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Corrigendum

Corrigendum to "*Larsenia salina* gen. nov., sp. nov., a new member of the family *Halomonadaceae* based on multilocus sequence analysis" [Syst. Appl. Microbiol. 37 (October (7)) (2014) 480–487]

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In a paper accepted for publication in Syst. Appl. Microbiol., we proposed a new genus and species of the family *Halomonadaceae*, with the new name *Larsenia salina*. However, the name *Larsenia* already exists in the botanical and in the zoological nomenclature and thus, the proposed name *Larsenia* is illegitimate according to Principle 2 of the International Code of Nomenclature of Prokaryotes. For this reason, we propose the correction of the new name *Larsenia salina* to *Larsenimonas salina* gen. nov., sp. nov. and enclose a correction of the proposed genus and species and their complete protologues.

Description of *Larsenimonas* gen. nov.

Larsenimonas (Lar.se.ni.mo'nas, Gr. fem. n. *monas*, unit, monad; N.L. fem. n. *Larsenimonas*, a monad named after Helge Larsen a pioneer of the study of halophilic microorganisms).

Cells are Gram-staining-negative, motile rods. Endospores are not formed. Organotrophic. Strictly aerobic, catalase and oxidase positive. Moderately halophilic. Major fatty acids are $C_{18:1}\omega7c/C_{18:1}\omega6c$, $C_{16:0}$, and $C_{16:1}\omega7c/C_{16:1}\omega6c$. The only respiratory quinone is Q-9 and the major polar lipids are phosphatidylglycerol, phosphatidylethanolamine, phospholipids and glycolipids. The genus *Larsenimonas* belongs to the family *Halomonadaceae* within the class *Gammaproteobacteria*. The type species is *Larsenimonas salina*.

Description of *Larsenimonas salina* sp. nov.

Larsenimonas salina (sa.li'na. L. fem. adj. *salina*, salted, saline).

Cells are Gram-staining-negative, motile, straight rods, 0.4–0.8 × 0.8–2.1 μm in size. Colonies are circular, entire, smooth, convex, yellow pigmented and 0.7–3.0 mm in diameter on 7.5% SW agar medium after 48 h incubation at 37 °C. Strictly aerobic. Moderately halophilic, growing at 3–25% (w/v) NaCl; with optimal growth at 7.5–10% (w/v) NaCl. No growth occurs in the absence of NaCl. Grows at 15–40 °C; showing optimal growth at 37 °C, and at pH values on the range 4.0–9.0; with optimal growth at pH 7.0. Anaerobic growth on nitrate or arginine negative. Catalase and oxidase positive. Gelatin, DNA, Tween 80 and aesculin are hydrolysed but starch is not hydrolysed. Nitrate is not reduced to nitrite. Acid is produced from D-glucose, D-arabinose, sucrose and D-trehalose but not from D-mannitol, D-amygdaline, L-citruline, DL-ethionine, inuline, lactose, melezitose, D-ribose, raffinose, sorbitol or xylitol. Indole or H_2S are not produced. Phosphatase is positive. Simmons' citrate is variable. Urease, arginine and phenylalanine deaminase tests are negative. The following compounds are utilized as sole sources of carbon and energy: D-galactose, D-glucose, D-ribose, glycerol, salicine, myo-inositol, benzoate, fumarate, hippurate and citrate. The following compounds are not utilized as sole sources of carbon and energy: D-fucose, aesculin, starch, butanol, dulcitol, methanol, formate, malate, propionate and tartrate. The following compounds are utilized as sole sources of carbon, nitrogen and energy: L-isoleucine, L-methionine and L-valine. The following compounds are not utilized as sole sources of carbon, nitrogen and energy: L-arginine, aspartate, L-threonine and tryptophan. The predominant cellular fatty acids are $C_{18:1}\omega7c/C_{18:1}\omega6c$, $C_{16:0}$, and $C_{16:1}\omega7c/C_{16:1}\omega6c$. The DNA G + C content is 54.5–55.9 mol% (T_m).

The type strain is M1-18^T (=CCM 8464^T = CECT 8192^T = IBRC-M 10767^T = LMG 27461^T). The DNA G + C content of the type strain is 54.5 mol% (T_m). This strain is unable to hydrolyze casein and is Simmons' citrate negative. Able to utilize L-cysteine and ethanol as sole carbon and energy source.

The authors would like to apologise for any inconvenience caused.

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