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1 **Effect of technological practices on individual betalains and antioxidant activity of**  
2 **Columbian betalain-rich raw materials**

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21  
22 ***RUNNING TITLE***

23 Technology on betalain and biological activity

24 ***KEYWORDS***

25 *Ullucus tuberosus*, *Opuntia dillenii*, betalains, antioxidant activity.

## 26 SUMMARY

27 The effect of different technological practices on ulluco (*ullucus tuberosus*) and  
28 *Opuntia dillenii* has been scrutinized. Their stability at different pHs and temperatures  
29 of storage or processing over time was monitored. Our attention was focused on the  
30 determination of individual betalains and antioxidant activity, not previously conducted  
31 in conjunction in these raw materials. On the basis of the results, it could be asserted  
32 that the *ullucus tuberosus* and *Opuntia dillenii* were more suitable for being added to  
33 low-acidic foods (pH 5 and 6), in the light of the higher values of betalains and  
34 antioxidant capacity (1.3-fold higher compared to pH 4). With regard to the  
35 temperature, cold storage conditions (4 °C) were optimal to increase or maintain as  
36 possible the initial content of betalains and antioxidant capacity. After cooking (80 °C),  
37 the identified betalains completely disappeared, but the low-acidic conditions (pH 6)  
38 favoured the greater antioxidant activity when that temperature was underwent.

## 39 INTRODUCTION

40 Nowadays, consumers are becoming more conscious about the importance of the  
41 consumption of natural foods, because of their many beneficial properties (Benavente-  
42 García *et al.*, 1997). There are several species rich in pigments (anthocyanins,  
43 chlorophylls and carotenoids) responsible for valuable healthy properties, but there is a  
44 group of phenolic coloured compounds (Cai *et al.*, 2003, Stintzing *et al.*, 2005) still  
45 poorly investigated, the so-called betalains.

46 Betalains are water-soluble compounds presented in a restricted number of families of  
47 the plant order Caryophyllales and of the genus *Amarita* of the Basidiomycetes  
48 (Waterman 2007). Betalamic acid is the common moiety structure, and the typology of  
49 betalains stems from the nature of the additional residues: red/purple betacyanins  
50 (condensation of betalamic acid and *cyclo*-Dopa, considering hydroxycinnamic acid

51 derivatives or sugars as residue) and yellow betaxanthins (imino condensation products  
52 between betalamic acid and amines or amino acids residues) (Herbach *et al.*, 2006)  
53 (Figure 1).

54 Betalains are widely distributed in a great variety of products, being red beet (*Beta*  
55 *vulgaris* L.) the most studied source of betalains par excellence (De Azeredo *et al.*,  
56 2009, Nemzer *et al.*, 2011). However, betalains have been also analysed in other  
57 sources, such as fruits of cactus pear (*Opuntia* sp.) or yellow beet (Stintzing *et al.*,  
58 2002a, Stintzing *et al.*, 2002b, Castellanos-Santiago and Yahia 2008, Cejudo-Bastante  
59 *et al.*, 2014a), flowers (Kugler *et al.*, 2007) and vegetables (Kugler *et al.*, 2004).

60 From a biological point of view, betalain-rich foods (red beet, *amaranthus*, cactus pear,  
61 among others) have an important free radical-scavenging activity (Escribano 1998;  
62 Kanner *et al.*, 2001; Butera *et al.*, 2002; Cai *et al.*, 2003; Dantas *et al.*, 2015).

63 Concretely, red beet and red pitaya have been established as two of the most powerful  
64 vegetables with respect to their antioxidant capacity (Vinson *et al.*, 1998, Vaillant *et al.*,  
65 2005), directly related with the presence and content of betacyanins. However, up to the  
66 present, very scarce studies about antioxidant activity of other betalain-rich products  
67 have been reported. In that sense, Campos *et al.* (2006) measured the antioxidant  
68 capacity and total betaxanthins of several Peruvian ulluco tubers and Ghazi *et al.* (2015)  
69 and Kumar *et al.* (2014) obtained the antioxidant activity of *Opuntia dillenii* extracts  
70 from Marocco and India, respectively. However, any scientific study about the  
71 relationship of individual betalains and antioxidant activity on these raw materials.

72 Therefore, this work was performed on Colombian ulluco (*ullucus tuberosus*) and  
73 cactus pear (*Opuntia dillenii*) and the interest was focused on the individual betalain  
74 characterization and antioxidant activity, not previously conducted in conjunction in  
75 those raw materials. Furthermore, not only a betalain characterization is needed, but

76 also even to spotlight the behaviour in terms of betalains and antioxidant activity when  
77 these raw materials underwent different commercial technological practices and are  
78 added as natural colorant to foodstuffs.

## 79 MATERIAL AND METHODS

### 80 Samples

81 Magenta ulluco samples, commonly known as Ulluco “Chincheño”, and red prickly  
82 pear samples of *Opuntia dillenii* were collected in the village of Catambuco and  
83 Remolinos, respectively, located in Nariño (Colombia). Both areas are located at  
84 approximately 8 km south of the city of San Juan de Pasto (Nariño, Colombia) and at  
85 2820 m above sea level, with an average temperature at around 12 °C. Ulluco is a tuber  
86 with a white pulp and a thin surface portion of magenta skin. *Opuntia dillenii* has a red-  
87 purple skin with cladode spines, and a red-purple pulp with several prickles.

88 Samples were carefully washed. Subsequently, *ullucus tuberosus* were manually peeled  
89 to separate the white pulp from the magenta peel, and spines and prickles of *Opuntia*  
90 *dillenii* were manually removed. The peelings of ulluco and the peel and pulp of  
91 *Opuntia dillenii* were cut into small pieces (1 cm<sup>2</sup>) and 1 L of methanol of  
92 methanol:water (60:40) was used for extraction for 24 h at 10 °C (maceration),  
93 according to the method proposed by Cejudo-Bastante *et al.* (2014b). After filtration,  
94 the organic solvent was evaporated at 35 °C using a rotary evaporator (Heidolph,  
95 Schwabach, Germany) and the re-dissolved with distilled water was lyophilized  
96 (Labconco, MO, USA) and stored at 4 °C until their analysis.

### 97 Preparation of extracts

98 Lyophilized samples (3 g) were added to 9.5 mL of a mixture of methanol:water  
99 (50:50), containing 50 mM of sodium ascorbate in order to avoid possible oxidations.

100 Subsequently, samples were centrifugated at 12000 x g at 10 °C for 5 min, according the

101 method described by Cejudo-Bastante *et al.* (2014b). Supernatants were recovered and  
102 the procedure was developed once more with the extraction solution and finally with  
103 100 % methanol in order to a complete discoloration of the plant tissue. Later, the  
104 extract obtained was concentrated in vacuum (30 °C) and re-suspended until 15 mL with  
105 purified water. All experiences were carried out in duplicate.

#### 106 **Technological treatments of the extracts**

107 Extracts were adjusted to different acidities (pH 4, 5 and 6) using hydrochloride acid 0.1  
108 and 1 M. Subsequently, each pH-sample were submitted to three different temperatures:  
109 4 °C, and 20° and 80 °C, putting samples into a refrigerator and an oven JP Selecta, S.A.  
110 (Barcelona, Spain), respectively. Samples were analysed at the beginning of the  
111 treatment ( $t_0$ ) and after a period of storage ( $t_r$ ), which was 12 days when samples were  
112 kept at 4 °C and 20 °C, and 1 day when they underwent 80 °C.

#### 113 **HPLC-DAD analysis**

114 The Agilent 1200 chromatographic system equipped with a quaternary pump, an UV-  
115 vis diode-array detector, an automatic injector, and ChemStation software (Palo Alto,  
116 CA, USA) were used to the HPLC separation, identification and quantification of  
117 betalains. Prior direct injection, the samples were filtered through a 0.45 µm nylon filter  
118 (E0034, Análisis Vínicos, Spain). All analyses were made in duplicate.

119 Betalains were separated in a Zorbax C18 column (250 x 4.6 mm, 5 µm particle size)  
120 maintained at 25 °C at a flow rate of 1 mL/min. The eluents were 1% formic acid in  
121 water (v/v, eluent A) and a mixture 80:20 of acetonitrile:water (eluent B). The injection  
122 of the volume for all extracts was 20 µL. The elution profile of betalains was different  
123 depending on the sample: ulluco, 0 min, 5% B; 10 min, 10% B; 20 min 10% B; 25 min  
124 20% B; 30 min 30% B; 35 min 100% B; 37 min 100% B; *Opuntia dillenii*, 0 min, 5%  
125 B; 10 min, 12% B; 20 min 14% B; 25 min 20% B; 30 min 30% B; 35 min 100% B; 37

126 min 100% B. In order to re-establish the initial conditions, a linear gradient from 100%  
127 B to 5% B was used during 2 min, and maintained for 2 min more. For all samples,  
128 UV–Vis spectra were recorded from 200 to 800 nm with a bandwidth of 1.0 nm.  
129 Betacyanins and betaxanthins were monitored at 535 and 482 nm, respectively. The  
130 identification of each chromatographic peak were tentative assigned by their visible  
131 spectral characteristics **in comparison with retention times**. Semi-quantification was  
132 done on the basis of the mean areas of individual betalains.

### 133 **Antioxidant activity**

134 The antioxidant capacity was measured *in vitro* on the basis of the ability to scavenge  
135 the ABTS<sup>•+</sup> radical (Jara-Palacios *et al.*, 2014). The ABTS<sup>•+</sup> radical was produced by  
136 the oxidation of 7 mM ABTS with potassium persulfate (2.45 mM) in water. After  
137 storage under dark conditions and room temperature for 16 h before its use, ABTS<sup>•+</sup>  
138 solution was diluted with phosphate-buffered saline (PBS) at pH 7.4, until reaching an  
139 absorbance of  $0.70 \pm 0.02$  at 734 nm. Then, 50  $\mu$ L of each sample was mixed with 2 mL  
140 of the ABTS<sup>•+</sup> diluted solution and vortexed for 10 s, and the absorbance was measured  
141 at 734 nm after 3 min of reaction at 30 °C. The results were achieved with the help of a  
142 calibration curve made with Trolox (30-1000  $\mu$ M). Three replicates were performed for  
143 each sample, and the results were expressed as Trolox equivalent antioxidant activity  
144 (TEAC;  $\mu$ mol of Trolox equivalent (TE) with the same **antioxidant capacity** of 1 L of  
145 sample).

### 146 **Statistical analysis**

147 The complete set of analytical data was submitted to analysis of variance (ANOVA),  
148 Principal Component Analysis (PCA) and cluster analysis (CA) using the statistical  
149 computer packages Statistica version 8.0 software (Statistica 2007).

## 150 **RESULTS AND DISCUSSION**

## 151 **Betalain profile**

152 In this research, several betalain compounds were identified and semi-quantified in both  
153 raw materials: betanin and isobetanin ( $R_1$ , H;  $R_2$ ,  $\beta$ -glucoside), as betacyanins, and  
154 indicaxanthin ( $R_3$ , proline), as betaxanthins (Fig. 1). The mean areas are exposed in  
155 Table 1. Moreover, Table S1 show the ANOVA analysis on the basis of the three  
156 factors studied: pH, temperature and time.

157 At the beginning of the treatment ( $t_0$ ), the content of betanin was significantly  
158 (ANOVA;  $p < 0.05$ ) higher than of the isobetanin in ulluco samples for all pHs (Table  
159 1), whereas the ratio betanin/isobetanin was nearly 1 in *Opuntia dillenii* samples.  
160 Similar results were observed by Svenson *et al.* (2008) in red ulluco samples, but any  
161 reference about individual betalains of *Opuntia dillenii* has been found in bibliography.

162 In order to statistically highlight the differences on the betalain content among *ullucus*  
163 *tuberosus* and *Opuntia dillenii*, the complete set of data were submitted to ANOVA,  
164 considering the effect of three factors: temperature, pH and time (Table S1). Cooked  
165 samples (80 °C) are not included in the statistical analysis due to the obvious significant  
166 differences in comparison with the rest of temperatures. Results revealed that significant  
167 ( $p < 0.05$ ) differences according “temperature” and “time” factors were observed in  
168 both raw materials, whereas “pH” did not displayed remarkable differences.

169 In an attempt to deepen on the effect of the three factors on individual betalains, a more  
170 detailed statistical analysis have been developed. Thus, at the beginning of the treatment  
171 ( $t_0$ ), the significant (ANOVA;  $p < 0.05$ ) highest content of betalains was found at low  
172 pH-values (pH 4) in *Opuntia dillenii*, although without significant differences among  
173 pHs in *ullucus tuberosus* (Table 1). The storage made the content of betalains  
174 significantly (ANOVA;  $p < 0.05$ ) decrease in both ulluco and *Opuntia dillenii* samples,  
175 although the behaviour over time was greatly influenced by the temperature.



176 Thus, under refrigeration conditions, the highest values of betacyanins of ulluco  
177 correspond to pH 5 (Table 1), with a percentage of diminution of betanin around 10 %  
178 and negligible of isobetanin. The rest of pHs (pH 4 and 6) suffered an important  
179 decrease (around 40 and 25%, respectively). In the case of *Opuntia dillenii*, the highest  
180 values of betacyanins and betaxanthins (and a consequence the best conditions of  
181 acidity at 4 °C) corresponded to low-acidic samples (pH 6), with the lowest percentage  
182 of diminution over time. It is also highlighted that the percentage of diminution of  
183 indicaxanthin at cold storage was lessened as pH increased.

184 The panorama changed when samples were kept under room temperature conditions (20  
185 °C). The content of betalains decreased as pH increased in ulluco samples, and the  
186 highest values of betalains (and, subsequently, the best conditions of conservation) were  
187 found when they were adjusted to pH 4. Contrary results exhibited *Opuntia dillenii*  
188 samples, which the highest values of betalain content correspond to samples adjusted to  
189 pH 6. Same value of pH were ascertained at maximum HPLC area in several species of  
190 *Opuntia* (Castellar *et al.*, 2003).

191 Besides, it is highlighted that the behaviour of the samples submitted to 80 °C greatly  
192 differed from the rest of temperatures, and the individual betalains completely  
193 disappeared in both kind of samples, maybe for the formation of derivative compounds  
194 as a consequence of the thermal process (Stintzing and Carle 2007).

195 In brief, if *Opuntia dillenii* was added to foods with pH value over 4 (for example,  
196 vegetables), higher quantities might be added to foodstuffs to counteract its lower  
197 betalain content. These results were in concordance with those obtained by Cejudo-  
198 Bastante *et al.* (2015) in a colorimetric study carried out on *Opuntia dillenii*,  
199 demonstrating the lower chromaticity provided at that pH value. On that matter, ulluco  
200 could be suitable for being added to acid or basic foodstuffs (vegetables or meats)

201 because it provides similar betalain contents. In order to preserve as possible the content  
202 of betalains in both raw materials, the best temperature of conservation were under  
203 refrigeration conditions. However, the choice of the best acidity conditions was  
204 dependent of the raw material, pH 5 for ulluco and pH 6 for *Opuntia dillenii*. These  
205 results complement those obtained by Cejudo-Bastante *et al.* (2014b) who affirmed that  
206 *ullucus tuberosus* could be more suitable for being added to low-acidity foodstuffs  
207 (meat industry) because it provides higher values of chroma, but assuming that possible  
208 yellow tonalities could confer over time.

### 209 **Antioxidant activity**

210 Table 1 show the values of **antioxidant capacity** of both raw materials (*Ullucus*  
211 *tuberosus* and *Opuntia dillenii*), and Table S1 the results of the ANOVA analysis on the  
212 basis of “pH”, “temperature” and “time” **factors**.

213 **As it can be seen, *ullucus tuberosus* manifested a significantly higher capacity in**  
214 **comparison with *Opuntia dillenii*, and regardless acidity conditions. Even so, both**  
215 ***Opuntia dillenii* and ulluco extracts were important sources of antioxidant activity, as**  
216 **other authors affirmed (Kumar *et al.*, 2014; Ghazi *et al.*, 2015).**

217 When ANOVA was applied to understand if significant differences occurred in  
218 antioxidant capacity according temperature, pH and time for all set of data, it is  
219 highlighted that only pH was significant (**ANOVA;  $p < 0.05$** ) in both samples (Table  
220 S1), with **remarkably** higher values of antioxidant activity in low-acidic samples (Table  
221 1). **Escribano *et al.* (1998) also demonstrated that antiradical activity was strongly**  
222 **influenced by the pH of *Beta vulgaris* roots.** Also “time” was a significant (**ANOVA;  $p$**   
223  **$< 0.05$** ) factor in *Opuntia dillenii*, mainly due the behaviour of the low-acidic samples  
224 (pH 6).

225 At the beginning of the treatment ( $t_0$ ), although the effect of pH on antioxidant capacity  
226 was similar for both ulluco and *Opuntia dillenii* (i.e., significant (ANOVA;  $p < 0.05$ )  
227 higher values in samples adjusted to pH 6), the behaviour after storage greatly depended  
228 on each raw material (Table 1).

229 In general, antioxidant capacity experimented a significant (ANOVA;  $p < 0.05$ )  
230 attenuation after storage. Although highly-acidic (pH 4) extracts of *ullucus tuberosus*  
231 and *Opuntia dillenii* (pH 4) showed scarce variations of antioxidant activity regardless  
232 of temperature, they were the samples with the lowest values of antioxidant capacity.  
233 That fact was in concordance with the behaviour of the content of individual betalains,  
234 which showed a significantly (ANOVA;  $p < 0.05$ ) lower amount under those acidity  
235 conditions. With regard to the intermediate- (pH 5) and low-acidic (pH 6) extracts,  
236 temperature played an important role in the evolution of the antioxidant capacity. Thus,  
237 when pH 5-extracts were submitted to cold conditions (4 °C), a remarkable decreased of  
238 antioxidant potential was observed, above all in ulluco extracts (around 40 %). That fact  
239 demonstrated that it could be more appropriate to conserve these samples at room  
240 temperature (20 °C) or be cooked (80 °C).

241 In ulluco, the values of antioxidant activity significantly decreased when extracts was  
242 submitted to room temperature and cooked, but significantly ( $p < 0.05$ ) increased when  
243 cold storage was experimented. However, in *Opuntia dillenii* samples, 4 °C and 80 °C  
244 made antioxidant capacity increase, keeping constant when room temperature was  
245 underwent. It is remarkable that any detailed study concerning antioxidant activity of  
246 these raw materials have been previously performed up.

247 In any case, if *ullucus tuberosus* and *Opuntia dillenii* were added to high-acidic  
248 foodstuffs (pH 4), it would provide significant lower antioxidant capacity, regardless of  
249 the temperature of storage. These raw materials could be also used in foods with

250 intermediate acidity (pH 5), although cold conditions made antioxidant activity  
251 decrease. Adding *ullucus tuberosus* to pH 5-foodstuffs and *Opuntia dillenii* to pH-6  
252 foods could better withstand the cooking temperatures (80 °C) because the highest  
253 values of antioxidant capacity were reached. That fact could be due to the formation of  
254 other compounds, depending on each raw material, that could provide biological  
255 function (Stintzing and Carle 2007). In any case, the best option, from an antioxidant  
256 capacity point of view, corresponded to the addition of both *Ullucus tuberosus* and  
257 *Opuntia dillenii* to low-acidic foodstuff, keeping them under cold storage.

### 258 **Multivariate analysis**

259 Principal Component Analysis (PCA), carried out on individual betalains and  
260 antioxidant capacity, was applied with the aim of establishing which of the variables  
261 were better for discriminating among samples, and carrying out an exploratory tool to  
262 uncover unknown trends in the data. **Samples at the three different pHs and**  
263 **temperatures at the beginning and after 12 days of storage were considered.** When the  
264 set of data was subjected to PCA, two main significant factors PCs arose according to  
265 Kaiser's criterion (eigenvalues > 1). Figure 2a and b show the score plot of the first two  
266 PCs for each fruit, which explained 97.33% and 99.08% of the total variance of *Opuntia*  
267 *dillenii* and *ullucus tuberosus*, respectively. As can be seen in Figure 2, **PC 1 organized**  
268 **samples according to the temperature in** three different groups. *Opuntia dillenii* samples  
269 were classified at the beginning of the treatment ( $t_0$ ), samples stored at 4 °C, and those  
270 kept at 20 and 80 °C (Fig. 2a), whereas ulluco (Fig. 2b) were **situated together those** at  
271 the beginning of the treatment and stored at 4 °C, coming close to those kept at 20 °C  
272 and finally 80 °C. It could be seen that, when temperature was increased (20 and 80 °C),  
273 ulluco samples adjusted to pH 5 tended to separate from the rest (Fig 2b). **However,**  
274 **scarce grouping according to pH were observed for *Opuntia dillenii*. In any case, the**

275 content of all individual betalains identified (betanin, isobetanin and indicaxanthin)  
276 decreased with temperature (PC 1). PC2 separated *Opuntia dillenii* samples adjusted to  
277 pH 6 and stored at 4 °C and 80 °C (Fig. 2a), and the high-acidic *ullucus tuberosus*  
278 samples (pH 4) (Fig. 2b), with the highest values of antioxidant activity.

279 With the aim of highlighting the possible similarity among samples on the basis of  
280 betalains and antioxidant activity, the data matrix was subjected to a hierarchical  
281 agglomerative cluster analysis of cases. The squared Euclidean distance as the metric  
282 and the Ward method as the amalgamation rule were taken into account. The  
283 dendograms obtain for each raw material are shown in Fig. S1a and b. As can be seen,  
284 the aggrupation of the samples were similar for each raw material. The original samples  
285 (t<sub>0</sub>) and those submitted to cold storage (4 °C) were closely grouped according to  
286 betalains and antioxidant capacity, and differentiated from those kept at room  
287 temperature and after cooking. In the light of the results, it could be asserted that the  
288 grouping obtained responds to the temperature more than to the pH conditions.

## 289 CONCLUSIONS

290 *Ullucus tuberosus* and *Opuntia dillenii* are two sources of betalains with an important  
291 antioxidant activity. They could be added to highly-acidic foods, but higher quantities  
292 of extract must be added to counteract their lower values of betalains and antioxidant  
293 activity. For both raw materials, their optimal conditions of employment (in order to  
294 yield the highest values of betalains and antioxidant activity) would be in low-acidic  
295 foodstuffs (pH 5 and 6), as long as the conservation was under cold storage conditions.  
296 Although further researchers are needed in relation to natural colorants, this study could  
297 be a great step forward in the field of food, cosmetic and drug industries, among others.

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405 **LEGENDS TO FIGURES**

406 **Figure 1.** Chemical structures of betacyanins and betaxanthins.

407 **Figure 2.** Plot of the *Opuntia dillenii* (a) and *Ullucus tuberosus* (b) in the space defined  
408 by the first two principal components with regard to betalains and **antioxidant capacity**.

409 **Figure S1.** Cluster analysis. Dendrogram obtained after hierarchical agglomerative  
410 cluster analysis performed on betalain compounds and **antioxidant capacity** of all  
411 samples studied. OP, *Opuntia dillenii*; UL, *Ullucus tuberosus*; d, days.