1 . 1 01 Pofomonoso E 12 (Journal of Antimicrobial Chemotherapy doi:10.1093/jac/dki488 Advance Access publication 23 January 2006 9 Characterization of a clinical isolate of Haemophilus influenzae with a high level of fluoroquinolone I resistance J. M. Rodríguez-Martínez^{1*}, L. López², I. García¹ and C A. Pascual^{1,2} 0 d e ¹School of Medicine, University of Seville, Seville, Spain; tl ²Hospital Virgen Macarena, Seville, Spain T D V e S Keywords: antibiotic resistance, susceptibility, ciprofloxacin il d p *Corresponding author. Tel: +34-954552863; Fax: +34c 954177413; E-mail: jmrodriguez@us.es Y il h ti f a b e Sir V Haemophilus influenzae is one of the bacterial pathogens most b likely to be involved in community-acquired respiratory infections tı in patients with chronic obstructive pulmonary disease. In this p species, resistance to fluoroquinolones occurs at very low fre-C quency although in such cases treatment failure has been reported.1 fi In 2003, a fluoroquinolone-resistant H. influenzae isolate was S recovered from the sputum of an 80-year-old male patient with a history of bronchiectasis and residual lesions from old tuber-7 culosis but with no history of treatment with fluoroquinolones. iı Identification of this H. influenzae (strain number 79105) was S carried out by Gram stain, tests for catalase, oxidase, X and V iı factor requirements (MAST), Api NH (bioMerieux, Madrid, Spain) and antisera to serotypes a-f (Difco, Sparks, MD, USA). Susceptibility tests were performed by disc diffusion and Etest V on HTM agar and Chocolate agar at 35°C and 5% CO2 and by microdilution in HTM broth according to CLSI (formerly NCCLS) guidelines.2 H. influenzae ATCC 49247 was used as a reference strain. Isolate 79105 was resistant to nalidixic acid, fluoroquino-7

lones and co-trimoxazole and showed MICs of >256, 32, 32, 8 and 2 mg/L for nalidixic acid, ciprofloxacin, norfloxacin, levofloxacin

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and moxifloxacin, respectively, and >4/76 mg/L for co-trimoxazole. Moreover, this strain was non-typeable and a non- β -lactamase producer like most of the previously described fluoroquinolone-resistant isolates.

Interestingly, this clinical strain showed dependence with nalidixic acid, ciprofloxacin and co-trimoxazole on HTM agar, growing only round the disc or Etest strip containing these antibiotics. In contrast this effect was not seen with organisms grown on Chocolate agar. Surprisingly, the dependence disappeared after several subcultures.

In our hospital the proportion of isolates of H. influenzae resistant to new quinolones is <1% and even lower rates have been reported in other studies.^{3,4} At the present time only a few isolates of H. influenzae with a high level of fluoroquinolone resistance (MIC of ciprofloxacin >16 mg/L) have been described.⁵ Since mutations in the DNA gyrase and topoisomerase IV genes are the most common mechanisms of fluoroquinolone resistance, the corresponding regions of the gyrA and parC genes of H. influenzae ATCC 49247 (as control) and of H. influenzae 79105 strains were sequenced (the primers used have been described previously).1 The sequence of the quinolone-resistant isolate showed two substitutions in GyrA (S84Y and D88A) and two in ParC (S84N and E88A). This combination of mutations has not been described previously and only one H. influenzae isolate has previously shown four substitutions in these genes.⁵ Additionally, the QRDR regions of the gyrB and parE genes were analysed with the set of previously described primers.1 The sequence of the quinolone-resistant isolate showed the E559G substitution in GyrB and the L445F substitution in ParE. In summary, this strain contained six mutations in four different genes associated with quinolone resistance. In all cases, the sequence of the ATCC 49247 H. influenzae strain was identical to those obtained from

To explain observed differences in susceptibility between ciprofloxacin and moxifloxacin, we tried to identify the presence of a possible efflux pump with a different affinity for those antimicrobials. Efflux of fluoroquinolones was determined indirectly by measuring the intracellular accumulation of norfloxacin by fluorometric assay. A standardized cell suspension (OD₅₂₀ = 2.0) was preincubated with 10 mg/L norfloxacin, following which half of the cell suspension was exposed to 0.2 mM carbonyl cyanide chlorophenylhydrazone (CCCP). Cell-fluorescence was measured with a fluorescence spectrophotometer (Hitachi 2000; at 279/445 nm). The addition of CCCP to the cells did not increase the level of intracellular norfloxacin. Thus, there was no evidence that efflux pumps inhibited by CCCP contributed to the observed differences in susceptibility of the strain to ciprofloxacin, norfloxacin and moxifloxacin.

In conclusion, an *H. influenzae* isolate resistant to fluoroquinolones by means of a new combination of mutations in topoisomerase genes has been described. It is unknown whether other different efflux pumps or additional mechanisms are implicated in quinolone resistance and explain the different susceptibility to ciprofloxacin and moxifloxacin in this strain.

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Transparency declarations

None to declare.

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