). In addition to the information on the induction of the expression of *adhA*, Fig. 2 includes also data for the expression of the nearby *sll1106* and *sll1107* genes, which exhibited a pattern analogous to that of *adhA*. This circumstance is considered below.

In order to analyze the relationship between *adhA* transcript and AdhA protein levels, Western blot experiments were performed. A basal amount of AdhA was detected in *Synechocystis* cells grown under photoautotrophic conditions. As shown in

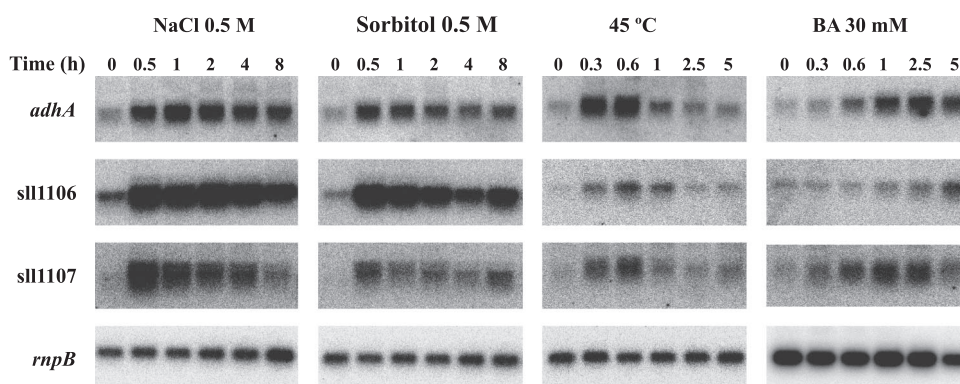


FIG. 2. Northern blotting analysis of the stress-inducible expression of the *adhA*, *sll1106*, and *sll1107* genes. *Synechocystis* sp. strain PCC 6803 cells at the exponential phase of growth were subjected to various conditions, and total RNA was extracted at different time points. Fifteen micrograms of total RNA was denatured, separated by electrophoresis in a 1.2% agarose gel, blotted, and hybridized with probes for the *adhA*, *sll1106*, and *sll1107* genes. The filters were stripped and hybridized with an *rnpB* gene probe as a loading control. All probes were synthesized as described in Materials and Methods. Three independent experiments were performed, and data plotted are the averages  $\pm$  standard errors.

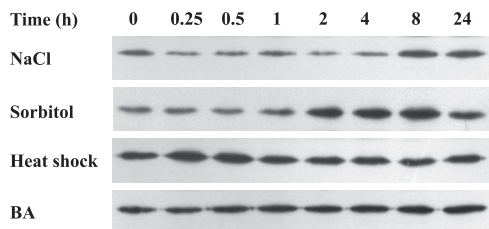


FIG. 3. Western blotting analysis of the stress-inducible expression of the AdhA protein. *Synechocystis* sp. strain PCC 6803 cells at the exponential phase of growth were subjected to various conditions as indicated. Cells were harvested at the times indicated, and 5 µg of total protein from soluble extracts was separated by 12% SDS-PAGE and subjected to Western blotting.

Fig. 3, the AdhA protein level increased under conditions of salt and hyperosmotic stresses in accordance with the observed induction of transcript levels. The kinetics of AdhA accumulation differed depending on the stress condition imposed (Fig.

3). While under hyperosmotic conditions, an increase in AdhA levels was readily observed after 2 h of treatment, under salt stress conditions, the protein started to accumulate after 8 h (Fig. 3). This difference could be explained by the fact that protein synthesis was previously reported to be nearly blocked within 4 h after the onset of salt stress (15). In contrast, the amount of the AdhA protein remained virtually unaltered under conditions of heat shock and upon the addition of BA. Induction experiments performed in the presence of chloramphenicol confirmed that no degradation of AdhA took place under any condition tested (data not shown).

The *sll1106* and *sll1107* genes share the same expression pattern as *adhA*. Immediately upstream of the *adhA* gene in the opposite DNA strand are two open reading frames, *sll1106* and *sll1107* (Fig. 4A). One of them, *sll1107*, was also identified, together with the *adhA* gene, as being a salt-inducible gene in a genome-wide DNA microarray assay (48); both genes were shown to be under the control of the Hik34-Rre1 two-compo-

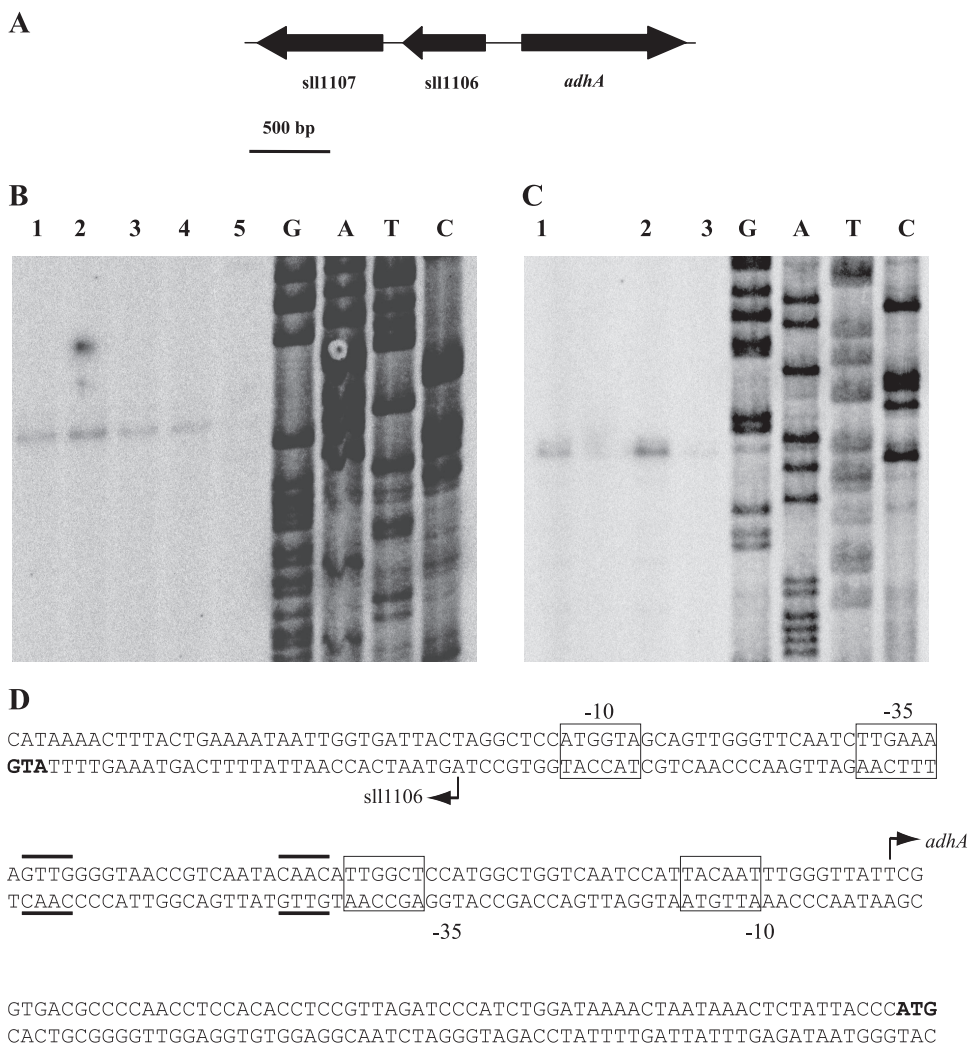


FIG. 4. Primer extension analysis of *adhA* and *sll1106* transcripts. (A) Schematic representation of the genomic region corresponding to the *adhA*, *sll1106*, and *sll1107* genes. (B) Primer extension analysis of *adhA* transcripts from *Synechocystis* cells exposed for 1 h to 30 mM BA (lane 1), heat shock (45°C) (lane 2), 0.5 M NaCl (lane 3), and 0.5 M sorbitol (lane 4) and from nonexposed cells (lane 5). (C) Primer extension analysis of *sll1106* transcripts from *Synechocystis* cells exposed to heat shock (45°C) (lane 1) and 0.5 M sorbitol (lane 2) and from nonexposed cells (lane 3). (D) Sequence of the *adhA*-*sll1106* intergenic region, with translation start codons in boldface type. The transcripts start points are marked with arrows. Putative -10 and -35 boxes are indicated with rectangles.

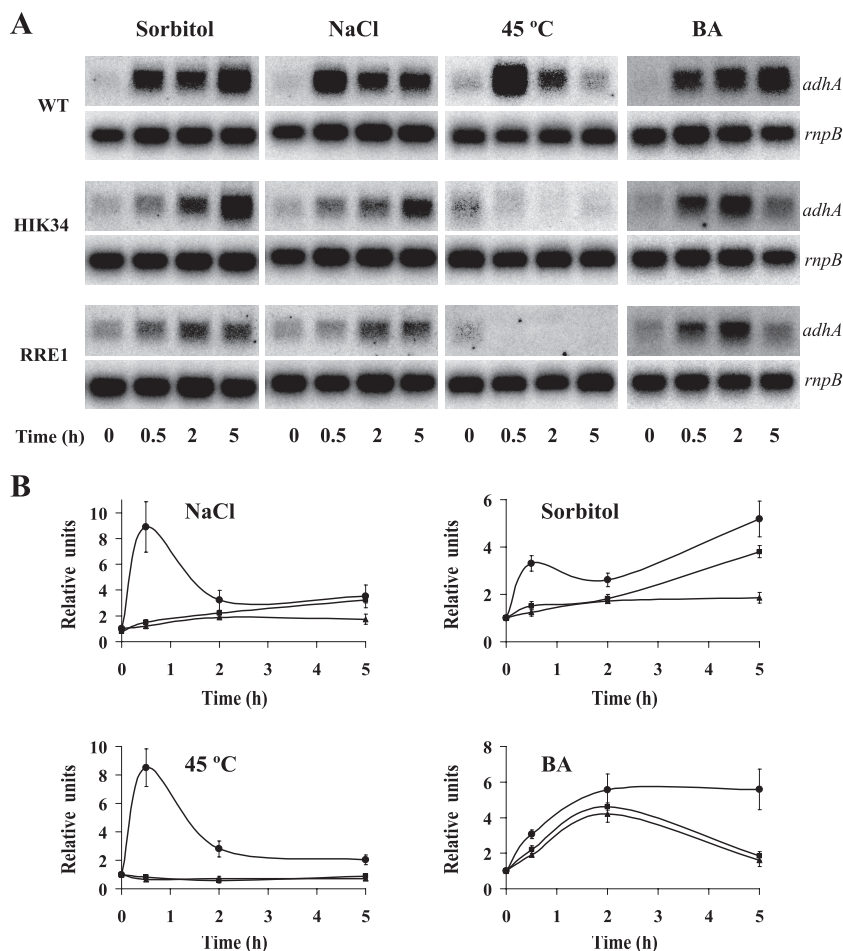


FIG. 5. Expression of the *adhA* gene under stress conditions in wild-type (WT),  $\Delta$ *hik34*, and  $\Delta$ *rre1* strains. (A) Northern blot analysis. Cultures at the exponential phase of growth were subjected to various stress conditions, and total RNA was extracted at different time points. Fifteen micrograms of total RNA was denatured, separated by electrophoresis in a 1.2% agarose gel, blotted, and hybridized with *adhA*. As a loading control, all the filters were rehybridized with *rnpB*. (B) Kinetics of expression of the *adhA* transcripts. The values in the graphs indicate the ratio of *adhA* transcripts in stressed cells to those in control cells. Three independent experiments were performed, and data plotted are the averages  $\pm$  standard errors. ●, wild-type strain; ■,  $\Delta$ *hik34* mutant strain; ▲,  $\Delta$ *rre1* mutant strain.

ment system. The proximity between *sll1106* and *adhA* and the coregulation of *sll1107* prompted us to test for a possible coordinated regulation of the *sll1106*, *sll1107*, and *adhA* genes. In order to test this hypothesis, filters used for Northern hybridizations with *adhA* were rehybridized with probes corresponding to *sll1106* and *sll1107*. The results (Fig. 2) showing the *sll1106* and *sll1107* genes to be induced under the same conditions that induced the expression of *adhA* suggest a possible coregulation in which the *sll1106-adhA* intergenic region could contain the promoter and the regulatory sequences for the expression of the three genes. However, transcripts of different sizes (516 nucleotides for *sll1106* and 777 nucleotides for *sll1107*) were detected by Northern blotting, indicating that both genes are probably transcribed under the control of independent promoters. On the other hand, the *sll1106* and *adhA* genes are separated by a 200-bp region. In order to establish the location of the *adhA* and *sll1106* promoter(s), starting points of *adhA* and *sll1106* transcripts were mapped by primer extension experiments. One single extension product, which appeared as a double band, was obtained for both genes

when RNA extracted from cells subjected to stress conditions was used (Fig. 4B and C). The *adhA* transcription start site was located 69 bp upstream of the ATG start codon, while the *sll1106* start site was located 31 bp upstream of the corresponding ATG start codon (Fig. 4D). Both sites were separated by a region of 100 bp, which might contain the promoters for both genes.

**The two-component system Hik34-Rre1 regulates the induction of *adhA* expression in response to stress conditions.** The *adhA* gene was previously identified as being a salt-stress-inducible gene under the control of the Hik34-Rre1 two-component system (48). In order to clarify if the histidine kinase Hik34 and the response regulator Rre1 regulate the expression of the *adhA* gene in response to the other conditions tested (Fig. 2), the pattern of expression of the *adhA* gene in wild-type cells was compared to that of  $\Delta$ *hik34* and  $\Delta$ *rre1* mutant strains. As shown in Fig. 5, the expression levels of the *adhA* gene in response to heat shock or BA were similar for both the  $\Delta$ *hik34* and  $\Delta$ *rre1* mutants and differed from that of the wild-type strain. In the case of heat shock, the induction of the expres-

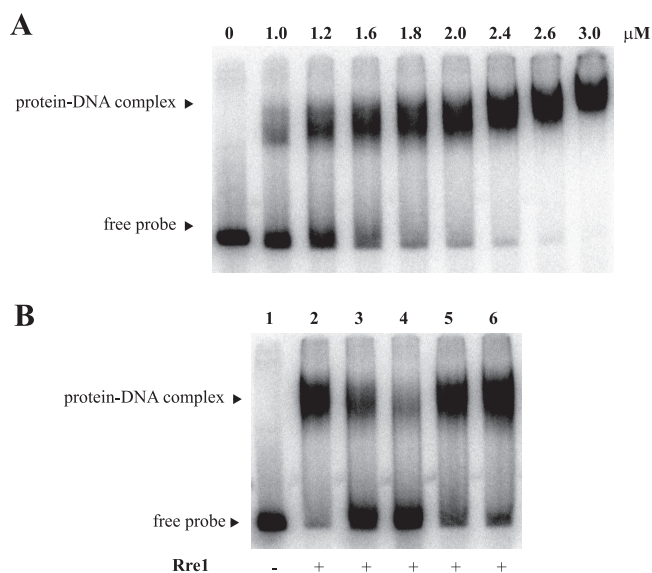


FIG. 6. Binding of Rre1 to the *adhA*-sll1106 promoter region. (A) Mobility shift assay of the *adhA*-sll1106 promoter-operator region with increasing amounts of Rre1. (B) Competitive assays were performed with Rre1 (1.6 μM) in the presence of either a 50-fold (lanes 3 and 5) or a 200-fold (lanes 4 and 6) excess of unlabeled self-competitor fragment (lanes 3 and 4) or unlabeled nonspecific competitor fragment (lanes 5 and 6).

sion of the *adhA* gene was completely abolished in both mutants. In the case of BA, the effect of either mutation was noticeable only at 5 h after treatment (Fig. 5). When mutant cells were exposed to either salt or hyperosmotic shock, the early induction of the *adhA* gene (30 min after exposure to stress) was abolished. In the case of the  $\Delta rre1$  mutant, this situation was maintained with time, although this was not the case for the  $\Delta hik34$  mutant.

**Rre1 binds to the promoter region of the *adhA* gene.** In order to test if the response regulator Rre1 affects the expression of the *adhA* gene directly by interactions with its promoter region, a DNA binding shift assay was performed. A radiolabeled DNA fragment corresponding to the sll1106-*adhA* intergenic region was incubated with increasing amounts of the purified Rre1 protein (see Materials and Methods). A single shifted band representing a DNA-protein complex was observed upon the addition of Rre1 (Fig. 6A). Sequence-specific binding was evident from competition assays in the presence of different concentrations of either specific or nonspecific competitor DNA. The Rre1-dependent band shift was diminished in the presence of a 200-fold excess of the same unlabeled fragment but was unaffected by the presence of an excess of the unlabeled DNA fragment (Fig. 6B).

## DISCUSSION

To the best of our knowledge, this is the first report of the molecular characterization of an alcohol dehydrogenase in cyanobacteria. We show that the protein encoded by the *Synechocystis adhA* gene is a zinc-dependent homotetrameric enzyme with NADP-dependent alcohol dehydrogenase activity acting on a broad variety of substrates. The higher catalytic efficiency

of *Synechocystis AdhA* for aldehyde reduction, with pentanal and cinnamaldehyde as the preferred substrates, points to the reduction of aromatic and medium-chain aliphatic aldehydes as a most plausible *in vivo* activity of the enzyme. *Synechocystis AdhA* exhibits a preferential specificity for NADP(H) but is also able to use NAD(H). This may be explained by the presence of a threonine at position 226 (see Fig. S1 in the supplemental material), which is more similar to the serine residue of the known NADPH-dependent ADH (52) than to the aspartate or glutamate residue of NADH-dependent ADH (29, 34). Thus, the *Synechocystis AdhA* enzyme shares biochemical features with both Y-ADH enzymes, which are tetrameric NAD-dependent enzymes, and CADH enzymes, which are dimeric and NADP dependent.

A search performed with the AdhA sequence (slr1192 protein) as a query using a nonredundant protein sequence database of the NCBI revealed more than 60% identity with putative MDRs from gammaproteobacteria. Among cyanobacteria, homologous sequences showing about 50 to 60% identity were found in some strains of *Microcystis* and *Synechococcus*. Sequence alignment of AdhA with related members of the MDR family from bacteria, plants, and yeasts yielded different clusters, with bacterial sequences forming a closed group independent from CADH and Y-ADH groups (see Fig. S2 in the supplemental material).

In plants, the function of the CADH family is related to lignin biosynthesis (5, 25). In bacteria, from which lignin is absent, CADH-like genes are related to the biosynthesis of lipids of the cell envelope (16, 62), but it is possible that they fulfill other specific functions. An alcohol dehydrogenase from *Acinetobacter* that shares 46% identity with the *adhA* gene from *Synechocystis* has been proposed to be involved in the biosynthesis of wax esters as cell reserves when cells are grown on *n*-alkanes (57). On the other hand, some Y-ADH-related enzymes, which show broad substrate specificity, fulfill a function related to primordial metabolic pathways such as alcohol fermentative activity (49). A variety of different fermentation pathways yielding products such as CO<sub>2</sub>, H<sub>2</sub>, formate, acetate, lactate, and ethanol have been reported to exist in cyanobacteria (50). A number of cyanobacteria are capable of heterotrophic growth under dark conditions at the expense of either endogenous glycogen or exogenous substrates. In most cases, heterotrophic growth takes place under aerobic conditions, but some of the facultative heterotrophic cyanobacteria are able to survive under dark anoxic conditions via fermentation of the glucose derived from endogenous substrates such as glycogen or some osmoprotectant compounds (50). Thus, AdhA might participate in any of such fermentative pathways in *Synechocystis*.

Here we show that a soluble AdhA protein is present in *Synechocystis* cells and that its expression is enhanced upon the exposure of cells to stress conditions. Although the level of the *adhA* transcript increases, the AdhA protein level remains unchanged under conditions of heat shock, making it unlikely that AdhA plays a role in the response to this condition. On the other hand, the induction of *adhA* transcription under conditions of salt stress or hyperosmotic stress is accompanied by an increase in the amount of AdhA. Both salt and hyperosmotic stress cause alterations in the physical integrity of the plasma membrane (24) and affect the activity of photosystem I (PSI)

and photosystem II in cyanobacteria (1–3, 32), thus altering ATP production and the NADP/NADPH balance. In *Synechocystis*, salt stress has been reported to enhance PSI-dependent cyclic electron flow (58) that generates ATP but not NADPH (36), thus participating in determining the ATP/NADPH ratio when photosynthesis operates under changing environmental conditions (45). The fact that AdhA from *Synechocystis* oxidizes NADPH much more efficiently than it reduces NADP<sup>+</sup> supports a possible role for this enzyme in oxidizing NADPH under such conditions and thus contributing to the maintenance of the ATP/NADPH balance, as was also previously proposed for two members of the CADH family in *Saccharomyces* (27, 28).

We have compared the growth of an  $\Delta adhA$  mutant with that of the wild-type strain under a variety of conditions. No differences in responses to heat shock, salt, or hyperosmotic stress have been observed. Both strains exhibit analogous growth rates when tested in the light under autotrophic, mixotrophic (added glucose), or photoheterotrophic [added glucose and 3-(3P,4P-dichlorophenyl)-1,1P-dimethylurea] conditions. Nevertheless, the *adhA* mutant strain was unable to grow heterotrophically (added glucose) in darkness, contrary to the wild-type strain (our unpublished data). These results strongly suggest an involvement of AdhA in heterotrophic metabolism in *Synechocystis*, which calls for further investigation.

The histidine kinase Hik34 is involved in the regulation of the expression of heat shock genes in *Synechocystis* (53). Four two-component systems are involved in the perception and transduction of salt and hyperosmotic stress signals in *Synechocystis*, with the Hik34-Rre1 system being the one responsible for controlling the expression of the *adhA* and *sl1107* genes under conditions of salt stress (39, 48). The results obtained in the present work concerning the effect of a mutation of either *hik34* or *rre1* on the expression of the *adhA* gene show that the Hik34-Rre1 system is involved in the early induction of *adhA* expression under conditions of heat shock and hyperosmotic stress, a fact not yet reported. Although this system is essential for the response of the *adhA* gene to heat shock, the enhancement of transcription of *adhA* in response to salt stress, hyperosmotic stress, or the presence of BA is only in part dependent on the operation of the Hik34-Rre1 system. Moreover, the differences observed between *hik34* and *rre1* mutants with regard to *adhA* gene transcription in response to hyperosmotic and salt stress suggest the involvement of an additional factor in the activation of Rre1.

We have demonstrated that the Rre1 protein isolated from *E. coli* binds specifically to the promoter region of the *adhA* gene in vitro. Although in general, it is accepted that response regulators exist in two conformations, an inactive state and an active state, with phosphorylation serving to shift the equilibrium toward the active state (60), recent findings (12) support the contention of an intermediate conformation between inactive and active states (51). This is the case for the CheY response regulator, which binds to its target in the unphosphorylated form and displays a basal level of kinase-independent activity in vivo (47). The phosphoacceptor domain of Rre1 is a homolog of that of CheY. The affinity for the specific DNA sequence in the *adhA* promoter of unphosphorylated Rre1 would be sufficient for the observed binding. Nevertheless, the specific sequence for binding remains to be elucidated. The

upstream divergently transcribed *sl1106* gene, which shows the same expression pattern, could share the regulatory sequences with the *adhA* gene, as suggested by the proximity of their corresponding –35 boxes. Two repeated palindromic GTTG sequences located between the putative –35 boxes of both the *adhA* and *sl1106* promoters (Fig. 4D) might be the actual binding sites for Rre1, allowing the activation of both promoters. In silico analysis predicts that the *sl1106* gene encodes a membrane protein that highly conserved among cyanobacteria. Unfortunately, the lack of information about its function hampers the establishment of any functional relationship between the *sl1106* and AdhA proteins. On the other hand, the assigned function of the *sl1107* gene product, a type IV pilus biogenesis protein, excludes a direct functional relationship with the AdhA protein.

In conclusion, *Synechocystis* possesses a zinc-dependent alcohol dehydrogenase (AdhA), which has a preference for NADPH as a reductant and aromatic and medium-chain aliphatic aldehydes as oxidants. The corresponding gene is upregulated in response to certain stress conditions via the Hik34-Rre1 two-component system. Its specific role in the heterotrophic growth of *Synechocystis* remains to be established.

#### ACKNOWLEDGMENTS

This work was supported by a grant from the Ramón Areces Foundation (to M.G.G.), grants BFU2004-0635 and BFU2007-60300 from the Ministry of Science and Education of Spain (to F.J.F.), and funds of Junta de Andalucía (BIO131 and BIO284).

We acknowledge Iwane Suzuki for kindly providing the  $\Delta hik34$  and  $\Delta rre1$  mutant strains. We acknowledge Anna Lindahl for the critical reading of the manuscript.

#### REFERENCES

- Allakhverdiev, S. I., Y. Nishiyama, I. Suzuki, Y. Tasaka, and N. Murata. 1999. Genetic engineering of the unsaturation of fatty acids in membrane lipids alters the tolerance of *Synechocystis* to salt stress. *Proc. Natl. Acad. Sci. USA* **96**:5862–5867.
- Allakhverdiev, S. I., A. Sakamoto, Y. Nishiyama, M. Inaba, and N. Murata. 2000. Ionic and osmotic effects of NaCl-induced inactivation of photosystems I and II in *Synechococcus* sp. *Plant Physiol.* **123**:1047–1056.
- Allakhverdiev, S. I., A. Sakamoto, Y. Nishiyama, and N. Murata. 2000. Inactivation of photosystems I and II in response to osmotic stress in *Synechococcus*. Contribution of water channels. *Plant Physiol.* **122**:1201–1208.
- Anderson, S. L., and L. McIntosh. 1991. Light-activated heterotrophic growth of the cyanobacterium *Synechocystis* sp. strain PCC 6803: a blue-light-requiring process. *J. Bacteriol.* **173**:2761–2767.
- Boudet, A. M. 2007. Evolution and current status of research in phenolic compounds. *Phytochemistry* **68**:2722–2735.
- Boulton, B., V. L. Singleton, L. F. Bisson, and R. E. Kunkee. 1996. Principles and practices of winemaking, p.139–172. Chapman and Hall, New York, NY.
- Cai, Y. P., and C. P. Wolk. 1990. Use of a conditionally lethal gene in *Anabaena* sp. strain PCC 7120 to select for double recombinants and to trap insertion sequences. *J. Bacteriol.* **172**:3138–3145.
- Chang, C., and R. C. Stewart. 1998. The two-component system. Regulation of diverse signaling pathways in prokaryotes and eukaryotes. *Plant Physiol.* **117**:723–731.
- Christie, P. J., M. Hahn, and V. Walbot. 1991. Low-temperature accumulation of alcohol dehydrogenase-1 mRNA and protein activity in maize and rice seedlings. *Plant Physiol.* **95**:699–706.
- Dawes, E. A., D. W. Ribbons, and P. J. Large. 1966. The route of ethanol formation in *Zyomonas mobilis*. *Biochem. J.* **98**:795–803.
- Deng, M. D., and J. R. Coleman. 1999. Ethanol synthesis by genetic engineering in cyanobacteria. *Appl. Environ. Microbiol.* **65**:523–528.
- Dyer, C. M., and F. W. Dahlquist. 2006. Switched or not?: the structure of unphosphorylated CheY bound to the N terminus of FliM. *J. Bacteriol.* **188**:7354–7363.
- Elhai, J., and C. P. Wolk. 1988. A versatile class of positive-selection vectors based on the nonviability of palindromic-containing plasmids that allows cloning into long polylinkers. *Gene* **68**:119–138.
- Ferino, F., and F. Chauvat. 1989. A promoter-probe vector-host system for the cyanobacterium, *Synechocystis* PCC6803. *Gene* **84**:257–266.



15. Fulda, S., S. Mikkat, F. Huang, J. Huckauf, K. Marin, B. Norling, and M. Hagemann. 2006. Proteome analysis of salt stress response in the cyanobacterium *Synechocystis* sp. strain PCC 6803. *Proteomics* **6**:2733–2745.
16. Galamba, A., K. Soetaert, P. Buysens, D. Monnaie, P. Jacobs, and J. Content. 2001. Molecular and biochemical characterisation of *Mycobacterium smegmatis* alcohol dehydrogenase C. *FEMS Microbiol. Lett.* **196**: 51–56.
17. Garcia-Dominguez, M., and F. J. Florencio. 1997. Nitrogen availability and electron transport control the expression of *glnB* gene (encoding PII protein) in the cyanobacterium *Synechocystis* sp. PCC 6803. *Plant Mol. Biol.* **35**:723–734.
18. Horvath, I., A. Glatz, V. Varvasovszki, Z. Torok, T. Pali, G. Balogh, E. Kovacs, L. Nadasdi, S. Benko, F. Joo, and L. Vigh. 1998. Membrane physical state controls the signaling mechanism of the heat shock response in *Synechocystis* PCC 6803: identification of *hsp17* as a “fluidity gene.” *Proc. Natl. Acad. Sci. USA* **95**:3513–3518.
19. Inaba, M., I. Suzuki, B. Szalontai, Y. Kanesaki, D. A. Los, H. Hayashi, and N. Murata. 2003. Gene-engineered rigidification of membrane lipids enhances the cold inducibility of gene expression in *Synechocystis*. *J. Biol. Chem.* **278**:12191–12198.
20. Jarillo, J. A., A. Leyva, J. Salinas, and J. M. Martinez-Zapater. 1993. Low temperature induces the accumulation of alcohol dehydrogenase mRNA in *Arabidopsis thaliana*, a chilling-tolerant plant. *Plant Physiol.* **101**:833–837.
21. Jornvall, H., and J. O. Hoog. 1995. Nomenclature of alcohol dehydrogenases. *Alcohol Alcohol.* **30**:153–161.
22. Jornvall, H., J. O. Hoog, and B. Persson. 1999. SDR and MDR: completed genome sequences show these protein families to be large, of old origin, and of complex nature. *FEBS Lett.* **445**:261–264.
23. Jornvall, H., B. Persson, and J. Jeffery. 1987. Characteristics of alcohol/polyol dehydrogenases. The zinc-containing long-chain alcohol dehydrogenases. *Eur. J. Biochem.* **167**:195–201.
24. Kanesaki, Y., I. Suzuki, S. I. Allakhverdiev, K. Mikami, and N. Murata. 2002. Salt stress and hyperosmotic stress regulate the expression of different sets of genes in *Synechocystis* sp. PCC 6803. *Biochem. Biophys. Res. Commun.* **290**:339–348.
25. Kim, S. J., K. W. Kim, M. H. Cho, V. R. Franceschi, L. B. Davin, and N. G. Lewis. 2007. Expression of cinnamyl alcohol dehydrogenases and their putative homologues during *Arabidopsis thaliana* growth and development: lessons for database annotations? *Phytochemistry* **68**:1957–1974.
26. Kimmerer, T. W., and T. T. Kozlowski. 1982. Ethylene, ethane, acetaldehyde, and ethanol production by plants under stress. *Plant Physiol.* **69**:840–847.
27. Larray, C., M. R. Fernandez, E. Gonzalez, X. Pares, and J. A. Biosca. 2002. Characterization of the *Saccharomyces cerevisiae* YMR318C (ADH6) gene product as a broad specificity NADPH-dependent alcohol dehydrogenase: relevance in aldehyde reduction. *Biochem. J.* **361**:163–172.
28. Larray, C., X. Pares, and J. A. Biosca. 2002. Characterization of a *Saccharomyces cerevisiae* NADP(H)-dependent alcohol dehydrogenase (ADHVII), a member of the cinnamyl alcohol dehydrogenase family. *Eur. J. Biochem.* **269**:5738–5745.
29. Lauvergeat, V., K. Kennedy, C. Feuillet, J. H. McKie, L. Gorrichon, M. Baltas, A. M. Boudet, J. Grima-Pettenati, and K. T. Douglas. 1995. Site-directed mutagenesis of a serine residue in cinnamyl alcohol dehydrogenase, a plant NADPH-dependent dehydrogenase, affects the specificity for the coenzyme. *Biochemistry* **34**:12426–12434.
30. Leskovac, V., S. Trivic, and D. Pericin. 2002. The three zinc-containing alcohol dehydrogenases from baker's yeast, *Saccharomyces cerevisiae*. *FEMS Yeast Res.* **2**:481–494.
31. Lopez-Maury, L., M. Garcia-Dominguez, F. J. Florencio, and J. C. Reyes. 2002. A two-component signal transduction system involved in nickel sensing in the cyanobacterium *Synechocystis* sp. PCC 6803. *Mol. Microbiol.* **43**:247–256.
32. Marin, K., Y. Kanesaki, D. A. Los, N. Murata, I. Suzuki, and M. Hagemann. 2004. Gene expression profiling reflects physiological processes in salt acclimation of *Synechocystis* sp. strain PCC 6803. *Plant Physiol.* **136**:3290–3300.
33. Marin, K., I. Suzuki, K. Yamaguchi, K. Ribbeck, H. Yamamoto, Y. Kanesaki, M. Hagemann, and N. Murata. 2003. Identification of histidine kinases that act as sensors in the perception of salt stress in *Synechocystis* sp. PCC 6803. *Proc. Natl. Acad. Sci. USA* **100**:9061–9066.
34. McKie, J. H., R. Jaouhari, K. T. Douglas, D. Goffner, C. Feuillet, J. Grima-Pettenati, A. M. Boudet, M. Baltas, and L. Gorrichon. 1993. A molecular model for cinnamyl alcohol dehydrogenase, a plant aromatic alcohol dehydrogenase involved in lignification. *Biochim. Biophys. Acta* **1202**:61–69.
35. Mikami, K., Y. Kanesaki, I. Suzuki, and N. Murata. 2002. The histidine kinase Hik33 perceives osmotic stress and cold stress in *Synechocystis* sp. PCC 6803. *Mol. Microbiol.* **46**:905–915.
36. Munekage, Y., M. Hashimoto, C. Miyake, K. Tomizawa, T. Endo, M. Tasaka, and T. Shikanai. 2004. Cyclic electron flow around photosystem I is essential for photosynthesis. *Nature* **429**:579–582.
37. Murata, N., and I. Suzuki. 2006. Exploitation of genomic sequences in a systematic analysis to access how cyanobacteria sense environmental stress. *J. Exp. Bot.* **57**:235–247.
38. Nordling, E., H. Jornvall, and B. Persson. 2002. Medium-chain dehydrogenases/reductases (MDR). Family characterizations including genome comparisons and active site modeling. *Eur. J. Biochem.* **269**:4267–4276.
39. Paithoonrangarid, K., M. A. Shoumskaya, Y. Kanesaki, S. Satoh, S. Tabata, D. A. Los, V. V. Zinchenko, H. Hayashi, M. Tanticharoen, I. Suzuki, and N. Murata. 2004. Five histidine kinases perceive osmotic stress and regulate distinct sets of genes in *Synechocystis*. *J. Biol. Chem.* **279**:53078–53086.
40. Perez-Perez, M. E., F. J. Florencio, and M. Lindahl. 2006. Selecting thioredoxins for disulphide proteomics: target proteomes of three thioredoxins from the cyanobacterium *Synechocystis* sp. PCC 6803. *Proteomics* **6**(Suppl. 1):S186–S195.
41. Persson, B., J. S. Zigler, Jr., and H. Jornvall. 1994. A super-family of medium-chain dehydrogenases/reductases (MDR) sub-lines including  $\zeta$ -crystallin, alcohol and polyol dehydrogenases, quinone oxidoreductases, enoyl reductases, VAT-1 and other proteins. *Eur. J. Biochem.* **226**:15–22.
42. Rippka, R., J. Deruelles, J. B. Waterbury, M. Herman, and R. Y. Stanier. 1979. Genetics assignments, strain histories and properties of pure cultures of cyanobacteria. *J. Gen. Microbiol.* **111**:1–61.
43. Rittmann, B. E., R. Krajmalnik-Brown, and R. U. Halden. 2008. Pre-genomic, genomic and post-genomic study of microbial communities involved in bioenergy. *Nat. Rev. Microbiol.* **6**:604–612.
44. Riveros-Rosas, H., A. Julian-Sanchez, R. Villalobos-Molina, J. P. Pardo, and E. Pina. 2003. Diversity, taxonomy and evolution of medium-chain dehydrogenase/reductase superfamily. *Eur. J. Biochem.* **270**:3309–3334.
45. Rumeau, D., G. Peltier, and L. Cournac. 2007. Chlororespiration and cyclic electron flow around PSI during photosynthesis and plant stress response. *Plant Cell Environ.* **30**:1041–1051.
46. Sambrook, J., E. F. Fritsch, and T. Maniatis. 1989. *Molecular cloning: a laboratory manual*, 2nd ed. Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY.
47. Schuster, M., R. E. Silversmith, and R. B. Bourret. 2001. Conformational coupling in the chemotaxis response regulator CheY. *Proc. Natl. Acad. Sci. USA* **98**:6003–6008.
48. Shoumskaya, M. A., K. Paithoonrangarid, Y. Kanesaki, D. A. Los, V. V. Zinchenko, M. Tanticharoen, I. Suzuki, and N. Murata. 2005. Identical Hik-Rre systems are involved in perception and transduction of salt signals and hyperosmotic signals but regulate the expression of individual genes to different extents in *Synechocystis*. *J. Biol. Chem.* **280**:21531–21538.
49. Skulachev, V. P. 1994. Bioenergetics: the evolution of molecular mechanisms and the development of bioenergetic concepts. *Antonie van Leeuwenhoek* **65**:271–284.
50. Stal, L. J., and R. Moezelaar. 1997. Fermentation in cyanobacteria. *FEMS Microbiol. Rev.* **21**:179–211.
51. Stock, A. M., and J. Guhaniyogi. 2006. A new perspective on response regulator activation. *J. Bacteriol.* **188**:7328–7330.
52. Sun, H. W., and B. V. Plapp. 1992. Progressive sequence alignment and molecular evolution of the Zn-containing alcohol dehydrogenase family. *J. Mol. Evol.* **34**:522–535.
53. Suzuki, I., Y. Kanesaki, H. Hayashi, J. J. Hall, W. J. Simon, A. R. Slabas, and N. Murata. 2005. The histidine kinase Hik34 is involved in thermotolerance by regulating the expression of heat shock genes in *Synechocystis*. *Plant Physiol.* **138**:1409–1421.
54. Suzuki, I., Y. Kanesaki, K. Mikami, M. Kanehisa, and N. Murata. 2001. Cold-regulated genes under control of the cold sensor Hik33 in *Synechocystis*. *Mol. Microbiol.* **40**:235–244.
55. Suzuki, I., D. A. Los, Y. Kanesaki, K. Mikami, and N. Murata. 2000. The pathway for perception and transduction of low-temperature signals in *Synechocystis*. *EMBO J.* **19**:1327–1334.
56. Tadege, M., I. I. Dupuis, and C. Kuhlemeier. 1999. Ethanol fermentation: new functions for an old pathway. *Trends Plant Sci.* **4**:320–325.
57. Tani, A., Y. Sakai, T. Ishige, and N. Kato. 2000. Thermostable NADP<sup>+</sup>-dependent medium-chain alcohol dehydrogenase from *Acinetobacter* sp. strain M-1: purification and characterization and gene expression in *Escherichia coli*. *Appl. Environ. Microbiol.* **66**:5231–5235.
58. van Thor, J. J., R. Jeanjean, M. Havaux, K. A. Sjollem, F. Joset, K. J. Hellingwerf, and H. C. Matthijs. 2000. Salt shock-inducible photosystem I cyclic electron transfer in *Synechocystis* PCC6803 relies on binding of ferredoxin:NADP(+) reductase to the thylakoid membranes via its CpcD phyco-bilisome-linker homologous N-terminal domain. *Biochim. Biophys. Acta* **1457**:129–144.
59. Vioque, A. 1992. Analysis of the gene encoding the RNA subunit of ribonucleic acid P from cyanobacteria. *Nucleic Acids Res.* **20**:6331–6337.
60. Volkman, B. F., D. Lipson, D. E. Wemmer, and D. Kern. 2001. Two-state allosteric behavior in a single-domain signaling protein. *Science* **291**:2429–2433.
61. West, A. H., and A. M. Stock. 2001. Histidine kinases and response regulator proteins in two-component signaling systems. *Trends Biochem. Sci.* **26**:369–376.
62. Wilkin, J. M., K. Soetaert, M. Stelandre, P. Buysens, G. Castillo, V. Demoulin, G. Botu, M. A. Laneelle, M. Daffe, and J. De Bruyn. 1999. Overexpression, purification and characterization of *Mycobacterium bovis* BCG alcohol dehydrogenase. *Eur. J. Biochem.* **262**:299–307.