



Update on In Vitro Diagnostic Tools and Treatments for Food Allergies

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Abstract: Food allergy (FA) is an adverse immunological reaction to a specific food that can trigger a wide range of symptoms from mild to life-threatening. This adverse reaction is caused by different immunological mechanisms, such as IgE-mediated, non-IgE-mediated and mixed IgE-mediated reactions. Its epidemiology has had a significant increase in the last decade, more so in developed countries. It is estimated that approximately 2 to 10% of the world's population has FA and this number appears to be increasing and also affecting more children. The diagnosis can be complex and requires the combination of different tests to establish an accurate diagnosis. However, the treatment of FA is based on avoiding the intake of the specific allergenic food, thus being very difficult at times and also controlling the symptoms in case of accidental exposure. Currently, there are other immunomodulatory treatments such as specific allergen immunotherapy or more innovative treatments that can induce a tolerance response. It is important to mention that research in this field is ongoing and clinical trials are underway to assess the safety and efficacy of these different immunotherapy approaches, new treatment pathways are being used to target and promote the tolerance response. In this review, we describe the new in vitro diagnostic tools and therapeutic treatments to show the latest advances in FA management. We conclude that although significant advances have been made to improve therapies and diagnostic tools for FA, there is an urgent need to standardize both so that, in their totality, they help to improve the management of FA.

Keywords: food allergy; immunology; immunotherapy; nanostructures; probiotic; herbal medicine

1. Introduction

Food allergy (FA) is an abnormal and exacerbated response of the immune system to certain food allergens by immunoglobulin E (IgE)-mediated [1], non-IgE-mediated or mixed reaction [2]. The mechanisms involved consist of the dysregulated immune responses and a skewing towards a type 2 immune response. This is associated with production of IgE antibodies and inflammatory cytokines, which are reviewed in detail in the next section. This immune response can result in various ways towards allergen foods, leading to a wide variety of symptoms and clinical manifestations.

The food allergens that most frequently cause the allergies are the following: cow's milk, eggs, fish, peanuts, peach, and soy amongst others [3]. FA patients can suffer a wide variety of symptoms ranging from mild to severe, such as effecting the digestive system, skin, respiratory tract and in extreme cases it can cause a severe allergic reaction known as anaphylaxis [4]. However, some severe reactions can also be associated with other risk factors (co-factors) such as exercise, nonsteroidal anti-inflammatory drugs (NSAIDs), alcohol, alterations in the intestinal microbiota, genetic and even environmental factors.



Citation: Brasal-Prieto, M.; Fernández-Prades, L.; Dakhaoui, H.; Sobrino, F.; López-Enríquez, S.; Palomares, F. Update on In Vitro Diagnostic Tools and Treatments for Food Allergies. *Nutrients* **2023**, *15*, 3744. https://doi.org/10.3390/ nu15173744

Academic Editor: Nicolette W. De Jong

Received: 25 July 2023 Revised: 15 August 2023 Accepted: 24 August 2023 Published: 26 August 2023



Copyright: © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). Theses underlying mechanisms are unknown [5,6], which represents a significant burden for health and quality of life.

FA in our society is growing, already affecting an estimated 1 to 4% of young children, 9 to 11% of adults from Europe and USA, respectively [7]. It is believed that several co-factors may be contributing to this increase.

FA diagnosis can be complex due to the nature of the immunological mechanisms involved in allergic reactions to foods. Therefore, the need to improve strategies for diagnosis and therapy is essential. Diagnosis of FA relies on the combination of clinical/reaction history, skin and IgE testing as well as oral food testing. This is currently favored by method of using in vitro diagnostic techniques such as the basophil activation test (BAT) or the mast cell activation test (MAT) [8] which complement the diagnosis.

In recent years, important advances have been made in the search for other options for the treatment of FA, mainly emphasizing immunomodulatory therapies such as specific allergen immunotherapies (AIT). For example, oral immunotherapy (OIT) using peanut (Arachis hypogea) allergen powder-dnfp (PTHA) in children with peanut allergy has been approved as a treatment in certain cases [9]. Although AIT has achieved a beneficial response for allergic patients, it does not cure the disease so avoiding foods that may contain allergens strictly continues to be applied. Advances in the field of AIT and other novel therapies (nanoparticle design, probiotics, symbiotic, and herbal extracts) have emerged as new options for the development of FA treatments, including the use on monoclonal antibody (anti-IgE). In this review, we provide an overview of the most recent major advances in the diagnosis (in vitro tools) and FA treatment.

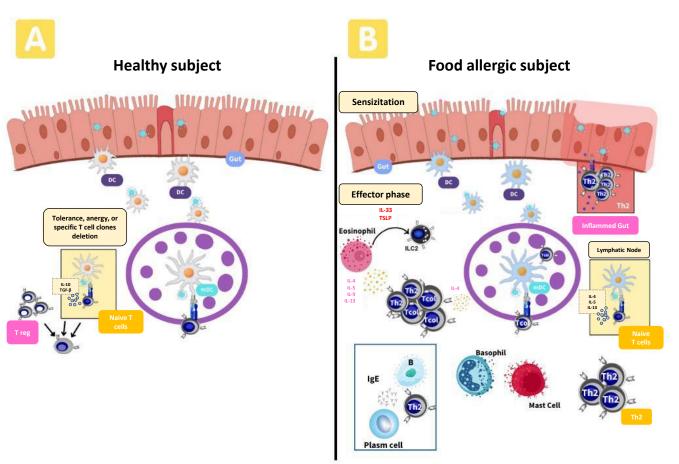
2. Immune Mechanism in FA

In FA, a physio-pathological reaction of the immune system is triggered by the ingestion of a food protein or food allergens. This leads to type I hypersensitivity and immediate reaction, which involves IgE-mediated release of antibodies against the soluble antigen. Theses reactions can be IgE-mediated, non-IgE-mediated, and mixed IgE reactions [10,11]. IgE-mediated FA reactions develop a multi-organ system anaphylaxis. Non-IgE-mediated FAs include a group of disorders characterized by subacute or chronic inflammatory processes affecting the gastrointestinal tract [12]. Mixed IgE and non-IgE-mediated reactions such as food protein-induced allergic proctocolitis, food protein-induced enterocolitis or eosinophilic gastrointestinal disorders [13] have variable symptoms.

2.1. IgE-Mediated Reactions

Sensitization to food allergens occurs in subjects after intake of foods which leads to an adverse inflammatory immune response [14]. The sensitization occurs in a microenvironment that shows damage to the epithelial permeability and dysbiosis (studied in Section 5.1.) which contributes to an imbalance of the host metabolism and immunometabolism. In response, allergens are taken up by dendritic cells (DCs) which interact with a naïve T cell and stimulate the effector response. In this process, DCs undergo phenotypic changes by increasing surface costimulatory receptors and cytokine production to induce a Th2 response (Figure 1) [15].

Recent studies have demonstrated the involvement of innate lymphoid cells (ILCs) emerging as a key in the cause of FA [16,17]. Among them, ILC2 are activated by interleukin (IL)-25, IL-33, thymic stromal lymphopoietin (TSLP) originates from their expansion and production of Th2 cytokines (IL-4, IL-5, and IL-13) [18]. Higher levels of these cytokines have been observed in FA patients [19]. With IL-33, after inducing the ILC2 activation, it produces high IL-4 levels which promotes a Th2 response and can inhibit the regulatory T cell (Treg) function in epithelia (skin and intestinal mucosa) [19]. Therefore, both Th2 and ILC2 release type 2 cytokines that promote B cell differentiation into allergen-specific IgE producing plasma cells which bind to the high-affinity IgE receptor ($Fc \in RI$) [11,19]. Moreover, in vitro assays had shown that both IL-33 and IgE-mediated activation of mast



cells inhibited the generation of a Treg response pattern from T cells in FA an animal model [20].

Figure 1. Immune cells involved in healthy subject's tolerance to food allergens (**A**) and with a food allergic subject with IgE-mediated reactions (**B**).

In the effector phase, re-exposure to the food allergen triggers rapid IgE-mediated degranulation of mast cells, basophils which release histamine and other inflammatory mediators (PGE2). This is accompanied by the adaptive Th2 response, associated with the manifestation of symptoms and the generation of allergen-specific B and T memory cells [11,19,21]. Additionally, it has recently been proposed that specific effector T cell subsets such as Th1, Th17, Tfh13, Th9 and Th22, might also contribute to ongoing FA [21,22].

2.2. Non-IgE-Mediated Reactions

This heterogeneous group of delayed reactions to foods harbor disorders including food protein-induced enterocolitis syndrome (FPIES), food protein-induced allergic proctocolitis (FPIAP), food protein-induced allergic enteropathy (FPE) and food protein-induced dysmotility disorders (gastro-esophageal reflux disorder (GORD) and constipation) [23]. Although the immune mechanism in these disorders remain unclear, they share the characteristics of the complete absence of IgE in the skin serum of patients with FA. Thus, they are being associated generally with a Th2 response pattern such as the classical IgE-mediated FA allergic reactions. In FPES, a pan-leukocyte activation has been observed with association of innate cells and increased gastrointestinal permeability, but the identification of food allergen-specific T lymphocyte is not conclusive [4,24].

Mixed IgE and non-IgE-mediated FAs include eosinophilic gastrointestinal disorders (EGIDs), such as eosinophilic esophagitis (EoE), cow's and soja's milk protein allergy (CMPA and SMPA, respectively) and atopic dermatitis [23]. Contrarily to FPIES, in the EoE disorder, T cells may play a central role in the development of this FA, [24]. Moreover, CMPA has been classified as IgE-mediated immediate reaction, non-IgE-mediated delayed reaction, or a mix of both [25,26].

Therefore, understanding the immunological mechanisms that occur in allergic reactions to foods is essential for an accurate diagnosis. The exploration of these mechanisms will allow us to identify the trigger allergen of the reaction, determine the severity of the allergy, amongst others. However, despite the recent studies, we found there are still limitations in the knowledge of the mechanisms, which means that in vitro diagnostic tools still need to be improved as well.

3. FA In Vitro Supporting Diagnostic Tools

The variability of complex immunological mechanisms contributes to inaccurate diagnosis and complicates the studies on the epidemiology of FA [27–29]. The diagnosis of FA is usually made through a combination of the patient's medical history, SPT, laboratory tests, and/or oral food challenges (OFC). Here, we will review the latest advances in the application of in vitro supporting diagnostic tools, including the current limitations (Table 1).

3.1. Allergen-Specific IgE In Vitro Testing

In vitro tests that measure serum sIgE allergen levels are conventionally used to diagnose FA, already being the standardized diagnostic tools [30,31]. The utility of allergenspecific IgE testing as an alternative to the OFC—the diagnostic standard—is being investigated. A study has determined that the combination of the sIgE levels (ImmunoCAP) and the basophil activation test (BAT) to Ses i 1 can decrease the need for OFC in sesame food allergy (SFA) patients. The authors showed Ses i 1 sIgE levels were not robust enough to be used for diagnosis; however, the simultaneous use of BAT and IgE showed positive correlations [32]. Recently, a systematic review using ImmunoCAP demonstrated that Cor a 9 and Cor a 14 drastically improved the specificity of hazelnut allergy diagnosis, compared to hazelnut extract (HE) sIgE. Using a cutoff of 0.35 ku/L, Cor a 14 sIgE had a specificity of 81.7%, compared to 10.8% for HE sIgE, although sensitivity of HE sIgE was slightly superior (95.5% for HE vs. 77.9% for Cor a 14) [33].

Sensitization to allergen components can be detected by using single plex (Immuno-CAP) or multiplex assays (ISAC) [34]. In particular, the multiplex assays offer a complete profile of multiple allergens in a single test. This is especially useful for identifying different allergens present in a patient and better understanding their sensitivities and allergies. A retrospective study for children with a suspected peanut allergy analyzed the Ara h 2 and Ara h sIgE levels using ImmunoCAP and ISAC. The results showed that the determination of Ara h 6 and Ara h 2 sIgE levels in ISAC was considered good predictors of peanut allergy in children. Even so, the levels of Ara h 2 were comparable to the levels obtained with ImmunoCAP. They concluded that the different peanut components using ISAC was an advantage and clinically useful to detect peanut allergic children [35]. It has been published that the sIgE levels measured with ISAC (Act d 1, Act d 2) showed a sensitivity (59.5%) similar to that of ImmunoCAP (with whole kiwi extract, 63.9%). Act d 1 from ISAC was associated with positive sIgE results from whole kiwi extract detected by ImmunoCAP [36], which indicated that the two in vitro tools showed a similar diagnostic capacity. In a Polish study of a large cohort of children determined sIgE to 112 allergen components using ISAC, and sIgE for hazelnut, Cor a 14, Cashew and Ana o 3, using ImmunoCAP. Both in vitro tools determined that Ana o 3 was the allergen component that predicted anaphylaxis. Although its quantity was lower in ISAC compared to ImmunoCAP, due to the sensitivity

of sIgE. Despite this, the identification of a single allergen component allowed the authors to determine the risk of severe anaphylaxis in FA [37].

In addition to these in vitro techniques, new systems based on IgE multiplex-immunoblot assay are being evaluated. Recently, a study evaluated improvement of lipid transfers protein syndrome (LTP) diagnosis using a EUROLINE-LTP strip. It was observed that there was a positive correlation between the sIgE levels of the single allergen components (Pru p 3, Mal d 3, Ara h 9, Cor a 8 and Jug r 3) with respect to the sIgE levels of the ImmunoCAP. This new IgE multiplex-immunoblot LTP assay showed a good diagnostic performance allowing the culprit food allergies assessment [38]. Another study evaluated the ImmunoCAP and the EUROLINE system for the sensitization profiles towards egg white, yolk extract with the allergen components Gal d 1, 2, 3, 4. The authors determined that sIgE to Gal d 1 (single component) was highly specific in hen's egg that affect corresponding allergic adults [39].

3.2. The Activation Test (BAT)

The BAT is a functional assay that measures the degree of degranulation of the basophils after stimulation with specific allergens, this being basophil reactivity (% CD63+ basophils) and basophil sensitivity (EC50), the main outcomes of the test [40]. Activation of basophils can be detected through upregulation of CD63 activation marker, then its expression is correlated with histamine released. EC50 is the concentration eliciting the half-maximal basophil activation and is a measure of basophil sensitivity [40]. The large number of articles published on BAT in the last year (2022) and the trend towards more regulation by the FDA, it is essential to understand methodological aspects also [41]. For example, passive sensitization experiments, selection of the optimal allergen concentration or the determination of the threshold. [42]. Considering these technical aspects, the BAT has limitations such as the presence of basophils that non-respond to the allergen. Peanut allergy studies (LEAP/LEAP-On studies) have reported that when BAT is performed for a single time, 14% of patients have unreactive basophils [43]. This non-responder status could be associated to transient changes in cell signaling proteins and can be reversed in different culturing conditions.

BAT in FA has already been studied extensively. In a peanut allergy study, the BAT for hazelnut, cashew nut, sesame, almond, peanut discriminated between allergic and nonallergic children, its sensitivity to peanut ranged between 96 and 100% [44]. A prospective study of patients from pediatric centers and universities, children between 0.5 and 17 years with confirmed allergy or sensitization to peanuts and/or tree nuts (almonds, cashews, hazelnuts, pistachios, walnuts) determined that BAT could predict allergic clinical status, and therefore may reduce the need for high-risk OFC in patients [45]. In addition, there was a study that also described these technique as a diagnostic tool in LTP allergy [46] showing that BAT could differentiate between LTP allergic patients and tolerant controls (Ara h 9), although neither the reactivity nor sensitivity could distinguish the severity of clinical symptoms. In a randomized controlled trial in children, it has been showed that BAT reduces the need for a food challenge test in children suspected of IgE-mediated cow's milk allergy (CMA) [47]. The results showed that only a 37% reduction was achieved in in the requirement for food challenge. Similar results were obtained in a clinical trial for children with egg allergy, here BAT for egg allergy was considered a better diagnostic test than to double-blind placebo-controlled food challenge. The results showed a 41% reduction in the number of OFC was achieved [48].

3.3. Mast Cell Activation (MAT)

An alternative test to the BAT is the mast cell activation test (MAT). This consists of measuring the degranulation of mast cells, through levels of CD63, CD107a expression and the release of mediators (prostaglandin D2 and β -hexosaminidase) [49]. MAT is carried out in a similar way to BAT, although it is less sensitive [50]. In a study testing of peanut-sensitized patients (allergic patients) vs. non-peanut-sensitized patients (non-allergic patients), MAT provided conclusive results to aid in the diagnosis of allergic patients [51].

Related to peanut allergy, a study analyzed the activation of mast cells with the presence of Ara h 2 in a group of allergic children. MAT used for Ara h 2 strongly correlated with Arah2-sIgE levels, indicating specific mast cell response, and constituting an alternative diagnostic pathway [52]. Another study demonstrated for the first time the utility of MAT in the diagnosis of LTP allergy, with higher specificity compared to sIgE determination. The results from the study concluded that MAT can be used as complementary tool in the diagnosis of LTP allergy and just with BAT increased the sensitivity up to 95% [53].

3.4. T Cells Assay

The study of allergen-specific T cells is limited by the low frequencies of these cells in blood and the lack of methods able to characterize them. However, last year's development of innovative techniques such as single-cell, genomic, epigenomic and immune repertoire sequencing [54] opened the door to progress the application of T cell assays as diagnostic tool for FA. CD8 T cells attenuate FA in some experimental models, while in humans, CD8 T cells have been shown to expand in response to wheat ingestion corresponding to celiac disease. Additionally, another study showed that CD8 T cells are activated by a peanut peptide in a dependent manner with peanut allergic individuals [55]. These CD8 T cells could express CCR4, suggesting that they were involved with a type 2 allergic immune response. A further study has identified a new Th2 effector, follicular subtypes with potential functional consequences in the pathogenesis and severity of allergic disease in patients with milk-triggered disease (EoE) [56]. A clinical trial identified characteristics of the peanut-specific CD4 T cell response in FA patients, correlated with high clinical sensitivity [57].

Therefore, theses in vitro diagnostic tools have been valuable to better understand the underlying mechanisms of allergic reactions to food and have helped to evaluate and develop treatments, which we will describe in the following sections.

Diagnostic Tools FA Advantages Limitations Specific IgE Sesame [32] Hazelnut [33] Standardized Poor in detection of Peanut allergic children [35] High throughput sensitization from clinically Kiwi [36] Can be automatized reactive FA. LTP (Multiple Food) [38] Egg white and yolk [39] BAT Lack of standardization. Peanut [44] LTP (Multiple Food) [45] High sensitivity Requires fresh blood. Cow's milk [47] Its specificity is variable. MAT Peanut [51,52] Lack of standardization. Requires plasma. LTP Its specificity is stable. Low sensitivity. (Multiple Food) [53]

Table 1. Diagnostic tests for FA including allergen-specific IgE, the basophil activation test, the mast cell activation test, and T cells assay.

Diagnostic Tools	FA	Advantages	Limitations
T cells			
	Peanut [55] Milk [56]	High throughput High sensitivity and specificity Concordant with clinical results	Need to improve the development of molecular techniques, and the characterization of allergens Larger amounts of blood.

Table 1. Cont.

LTP: lipid transfers protein; FA: food allergy.

4. FA Immunotherapy Treatment

Immunomodulatory treatments in FA are therapeutic approaches that regulate the exaggerated immune response (Table 2). These treatments are in various stages of research and development, and some of these have been already used in clinical trials with promising results.

4.1. Allergen Immunotherapy

Allergen immunotherapy (AIT) has been shown to increase the reactivity threshold in most FA individuals [58]. Regarding this, different clinical trials have demonstrated that oral immunotherapy (OIT) for food allergens is safe and effective, improving the quality of life for peanut allergic patients. OIT for peanut has been approved by the FDA and EMA for its use in FA [9,59]. Peanut OIT is safe for children 1 to 3 years of age, from which 71% became desensitized, tolerating peanut protein [60]. Similar results were observed in patients (aged 7 to 55 years) treated with peanut OIT, highlighting that sustained unresponsiveness (SU) was only achievable in less than 35% of those who were successfully desensitized and the SU was maintained for a year [61]. It demonstrated that peanut OIT induces the blocking antibodies [62], low basophil activation and peanutspecific IgE [63].

Epicutaneous immunotherapy (EPIT) employs a non-invasive delivery system of the food allergen. Regarding EPIT, a clinical trial using peanut patches among peanut-allergic children, this assessed the efficacy and adverse events of EPIT. Its results provided a modest success, improving threshold sensitivity to one peanut (300 mg protein) for 35.3% after one year of therapy [64] and for patients after 130 weeks of desensitization using EPIT, this reached a threshold of 400 mg peanut protein [65].

Sublingual immunotherapy (SLIT) is another alternative to OIT and EPIT, which has proven to be safe and effective [66,67]. Peanut SLIT induced long-term desensitization in peanut allergic patients after 3 to 5 years of treatment. Recently, a study reported that during 16 weeks of SLIT with recombinant (r) Mal d 1, but not rBet v 1, significantly improved also for birch pollen-related apple allergy and showed that allergen-sIgE-blocking IgG antibodies were associated with clinical efficacy [68]. Although the SLIT treatment was conducted with only two patients, the results showed that SLIT was able to reduce the levels of antigen-sIgE in severe egg allergy also [69]. Furthermore, in a prospective study where patients diagnosed with LTP allergy and treated with Pru p 3 SLIT were included. One year after the start of Pru p 3 SLIT, the patients had negative OFC to peach and after 2 years of treatment, the OFC remained negative for walnuts and/or peanuts confirming the safety of the therapy [70].

Taking all these results into account and the importance of IgG antibodies in AIT, it has been proposed that IgG antibodies could have a modulatory role in FA. A study has described that FA is associated with increased levels of food-specific IgG and that they interfere with IgE interaction by regulating mast cell and basophil functions [71]. A study for children with suspected FA determined different patterns of sIgG in persistent (peanut) and transient (milk and egg) FA. The authors measured sIgG levels and sIgG isotypes regarding these foods, this showed that for peanut the sIgG, sIgG1, sIgG2, sIgG3

and sIgG4 were higher in peanut-allergic than in non-peanut-allergic patients. However, there is no difference in allergen-specific IgG isotypes when observed between allergic and non-allergic for milk or egg. With exception for milk-specific IgG4 that was higher in non-cows-milk-allergic than in cows-milk-allergic children [72]. They found no evidence that IgG was able to bind to receptors on the surface of mast cells or basophils or to suppress IgE-mediated activation of mast cells or basophils after allergen stimulation. sIgG4 is the most interesting of the IgG antibodies due to its inherent anti-inflammatory properties and its clinical relevance [73,74]. It has been reported that treatment with AIT in patients with peanut allergy, regulated the sIgG4 levels [75]

4.2. Nanoparticles: Platform AIT

Although the use of simple immunotherapies modify the immune response towards food tolerance, there are alternative strategies that promote desensitization such as the use of nanoparticles [11]. A large number of studies are investigating the safety and efficiency of the use of nanoparticle-encapsulated purified peanut extract in peanut allergic patients and animal models [76], masked administration of allergens (peanut) are encapsulated in poly(lactide-co-glycolide) (PLG). Nanoparticles attenuated the anaphylactic response in mouse models with peanut allergy, inducing a more tolerogenic phenotype and conferring protection from intragastric allergen challenge [77]. An innovative therapy has been developed based on the use of lipid nanoparticles (LNP) with encapsulated mRNA encoding peanut allergen epitopes. LNP demonstrated an increase in IL-10-producing Treg cells, suppression of Th2-mediated cytokine production, IgE synthesis and mast cell release in a peanut animal model [78,79].

Nanoparticles with T cells epitopes of arginine kinase with CpG attenuated shrimp allergen enhancing the FOXP3 expression and IL-10 production with a decrease in the Th2 differentiation [80]. Associated with CpG, another study showed that oral pre-treatment with β -lactoglobulin derived peptide and CpG co-encapsulated in PLG nanoparticles prior to sensitization, attenuates the development of CMA in mice [81,82].

New glycosystems functionalized with mannose or fucose and specific ligands combined with Pru p 3 peptides are focused on the modulation of the immune response via C-type lectin receptors (CLRs) [83,84] or Toll-Like Receptors (TLRs) [85]. Regarding this, glycosylated nanostructures combined with immunotherapy, induced long-lasting tolerance with specific transcriptional and methylations changes on DCs [86,87], Treg cells [88] in a peach allergy mouse model.

4.3. Hypoallergenic Proteins: Product AIT

The modification of allergens, through a physical or chemical alteration of their structures have been developed to improve the tolerance response, inducing Th1 and Treg responses. Within the hypoallergenic proteins, synthetic peptides and recombinant proteins have been designed as immunotherapy or vaccines to treat FA [11,89].

Allergy to cow's milk requires the avoidance of cow's milk proteins, currently the use of hypoallergenic milk protein formulation (CM-based hydrolysates or hydrolyzed rice) has been a strategy for the management of CMA [90–92]. The effects of these formulations have showed a tolerance response in human cells [93]. From the hydrolyzed formulas, the structural alteration of the Bos d 5 allergen (B5M) has been identified as a product for use in a hypoallergenic vaccine for CM allergy. B5M induced IgG antibodies and inhibited the degranulation of basophils induced by Bos d 5 [94].

Hypoallergenic derivatives of *Scylla-paramamosain* (mud crab), heat-stable tropomyosin (TM) and myosin light chain (MLC) have been preliminarily explored in vitro. A study recently showed that hypoallergenic derivatives of heat-stable allergens (mtTM, mtMLC) alleviated FA symptoms in a crab allergy model, inducing a significant IL-10 production which equilibrated Th1/Th2 cells [95]. Regarding shrimp allergy, TM has been modified by glycation (GTM) and combined with Al(OH)3 to form hypoallergenic complex and be used in AIT. This hypoallergenic complex has been able to induce desensitization in

allergic reactions in shrimp allergic patients [96,97]. The glycation in the sesame proteins has been shown to reduce the allergenicity of sesame proteins effectively, identifying new hypoallergenic products to treat FA [98].

In addition, hypoallergenic wheat line (1BS-18H) lacking ω 5-gliadin, induced oral tolerance to wheat gluten proteins in a wheat allergy rat model [99]. In this study, the results demonstrated that the early ingestion of 1BS-18H wheat before immunization induced oral tolerance to gluten and ω 5-gliadin, a suppression of gluten-sIgE and IgG₁ levels with an induction of Treg cells.

Although the recombinant Mad d 1 shows high allergenicity [100], recently a study showed that the recombinant Mal d 1 combined with immunotherapy blocked the IgE-mediated reactions also improving apple allergy [68].

4.4. Monoclonal Antibodies (Anti-IgE): Adjuvant AIT

Monoclonal antibodies (anti-IgE) have been considered as adjuvants in food AIT treatments [101] and as monotherapies [102]. These are based on the IgE neutralization. which reduces the sensitivity of the immune system to food allergens, reducing the activation of mast cells and basophils [103].

As monotherapy, an observational study has reported that patients with FA and severe asthma treated with omalizumab were able to increase the allergen threshold for milk, egg, wheat, hazelnut, also control of severe asthma. This resulted in an improvement in the quality of life [102]. An observational study which included children with severe CMA who did not respond successfully to OIT were treated with omalizumab. Interestingly, their CM threshold and IgG4 milk-specific protein levels were significantly increased [104]. Therefore, as monotherapy it can help patients to consume multiple foods and allow for increasing the dose of where its limited.

However, in recent years, several studies have revealed its role as adjuvant for AIT. A phase 2 randomized controlled multisite study using omalizumab combined with OIT decreased time to desensitization and simultaneously desensitizes multiple food allergens [105]. In addition, the combination of omalizumab with peanut OIT induced an increase in peanut intake, a reduction in sIgE, an increase in sIgG4 for peanut, Ara h 1 and Ara h 2 [106,107]. Another study similar determined that OIT with adjunctive anti-IgE can induce immunological changes decreasing type 2 immune response (IL-4 peanut-reactive CD4 T cells, downregulation of CD86 expression in antigen-presenting cell subsets and reduction in pro-inflammatory cytokines) [104]. Additionally, in a cohort of 181 patients, omalizumab dose-related efficacy in OIT was adjusted based on body weight, regardless of total IgE level [108]. Taken together, these results suggest that as an adjunct to OIT, omalizumab can facilitate rapid desensitization, regulate IgE and IgG4 and induce the change immunologically.

Immunotherapy	Treatment Type/FA	Advantages	Limitations
Allergen im- munotherapy	OIT: Peanut [60–63] EPIT: Peanut [65] SLIT: Peanut, Apple, Egg and LTP (Multiple Food) [66–70]	 -AIT is specific treatment that can address underlying cause of the allergic reaction. -Long-term effects, reducing the severity of allergic reactions. -ATI eliminates the need for other medications to control allergic reactions 	-Lack of clinical studies to evaluate its safety and efficacy -Risks of suffering severe allergic reactions during treatment -Prolonged duration of treatment
Nanoparticles	PLG and LNP: Peanut [77–79] T cells epitopes + CpG: Shrimp and CM [80–82] Glycoparticles: Peach [86–88]	-Controlled delivery of allergens. -Improved absorption and bioavailability	-More studies are needed to evaluate their long-term safety and effectiveness -Technical complexity -Lack of regulation and approval for its implementation as therapies

Table 2. Immunotherapeutic approaches for FA: advantages and limitation.

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Immunotherapy	Treatment Type/FA	Advantages	Limitations
Hypoallergenic proteins	Hydrolyzed protein formulation and B5M: CMA. [90–92,94] mtTM, mtMLC: Crab [95] GTM: Shrimp [96,97] 1BS-18H: Wheat [98,99] r Mal d 1: Apple [68]	-Reduced risk of allergic reactions.	-Potential nutrient deficiency -Cost and availability -Altered taste and texture
Monoclonal antibody (anti-IgE)	Monotherapy: Multiple Food and CM [102,104] Multiple therapy: Multiple Food and peanut [105–107]	-Reduced risk of allergic reactions (IgE inhibition). -Increasing food intake. -Improves the quality of life of patients.	-Cost -Side effects -Duration of treatment

Table 2. Cont.

AIT: Allergen immunotherapy; OIT: oral immunotherapy; EPIT: epicutaneous immunotherapy SLIT: sublingual Immunotherapy; PLG: poly(lactide-co-glycolide); LNP: lipid nanoparticle; CMA: cow's milk allergy. r: recombinant.

5. New Therapeutic Approaches

Despite advances in specific treatments, they present certain limitations (Table 3); because of this, the interest in alternative medicine is increasing. Although there are not defined limits between these new natural approaches, we can classify them into three groups: probiotics, herbal medicine and dietary supplements.

5.1. Probiotics and Symbiotics

The microbiome has recently been described as a crucial interface between environmental factors and the development of FA, among other allergic diseases. This relationship has been exploited through the microbial immune modulatory effects of probiotics, prebiotics and symbiotics for the generation of food allergen tolerance, maintaining the Th1/Th2 cell balance, thus improving intestinal barrier function, controlling the intestinal microbiota and its metabolism [109].

The effect of lactic acid bacteria on food protein allergies in infants is studied most, showing a regulation of the intestinal flora of allergic infants, hydrolysis of allergens, inhibition of the inflammatory response, enhancement of the intestinal barrier and modulation of immune cell differentiation [110].

Several studies in FA animal models have demonstrated how some probiotics show the capacity to increase the ratio of effector Treg cells and enhance the secretion of regulatory cytokines (IL-10) such as *Clostridium butyricum* and *Lactobacillus gasseri* [111,112] including the probiotics from the *Bifidobacterium* species [113].

In recent study, the activation of the TLR4 pathway by *Bifidobacterium animalis* KV9 and *Lactobacillus vaginalis* FN3 also led to modulation of the Th1/Th2 balance, attenuating allergic responses in FA mice [114]. The study demonstrated that KV9 and FN3 possessed anti-allergic activities, they modulated the expression of IRF-1 and IRF-4.

Decreasing antigen-specific immunoglobulins has also been an important goal in probiotic therapies. In a murine model of CMA, 3 probiotics with anti-allergic properties have been identified, showing a reduction in the levels of IgE, IgG1, IgG2a, and β -lactoglobulinspecific mast cell protease [115]. In other ovalbumin (OVA)-induced FA animal models, the probiotic *Akkermansia muciniphila* BAA-835 attenuated the levels of IgE anti-OVA and eosinophils [116].

There are numerous interventional trials utilizing prebiotics, probiotics or symbiotics to investigate its effects on FA. In a randomized controlled trial based on the coadministration of probiotic (*Lactobacillus rhamnosus*) with peanut OIT, this reduced peanut-specific IgE levels and increased peanut-specific IgG4 [117]. In addition, the use of probiotic peanut OIT leads to substantial and continued improvement for quality of life 4 years after treatment [118,119]. This outcome has also been described with the combination of heat killed

Lactiplantibacillus plantarum and OIT in CMA. In fact, this clinical trial showed improved tolerance to CM and an increase in the sIgG4 level with reduction in IL-5 and IL-9 [120].

A symbiotic-containing fructo-oligosaccharides and *Bifidobacterium breve* M-16V helped transform the gut microbial composition of non-IgE CM allergic infants to resemble that of healthy infants [121,122]. A pilot study with a symbiotic showed potential in the improvement of symptoms in infants with CMA; however, it lacked a proper a control group [123].

5.2. Herbal Medicine

In the past, herbal plants have been used for medical issues. Nowadays, many studies are being conducted to identify anti-allergic agents from these plants to treat IgE-mediated and non-IgE-mediated allergic reactions [124].

FA Herbal Formula 2 (FAHF-2), composed by 9 different herbs was the first plant origin-based drug approved by US FDA for FA treatment (2007). New techniques have been applied to improve some of its disadvantages. Two types of formulas have been purified, this being B-FAHF-2 (butanol purified FA herbal formula-2) and E-B-FAHF-2 (ethyl acetate and butanol purified FA herbal formula-2) [125]. Recently, in vivo experiments into murine models of peanut allergy showed that E-B-FHFA-2 and its active compound (berberine) protected the mice from anaphylaxis, decreased IgE levels and IgE plasma cells [126].

Apart from FAFH-2, in addition to its improved versions, many other plant origin compounds have been tested in vitro and in vivo for FA treatments. In this sense, N-nornuciferine and lirinidine alkaloids from lotus seed pods showed potent anti-food allergic activity in RBL-2H3 cells measured as β -hexosaminidase activity [127].

The oleuropein was evaluated for the prevention of OVA- induced FA. The results of their studies showed that sensitized mice treated with oleuropein had decreased levels of IgE, IgG and histamine. Moreover, oleuropein enhanced intestinal epithelium, altering mucosal mast cells and Treg cells [128]. Additionally, other compounds as the isoflavones isolated from kakkonto, a Japanese herbal medicine (Genistein and genistin) suppressed allergic symptoms in OVA-induced FA mice [129].

5.3. Dietary Supplements

Some components of food have been related to pro-inflammatory effects (fat, sugar, folate deficit) and others to anti-inflammatory effects (omega-3 PUFAs, vitamin D, polyphenols). Some studies have focused on diet supplementation as a treatment for FA disorders [130].

A study observed that ginger could upregulate the expression of the retinoic acid (RA) receptor in the gut, which suggested that immune responses mediated through RA regulation could contribute to the suppression of FA inflammation [131].

An OVA-sensitized mouse model supplemented with olive oil had a reduction in allergic symptoms, with a reduction in cytokines associated with Th2 cells and an increase in cytokines released by Treg cells [132]. The combined supplementation with arachidonic and docosahexaenoic acids during suckling period presented beneficial effects on OVA oral tolerance of the rat offspring, revealing that this combination decreased Th2 immune response due to an increase in Th1 cytokine levels [133]. A meta-analysis study concluded that omega-3 supplementation during pregnancy reduces the risk of FA in children. On the other hand, omega-3 supplementation during childhood did not show any beneficial effects [134].

Therapeutic Approaches	Model/FA	Achievements
Probiotics and symbiotics	Clostridium butyricum, Lactobacillus gasseri and Bifidobacterium species [111–113] and Lactobacillus vaginalis [114]: FA animal model. Lactobacillus paracasei L9: CM [135] Leuconostoc citreum: FA animal model [116] Akkermansia muciniphila: FA animal model [136]. Lactobacillus rhamnosus: Penaut [117] Synbiotic-containing fructooligosaccharides and Bifidobacterium breve M-16V:CM [121,122].	 -Increase the ratio of effector Treg cells and enhance the secretion of regulatory cytokines. -Modulation of Th1/Th2 balance and attenuation of allergic reaction. -IgE reduction -Transformation of gut microbial to improve the healthy infants.
Herbal medicine	E-B-FAHF-2 and B-FAHF-2: Peanut [125,126]. Berberine: peanut and cholera toxine animal model [126]. Oleuropein: FA animal model [128].	-Reduction in anaphylaxis symptoms. -Reduction in histamine and IgE plasma levels. -Reduction in B cells in spleen and modification of gut microbiota
Dietary supplements	Ginger: CACO2 cells [131]. Olive oil: FA animal model [132]. Arachidonic and docosahexaenoic acids (PUFAs): mother during suckling period levels [133]. Omega-3 supplementation: mothers during pregnancy [134].	-Suppression of FA inflammation. -Reduction in allergic symptoms. -Th2 cells reduction and increase in Treg. -Increase in oral tolerance in children. -Increase in Th1 cytokines levels. -Reduction in the risk of FA in children.

Table 3. Therapeutic approaches and their achievements.

FA: food allergy; CMA: cow's milk allergy.

6. Conclusions

FAs are complex and mediated by a variety of immunological mechanisms. The IgE-mediated response is the most common, but cellular mechanisms and other types of immunological reactions can also be involved. Understanding these mechanisms is essential for the proper diagnosis and treatment of FA. In this sense, it is essential to note that diagnostic tools continue to develop constantly, since they have certain limitations, such as needing to be agreed upon or the low sensitivity. Nonetheless, theses in vitro diagnostic tools have helped to evaluate and develop new treatments for FA.

Regarding immunomodulatory approaches as treatment for FA, many of them are still in the research and development stages. However, all of them are focused on reducing the allergic response by inducing a tolerance response. Despite this, they still present certain weaknesses that give rise to new therapeutic approaches. More studies are still needed to investigate the role of IgG in FA.

Overall, new therapeutic approaches for the prevention and treatment of FAs have shown clinical potential; however, the results of these trials remain controversial. With contradiction, these approaches face several limitations including the heterogeneity in dosages/administration in trials and the mechanism of probiotics or herbal medicine for FA remains unknown. Further studies are required to discern the actual benefits of these new therapies in FA.

Author Contributions: Conceptualization, S.L.-E. and F.P.; writing—original draft preparation, M.B.-P., L.F.-P., S.L.-E. and F.P. writing—review and editing, H.D., S.L.-E. and F.P., visualization, F.S.; supervision, F.P. and F.S. All authors have read and agreed to the published version of the manuscript.

Funding: This work was supported by grants by "VII Plan Propio de Investigación y Transferencia" from University of Seville (Project-20220000421 and Project-2023/00000482), and by Ministry of Science and Innovation trough research Ramon y Cajal program (RYC2021-031256-I) from Spain.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: Not applicable.

Conflicts of Interest: The authors declare no conflict of interest.

Abbreviations

ATI	Allergen immunotherapy
BAT	Basophil activation test
CMPA	Cow milk protein
EGID	Eosinophilic gastrointestinal disorders
EoE	Eosinophilic esophagitis
EMA	European Medicines Agency
FA	Food allergy
FPIES	Food protein-induced enterocolitis syndrome
FPIAP	Food protein-induced allergic proctocolitis
FPE	Food protein-induced allergic enteropathy
FDA	Food and Drug Administration
GORD	Gastro-esophageal reflux disorder
MAT	Mast cell activation test
NSAIDs	Nonsteroidal anti-inflammatory drugs
OFC	Oral food challenge
OIT	Oral immunotherapy
SPT	Skin prick test
SMPA	Soja's milk protein allergy
sIgE	Specific immunoglobulin E
TSLP	Thymic stromal lymphopoietin

References

- Ballegaard, A.R.; Bogh, K.L. Intestinal protein uptake and IgE-mediated food allergy. *Food Res. Int.* 2023, 163, 112150. [CrossRef] [PubMed]
- 2. Mendonca, C.E.; Andreae, D.A. Food Allergy. Prim. Care 2023, 50, 205–220. [CrossRef]
- Feng, H.; Liu, Y.; Xiong, X.; Xu, Q.; Zhang, Z.; Wu, Y.; Lu, Y. Epidemiological survey of self-reported food allergy among university students in China. *Medicine* 2022, 101, e29606. [CrossRef] [PubMed]
- Baker, M.G.; Cecilia Berin, M.; Sicherer, S. Update on Food Protein-Induced Enterocolitis Syndrome (FPIES). Curr. Allergy Asthma Rep. 2022, 22, 113–122. [CrossRef] [PubMed]
- Turner, P.J.; Arasi, S.; Ballmer-Weber, B.; Baseggio Conrado, A.; Deschildre, A.; Gerdts, J.; Halken, S.; Muraro, A.; Patel, N.; Van Ree, R.; et al. Risk factors for severe reactions in food allergy: Rapid evidence review with meta-analysis. *Allergy* 2022, 77, 2634–2652. [CrossRef]
- Munoz-Cano, R.; San Bartolome, C.; Casas-Saucedo, R.; Araujo, G.; Gelis, S.; Ruano-Zaragoza, M.; Roca-Ferrer, J.; Palomares, F.; Martin, M.; Bartra, J.; et al. Immune-Mediated Mechanisms in Cofactor-Dependent Food Allergy and Anaphylaxis: Effect of Cofactors in Basophils and Mast Cells. *Front. Immunol.* 2020, *11*, 623071. [CrossRef]
- 7. Sampath, V.; Abrams, E.M.; Adlou, B.; Akdis, C.; Akdis, M.; Brough, H.A.; Chan, S.; Chatchatee, P.; Chinthrajah, R.S.; Cocco, R.R.; et al. Food allergy across the globe. *J. Allergy Clin. Immunol.* **2021**, *148*, 1347–1364. [CrossRef]
- 8. Paranjape, A.; Tsai, M.; Mukai, K.; Hoh, R.A.; Joshi, S.A.; Chinthrajah, R.S.; Nadeau, K.C.; Boyd, S.D.; Galli, S.J. Oral Immunotherapy and Basophil and Mast Cell Reactivity in Food Allergy. *Front. Immunol.* **2020**, *11*, 602660. [CrossRef]
- Fernandez-Rivas, M.; Vereda, A.; Vickery, B.P.; Sharma, V.; Nilsson, C.; Muraro, A.; Hourihane, J.O.; DunnGalvin, A.; du Toit, G.; Blumchen, K.; et al. Open-label follow-on study evaluating the efficacy, safety, and quality of life with extended daily oral immunotherapy in children with peanut allergy. *Allergy* 2022, 77, 991–1003. [CrossRef]
- Renz, H.; Allen, K.J.; Sicherer, S.H.; Sampson, H.A.; Lack, G.; Beyer, K.; Oettgen, H.C. Food allergy. Nat. Rev. Dis. Primers 2018, 4, 17098. [CrossRef]
- 11. Mayorga, C.; Palomares, F.; Canas, J.A.; Perez-Sanchez, N.; Nunez, R.; Torres, M.J.; Gomez, F. New Insights in Therapy for Food Allergy. *Foods* **2021**, *10*, 1037. [CrossRef]
- 12. Zhang, S.; Sicherer, S.; Berin, M.C.; Agyemang, A. Pathophysiology of Non-IgE-Mediated Food Allergy. *Immunotargets Ther.* **2021**, 10, 431–446. [CrossRef] [PubMed]
- Calvani, M.; Anania, C.; Cuomo, B.; D'Auria, E.; Decimo, F.; Indirli, G.C.; Marseglia, G.; Mastrorilli, V.; Sartorio, M.U.A.; Santoro, A.; et al. Non-IgE- or Mixed IgE/Non-IgE-Mediated Gastrointestinal Food Allergies in the First Years of Life: Old and New Tools for Diagnosis. *Nutrients* 2021, 13, 226. [CrossRef]
- Yu, W.; Freeland, D.M.H.; Nadeau, K.C. Food allergy: Immune mechanisms, diagnosis and immunotherapy. *Nat. Rev. Immunol.* 2016, 16, 751–765. [CrossRef] [PubMed]
- 15. Ellenbogen, Y.; Jimenez-Saiz, R.; Spill, P.; Chu, D.K.; Waserman, S.; Jordana, M. The Initiation of Th2 Immunity towards Food Allergens. *Int. J. Mol. Sci.* 2018, 19, 1447. [CrossRef] [PubMed]
- Palomares, F.; Gomez, F.; Bogas, G.; Maggi, L.; Cosmi, L.; Annunziato, F.; Nunez, R.; Perez, N.; Munoz-Cano, R.; Torres, M.J.; et al. Innate lymphoid cells type 2 in LTP-allergic patients and their modulation during sublingual immunotherapy. *Allergy* 2021, 76, 2253–2256. [CrossRef]

- Zheng, H.; Zhang, Y.; Pan, J.; Liu, N.; Qin, Y.; Qiu, L.; Liu, M.; Wang, T. The Role of Type 2 Innate Lymphoid Cells in Allergic Diseases. Front. Immunol. 2021, 12, 586078. [CrossRef]
- Kabata, H.; Moro, K.; Koyasu, S. The group 2 innate lymphoid cell (ILC2) regulatory network and its underlying mechanisms. *Immunol. Rev.* 2018, 286, 37–52. [CrossRef] [PubMed]
- Sahiner, U.M.; Layhadi, J.A.; Golebski, K.; Istvan Komlosi, Z.; Peng, Y.; Sekerel, B.; Durham, S.R.; Brough, H.; Morita, H.; Akdis, M.; et al. Innate lymphoid cells: The missing part of a puzzle in food allergy. *Allergy* 2021, *76*, 2002–2016. [CrossRef]
- Benede, S.; Tordesillas, L.; Berin, C. Demonstration of distinct pathways of mast cell-dependent inhibition of Treg generation using murine bone marrow-derived mast cells. *Allergy* 2020, 75, 2088–2091. [CrossRef]
- Locke, A.; Hung, L.; Upton, J.E.M.; O'Mahony, L.; Hoang, J.; Eiwegger, T. An update on recent developments and highlights in food allergy. *Allergy* 2023. [CrossRef] [PubMed]
- 22. Crespo, J.F.; Cabanillas, B. Recent advances in cellular and molecular mechanisms of IgE-mediated food allergy. *Food Chem.* 2023, 411, 135500. [CrossRef]
- 23. Al-Iede, M.; Sarhan, L.; Alshrouf, M.A.; Said, Y. Perspectives on Non-IgE-Mediated Gastrointestinal Food Allergy in Pediatrics: A Review of Current Evidence and Guidelines. *J. Asthma Allergy* **2023**, *16*, 279–291. [CrossRef] [PubMed]
- 24. Cianferoni, A. Non-IgE Mediated Food Allergy. Curr. Pediatr. Rev. 2020, 16, 95–105. [CrossRef] [PubMed]
- Martorell-Aragones, A.; Echeverria-Zudaire, L.; Alonso-Lebrero, E.; Bone-Calvo, J.; Martin-Munoz, M.F.; Nevot-Falco, S.; Piquer-Gibert, M.; Valdesoiro-Navarrete, L.; Food allergy committee of SEICAP. Position document: IgE-mediated cow's milk allergy. *Allergol. Immunopathol.* 2015, 43, 507–526. [CrossRef]
- Cronin, C.; Ramesh, Y.; De Pieri, C.; Velasco, R.; Trujillo, J. 'Early Introduction' of Cow's Milk for Children with IgE-Mediated Cow's Milk Protein Allergy: A Review of Current and Emerging Approaches for CMPA Management. *Nutrients* 2023, 15, 1397. [CrossRef]
- De Martinis, M.; Sirufo, M.M.; Viscido, A.; Ginaldi, L. Food Allergy Insights: A Changing Landscape. Arch. Immunol. Ther. Exp. 2020, 68, 8. [CrossRef]
- Sirufo, M.M.; Suppa, M.; Ginaldi, L.; De Martinis, M. Does Allergy Break Bones? Osteoporosis and Its Connection to Allergy. Int. J. Mol. Sci. 2020, 21, 712. [CrossRef]
- 29. De Martinis, M.; Sirufo, M.M.; Suppa, M.; Ginaldi, L. New Perspectives in Food Allergy. Int. J. Mol. Sci. 2020, 21, 1474. [CrossRef]
- Arasi, S.; Barni, S.; Mastrorilli, C.; Comberiati, P.; Chiera, F.; Pelosi, U.; Paravati, F.; Caimmi, D. Role of in vitro testing in food allergy. *Pediatr. Allergy Immunol.* 2020, 31 (Suppl. S26), 36–38. [CrossRef]
- Ansotegui, I.J.; Melioli, G.; Canonica, G.W.; Caraballo, L.; Villa, E.; Ebisawa, M.; Passalacqua, G.; Savi, E.; Ebo, D.; Gomez, R.M.; et al. IgE allergy diagnostics and other relevant tests in allergy, a World Allergy Organization position paper. *World Allergy Organ. J.* 2020, 13, 100080. [CrossRef] [PubMed]
- Goldberg, M.R.; Appel, M.Y.; Nachshon, L.; Holmqvist, M.; Epstein-Rigbi, N.; Levy, M.B.; Lidholm, J.; Elizur, A. Combinatorial advantage of Ses i 1-specific IgE and basophil activation for diagnosis of sesame food allergy. *Pediatr. Allergy Immunol.* 2021, 32, 1482–1489. [CrossRef] [PubMed]
- Nilsson, C.; Berthold, M.; Mascialino, B.; Orme, M.; Sjolander, S.; Hamilton, R. Allergen components in diagnosing childhood hazelnut allergy: Systematic literature review and meta-analysis. *Pediatr. Allergy Immunol.* 2020, 31, 186–196. [CrossRef] [PubMed]
- Maesa, J.M.; Dobrzynska, A.; Banos-Alvarez, E.; Isabel-Gomez, R.; Blasco-Amaro, J.A. ImmunoCAP ISAC in food allergy diagnosis: A systematic review of diagnostic test accuracy. *Clin. Exp. Allergy* 2021, *51*, 778–789. [CrossRef]
- 35. Brand, H.K.; Schreurs, M.W.J.; Emons, J.A.M.; Gerth van Wijk, R.; de Groot, H.; Arends, N.J.T. Peanut components measured by ISAC: Comparison with ImmunoCap and clinical relevance in peanut allergic children. *Clin. Mol. Allergy* **2021**, *19*, 14. [CrossRef]
- D'Amelio, C.M.; Bernad, A.; Garcia-Figueroa, B.E.; Garrido-Fernandez, S.; Azofra, J.; Beristain, A.; Bueno-Diaz, C.; Garrido-Arandia, M.; Gastaminza, G.; Ferrer, M.; et al. Unraveling the Diagnosis of Kiwifruit Allergy: Usefulness of Current Diagnostic Tests. J. Investig. Allergol. Clin. Immunol. 2022, 32, 206–212. [CrossRef]
- 37. Blazowski, L.; Majak, P.; Kurzawa, R.; Kuna, P.; Jerzynska, J. Food allergy endotype with high risk of severe anaphylaxis in children-Monosensitization to cashew 2S albumin Ana o 3. *Allergy* **2019**, *74*, 1945–1955. [CrossRef]
- Sara, B.V.; Ulrike, F.; Bettina, B.; Yvonne, W.; Teresa, P.; Clara, S.B.; Giovanna, A.S.; Rocio, C.S.; Maria, T.; Rocio, L.; et al. Improving In Vitro Detection of Sensitization to Lipid Transfer Proteins: A New Molecular Multiplex IgE Assay. *Mol. Nutr. Food Res.* 2023, 67, e2200906. [CrossRef]
- 39. Ehlers, A.M.; Otten, H.G.; Wierzba, E.; Flugge, U.; Le, T.M.; Knulst, A.C.; Suer, W. Detection of specific IgE against linear epitopes from Gal d 1 has additional value in diagnosing hen's egg allergy in adults. *Clin. Exp. Allergy* **2020**, *50*, 1415–1423. [CrossRef]
- 40. Santos, A.F.; Alpan, O.; Hoffmann, H.J. Basophil activation test: Mechanisms and considerations for use in clinical trials and clinical practice. *Allergy* **2021**, *76*, 2420–2432. [CrossRef]
- Alpan, O.; Wasserman, R.L.; Kim, T.; Darter, A.; Shah, A.; Jones, D.; McNeil, D.; Li, H.; Ispas, L.; Rathkopf, M.; et al. Towards an FDA-cleared basophil activation test. *Front. Allergy* 2022, *3*, 1009437. [CrossRef] [PubMed]
- 42. Ebo, D.G.; Bridts, C.H.; Mertens, C.H.; Sabato, V. Principles, potential, and limitations of ex vivo basophil activation by flow cytometry in allergology: A narrative review. *J. Allergy Clin. Immunol.* **2021**, 147, 1143–1153. [CrossRef]
- Santos, A.F.; Du Toit, G.; O'Rourke, C.; Becares, N.; Couto-Francisco, N.; Radulovic, S.; Khaleva, E.; Basting, M.; Harris, K.M.; Larson, D.; et al. Biomarkers of severity and threshold of allergic reactions during oral peanut challenges. *J. Allergy Clin. Immunol.* 2020, 146, 344–355. [CrossRef] [PubMed]

- Santos, A.F.; Bergmann, M.; Brough, H.A.; Couto-Francisco, N.; Kwok, M.; Panetta, V.; Haddad, D.; Lack, G.; Eigenmann, P.; Caubet, J.C. Basophil Activation Test Reduces Oral Food Challenges to Nuts and Sesame. J. Allergy Clin. Immunol. Pract. 2021, 9, 2016–2027. [CrossRef]
- Duan, L.; Celik, A.; Hoang, J.A.; Schmidthaler, K.; So, D.; Yin, X.; Ditlof, C.M.; Ponce, M.; Upton, J.E.M.; Lee, J.S.; et al. Basophil activation test shows high accuracy in the diagnosis of peanut and tree nut allergy: The Markers of Nut Allergy Study. *Allergy* 2021, 76, 1800–1812. [CrossRef] [PubMed]
- 46. Canas, J.A.; Perez-Sanchez, N.; Lopera-Doblas, L.; Palomares, F.; Molina, A.; Bartra, J.; Torres, M.J.; Gomez, F.; Mayorga, C. Basophil Activation Test Utility as a Diagnostic Tool in LTP Allergy. *Int. J. Mol. Sci.* **2022**, 23, 4979. [CrossRef] [PubMed]
- Ruinemans-Koerts, J.; Schmidt-Hieltjes, Y.; Jansen, A.; Savelkoul, H.F.J.; Plaisier, A.; van Setten, P. The Basophil Activation Test reduces the need for a food challenge test in children suspected of IgE-mediated cow's milk allergy. *Clin. Exp. Allergy* 2019, 49, 350–356. [CrossRef]
- Krawiec, M.; Radulovic, S.; Foong, R.X.; Marques-Mejias, A.; Bartha, I.; Kwok, M.; Jama, Z.; Harrison, F.; Ricci, C.; Lack, G.; et al. Diagnostic utility of allergy tests to predict baked egg and lightly cooked egg allergies compared to double-blind placebo-controlled food challenges. *Allergy* 2023. [CrossRef]
- Bahri, R.; Custovic, A.; Korosec, P.; Tsoumani, M.; Barron, M.; Wu, J.; Sayers, R.; Weimann, A.; Ruiz-Garcia, M.; Patel, N.; et al. Mast cell activation test in the diagnosis of allergic disease and anaphylaxis. *J. Allergy Clin. Immunol.* 2018, 142, 485–496. [CrossRef]
- 50. Peters, R.L.; Krawiec, M.; Koplin, J.J.; Santos, A.F. Update on food allergy. Pediatr. Allergy Immunol. 2021, 32, 647–657. [CrossRef]
- Santos, A.F.; Couto-Francisco, N.; Becares, N.; Kwok, M.; Bahnson, H.T.; Lack, G. A novel human mast cell activation test for peanut allergy. J. Allergy Clin. Immunol. 2018, 142, 689–691. [CrossRef] [PubMed]
- Ji, C.; Huang, Y.; Yeung, L.H.; Hemmings, O.; Jama, Z.; Kwok, M.; Lack, G.; Santos, A.F. Ara h 2-Specific IgE Presence Rather Than Its Function Is the Best Predictor of Mast Cell Activation in Children. *J. Allergy Clin. Immunol. Pract.* 2023, 11, 1154–1161. [CrossRef] [PubMed]
- Cespedes, J.A.; Lebron-Martin, C.; Garcia-Oton, R.; Delgado, M.J.; Martin-Astorga, M.D.C.; Perez-Sanchez, N.; Gomez, F.; Torres, M.J.; Canas, J.A.; Aranda, C.J.; et al. In vitro supporting diagnostic tools in plant-food allergy. *Allergy* 2023. *Online ahead of print*.. [CrossRef] [PubMed]
- 54. Lewis, S.A.; Peters, B. T-cell epitope discovery and single-cell technologies to advance food allergy research. *J. Allergy Clin. Immunol.* **2023**, *151*, 15–20. [CrossRef]
- Yu, W.; Zhou, X.; Dunham, D.; Lyu, S.C.; Manohar, M.; Zhang, W.; Zhao, F.; Davis, M.M.; Nadeau, K. Allergen-specific CD8(+) T cells in peanut-allergic individuals. J. Allergy Clin. Immunol. 2019, 143, 1948–1952. [CrossRef]
- Morgan, D.M.; Ruiter, B.; Smith, N.P.; Tu, A.A.; Monian, B.; Stone, B.E.; Virk-Hundal, N.; Yuan, Q.; Shreffler, W.G.; Love, J.C. Clonally expanded, GPR15-expressing pathogenic effector T(H)2 cells are associated with eosinophilic esophagitis. *Sci. Immunol.* 2021, *6*, eabi5586. [CrossRef] [PubMed]
- Ruiter, B.; Smith, N.P.; Monian, B.; Tu, A.A.; Fleming, E.; Virkud, Y.V.; Patil, S.U.; Whittaker, C.A.; Love, J.C.; Shreffler, W.G. Expansion of the CD4(+) effector T-cell repertoire characterizes peanut-allergic patients with heightened clinical sensitivity. J. Allergy Clin. Immunol. 2020, 145, 270–282. [CrossRef]
- Sampath, V.; Sindher, S.B.; Alvarez Pinzon, A.M.; Nadeau, K.C. Can food allergy be cured? What are the future prospects? *Allergy* 2020, 75, 1316–1326. [CrossRef]
- Vickery, B.P.; Vereda, A.; Nilsson, C.; du Toit, G.; Shreffler, W.G.; Burks, A.W.; Jones, S.M.; Fernandez-Rivas, M.; Blumchen, K.; Hourihane, J.O.B.; et al. Continuous and daily oral immunotherapy for peanut allergy: Results from a 2-year open-label follow-on study. J. Allergy Clin. Immunol. Pract. 2021, 9, 1879–1889. [CrossRef]
- Jones, S.M.; Kim, E.H.; Nadeau, K.C.; Nowak-Wegrzyn, A.; Wood, R.A.; Sampson, H.A.; Scurlock, A.M.; Chinthrajah, S.; Wang, J.; Pesek, R.D.; et al. Efficacy and safety of oral immunotherapy in children aged 1-3 years with peanut allergy (the Immune Tolerance Network IMPACT trial): A randomised placebo-controlled study. *Lancet* 2022, 399, 359–371. [CrossRef]
- Chinthrajah, R.S.; Purington, N.; Andorf, S.; Long, A.; O'Laughlin, K.L.; Lyu, S.C.; Manohar, M.; Boyd, S.D.; Tibshirani, R.; Maecker, H.; et al. Sustained outcomes in oral immunotherapy for peanut allergy (POISED study): A large, randomised, double-blind, placebo-controlled, phase 2 study. *Lancet* 2019, 394, 1437–1449. [CrossRef] [PubMed]
- Santos, A.F.; James, L.K.; Kwok, M.; McKendry, R.T.; Anagnostou, K.; Clark, A.T.; Lack, G. Peanut oral immunotherapy induces blocking antibodies but does not change the functional characteristics of peanut-specific IgE. J. Allergy Clin. Immunol. 2020, 145, 440–443. [CrossRef] [PubMed]
- 63. Tsai, M.; Mukai, K.; Chinthrajah, R.S.; Nadeau, K.C.; Galli, S.J. Sustained successful peanut oral immunotherapy associated with low basophil activation and peanut-specific IgE. J. Allergy Clin. Immunol. **2020**, 145, 885–896.e6. [CrossRef] [PubMed]
- 64. Fleischer, D.M.; Greenhawt, M.; Sussman, G.; Begin, P.; Nowak-Wegrzyn, A.; Petroni, D.; Beyer, K.; Brown-Whitehorn, T.; Hebert, J.; Hourihane, J.O.; et al. Effect of Epicutaneous Immunotherapy vs Placebo on Reaction to Peanut Protein Ingestion among Children with Peanut Allergy: The PEPITES Randomized Clinical Trial. JAMA 2019, 321, 946–955. [CrossRef]
- Scurlock, A.M.; Burks, A.W.; Sicherer, S.H.; Leung, D.Y.M.; Kim, E.H.; Henning, A.K.; Dawson, P.; Lindblad, R.W.; Berin, M.C.; Cho, C.B.; et al. Epicutaneous immunotherapy for treatment of peanut allergy: Follow-up from the Consortium for Food Allergy Research. J. Allergy Clin. Immunol. 2021, 147, 992–1003. [CrossRef] [PubMed]

- 66. Schworer, S.A.; Kim, E.H. Sublingual immunotherapy for food allergy and its future directions. *Immunotherapy* **2020**, *12*, 921–931. [CrossRef]
- 67. DuBuske, L. Efficacy and safety of sublingual allergen immunotherapy. Allergy Asthma Proc. 2022, 43, 272–280. [CrossRef]
- 68. Sanchez Acosta, G.; Kinaciyan, T.; Kitzmuller, C.; Mobs, C.; Pfutzner, W.; Bohle, B. IgE-blocking antibodies following SLIT with recombinant Mal d 1 accord with improved apple allergy. *J. Allergy Clin. Immunol.* **2020**, *146*, 894–900.e2. [CrossRef]
- 69. Sagara, N.; Fujita, S.; Suzuki, R.; Aota, A.; Akashi, K.; Katsunuma, T. Successful sublingual immunotherapy for severe egg allergy in children: A case report. *Allergy Asthma Clin. Immunol.* **2021**, *17*, 2. [CrossRef]
- 70. Beitia, J.M.; Vega Castro, A.; Cardenas, R.; Pena-Arellano, M.I. Pru p 3 Sublingual Immunotherapy in Patients with Lipid Transfer Protein Syndrome: Is It Worth? *Int. Arch. Allergy Immunol.* **2021**, *182*, 447–454. [CrossRef]
- 71. Kanagaratham, C.; El Ansari, Y.S.; Lewis, O.L.; Oettgen, H.C. IgE and IgG Antibodies as Regulators of Mast Cell and Basophil Functions in Food Allergy. *Front. Immunol.* **2020**, *11*, 603050. [CrossRef]
- 72. McKendry, R.T.; Kwok, M.; Hemmings, O.; James, L.K.; Santos, A.F. Allergen-specific IgG show distinct patterns in persistent and transient food allergy. *Pediatr. Allergy Immunol.* **2021**, *32*, 1508–1518. [CrossRef] [PubMed]
- Qin, L.; Tang, L.F.; Cheng, L.; Wang, H.Y. The clinical significance of allergen-specific IgG4 in allergic diseases. *Front. Immunol.* 2022, 13, 1032909. [CrossRef] [PubMed]
- 74. Platts-Mills, T.A.E.; Keshavarz, B.; Wilson, J.M.; Li, R.C.; Heymann, P.W.; Gold, D.R.; McGowan, E.C.; Erwin, E.A. An Overview of the Relevance of IgG4 Antibodies in Allergic Disease with a Focus on Food Allergens. *Children* **2021**, *8*, 418. [CrossRef] [PubMed]
- Smeekens, J.M.; Baloh, C.; Lim, N.; Larson, D.; Qin, T.; Wheatley, L.; Kim, E.H.; Jones, S.M.; Burks, A.W.; Kulis, M.D. Peanut-Specific IgG4 and IgA in Saliva Are Modulated by Peanut Oral Immunotherapy. *J. Allergy Clin. Immunol. Pract.* 2022, 10, 3270–3275. [CrossRef]
- 76. Sindher, S.B.; Long, A.; Chin, A.R.; Hy, A.; Sampath, V.; Nadeau, K.C.; Chinthrajah, R.S. Food allergy, mechanisms, diagnosis and treatment: Innovation through a multi-targeted approach. *Allergy* **2022**, *77*, 2937–2948. [CrossRef]
- Hughes, K.R.; Saunders, M.N.; Landers, J.J.; Janczak, K.W.; Turkistani, H.; Rad, L.M.; Miller, S.D.; Podojil, J.R.; Shea, L.D.; O'Konek, J.J. Masked Delivery of Allergen in Nanoparticles Safely Attenuates Anaphylactic Response in Murine Models of Peanut Allergy. *Front. Allergy* 2022, *3*, 829605. [CrossRef]
- Xu, X.; Wang, X.; Liao, Y.P.; Luo, L.; Xia, T.; Nel, A.E. Use of a Liver-Targeting Immune-Tolerogenic mRNA Lipid Nanoparticle Platform to Treat Peanut-Induced Anaphylaxis by Single- and Multiple-Epitope Nucleotide Sequence Delivery. ACS Nano 2023, 17, 4942–4957. [CrossRef]
- 79. Liu, Q.; Wang, X.; Liao, Y.P.; Chang, C.H.; Li, J.; Xia, T.; Nel, A.E. Use of a Liver-targeting Nanoparticle Platform to Intervene in Peanut-induced anaphylaxis through delivery of an Ara h2 T-cell Epitope. *Nano Today* **2022**, *42*, 101370. [CrossRef]
- 80. Hong, J.; Gao, Q.; Xiao, X.; Cao, H.; Yuan, R.; Liu, Z.; Chen, T. T cell epitope of arginine kinase with CpG co-encapsulated nanoparticles attenuates a shrimp allergen-induced Th2-bias food allergy. *Biosci. Biotechnol. Biochem.* 2020, *84*, 804–814. [CrossRef]
- Liu, M.; Thijssen, S.; Hennink, W.E.; Garssen, J.; van Nostrum, C.F.; Willemsen, L.E.M. Oral pretreatment with beta-lactoglobulin derived peptide and CpG co-encapsulated in PLGA nanoparticles prior to sensitizations attenuates cow's milk allergy development in mice. *Front. Immunol.* 2022, *13*, 1053107. [CrossRef]
- Liu, M.; Thijssen, S.; van Nostrum, C.F.; Hennink, W.E.; Garssen, J.; Willemsen, L.E.M. Inhibition of cow's milk allergy development in mice by oral delivery of beta-lactoglobulin-derived peptides loaded PLGA nanoparticles is associated with systemic whey-specific immune silencing. *Clin. Exp. Allergy* 2022, *52*, 137–148. [CrossRef]
- Palomares, F.; Ramos-Soriano, J.; Gomez, F.; Mascaraque, A.; Bogas, G.; Perkins, J.R.; Gonzalez, M.; Torres, M.J.; Diaz-Perales, A.; Rojo, J.; et al. Pru p 3-Glycodendropeptides Based on Mannoses Promote Changes in the Immunological Properties of Dendritic and T-Cells from LTP-Allergic Patients. *Mol. Nutr. Food Res.* 2019, 63, e1900553. [CrossRef]
- Palomares, F.; Gomez, F.; de la Fuente, M.C.; Perez-Sanchez, N.; Torres, M.J.; Mayorga, C.; Rojo, J.; Ramos-Soriano, J. Fucodendropeptides induce changes in cells of the immune system in food allergic patients via DC-SIGN receptor. *Carbohydr. Res.* 2022, 517, 108580. [CrossRef]
- Losada Mendez, J.; Palomares, F.; Gomez, F.; Ramirez-Lopez, P.; Ramos-Soriano, J.; Torres, M.J.; Mayorga, C.; Rojo, J. Immunomodulatory Response of Toll-like Receptor Ligand-Peptide Conjugates in Food Allergy. ACS Chem. Biol. 2021, 16, 2651–2664. [CrossRef] [PubMed]
- Nunez, R.; Rodriguez, M.J.; Palomares, F.; Gomez, F.; Jabato, F.M.; Cordoba-Caballero, J.; Seoane, P.; Losada, J.; Rojo, J.; Torres, M.J.; et al. Transcriptional changes in dendritic cells underlying allergen specific induced tolerance in a mouse model. *Sci. Rep.* 2022, 12, 2797. [CrossRef] [PubMed]
- Nunez, R.; Rodriguez, M.J.; Lebron-Martin, C.; Martin-Astorga, M.D.C.; Palomares, F.; Ramos-Soriano, J.; Rojo, J.; Torres, M.J.; Canas, J.A.; Mayorga, C. Methylation changes induced by a glycodendropeptide immunotherapy and associated to tolerance in mice. *Front. Immunol.* 2022, 13, 1094172. [CrossRef]
- Nunez, R.; Rodriguez, M.J.; Lebron-Martin, C.; Martin-Astorga, M.D.C.; Ramos-Soriano, J.; Rojo, J.; Torres, M.J.; Canas, J.A.; Mayorga, C. A synthetic glycodendropeptide induces methylation changes on regulatory T cells linked to tolerant responses in anaphylactic-mice. *Front. Immunol.* 2023, 14, 1165852. [CrossRef] [PubMed]
- 89. Yang, L.; Kulis, M. Hypoallergenic Proteins for the Treatment of Food Allergy. Curr. Allergy Asthma Rep. 2019, 19, 15. [CrossRef]

- Vandenplas, Y.; Brough, H.A.; Fiocchi, A.; Miqdady, M.; Munasir, Z.; Salvatore, S.; Thapar, N.; Venter, C.; Vieira, M.C.; Meyer, R. Current Guidelines and Future Strategies for the Management of Cow's Milk Allergy. J. Asthma Allergy 2021, 14, 1243–1256. [CrossRef]
- D'Auria, E.; Salvatore, S.; Acunzo, M.; Peroni, D.; Pendezza, E.; Di Profio, E.; Fiore, G.; Zuccotti, G.V.; Verduci, E. Hydrolysed Formulas in the Management of Cow's Milk Allergy: New Insights, Pitfalls and Tips. *Nutrients* 2021, 13, 2762. [CrossRef] [PubMed]
- 92. Fiocchi, A.; Barrio-Torres, J.; Dupont, C.; Howells, H.E.; Shamir, R.; Venter, C.; Meyer, R. Hydrolyzed rice formula for dietary management of infants with cow's milk allergy. *World Allergy Organ. J.* 2022, *15*, 100717. [CrossRef]
- Paparo, L.; Picariello, G.; Bruno, C.; Pisapia, L.; Canale, V.; Sarracino, A.; Nocerino, R.; Carucci, L.; Cosenza, L.; Cozzolino, T.; et al. Tolerogenic Effect Elicited by Protein Fraction Derived from Different Formulas for Dietary Treatment of Cow's Milk Allergy in Human Cells. *Front. Immunol.* 2020, *11*, 604075. [CrossRef]
- Cui, Y.; Yu, Y.; Lin, S.; Xu, L.; Li, L.; Cai, C.; Li, H. Alteration of Allergen Fold of Bos d 5 into a Hypoallergenic Vaccine for Immunotherapy of Cow's Milk Allergy. *Int. Arch. Allergy Immunol.* 2022, 183, 93–104. [CrossRef] [PubMed]
- Li, M.S.; Xia, F.; Liu, Q.M.; Chen, Y.Y.; Yun, X.; Liu, M.; Chen, G.X.; Wang, L.; Cao, M.J.; Liu, G.M. Hypoallergenic derivatives of Scylla paramamosain heat-stable allergens alleviated food allergy symptoms in Balb/c mice. *Food Funct.* 2022, 13, 11518–11531. [CrossRef]
- Zhang, Z.; Li, Z.; Lin, H. Reducing the Allergenicity of Shrimp Tropomyosin and Allergy Desensitization Based on Glycation Modification. J. Agric. Food Chem. 2021, 69, 14742–14750. [CrossRef]
- 97. Zhang, Z.; Li, X.M.; Li, Z.; Lin, H. Investigation of glycated shrimp tropomyosin as a hypoallergen for potential immunotherapy. *Food Funct.* **2021**, *12*, 2750–2759. [CrossRef]
- Jiang, S.; Wang, T.; Chen, K.; Wang, H.; Meng, X. Assessment of the effect of glycation on the allergenicity of sesame proteins. *Food Res. Int.* 2023, 168, 112771. [CrossRef]
- Yamada, Y.; Yokooji, T.; Kunimoto, K.; Inoguchi, K.; Ogino, R.; Taogoshi, T.; Morita, E.; Matsuo, H. Hypoallergenic Wheat Line (1BS-18H) Lacking omega5-Gliadin Induces Oral Tolerance to Wheat Gluten Proteins in a Rat Model of Wheat Allergy. *Foods* 2022, 11, 2181. [CrossRef]
- Kaeswurm, J.A.H.; Nestl, B.; Richter, S.M.; Emperle, M.; Buchweitz, M. Purification and Characterization of Recombinant Expressed Apple Allergen Mal d 1. *Methods Protoc.* 2020, *4*, 3. [CrossRef] [PubMed]
- Dantzer, J.A.; Wood, R.A. Omalizumab as an adjuvant in food allergen immunotherapy. *Curr. Opin. Allergy Clin. Immunol.* 2021, 21, 278–285. [CrossRef] [PubMed]
- 102. Fiocchi, A.; Artesani, M.C.; Riccardi, C.; Mennini, M.; Pecora, V.; Fierro, V.; Calandrelli, V.; Dahdah, L.; Valluzzi, R.L. Impact of Omalizumab on Food Allergy in Patients Treated for Asthma: A Real-Life Study. J. Allergy Clin. Immunol. Pract. 2019, 7, 1901–1909. [CrossRef] [PubMed]
- 103. Chinuki, Y.; Yagami, A.; Adachi, A.; Matsunaga, K.; Ugajin, T.; Yokozeki, H.; Hayashi, M.; Katayama, I.; Kohno, K.; Shiwaku, K.; et al. In vitro basophil activation is reduced by short-term omalizumab treatment in hydrolyzed wheat protein allergy. *Allergol. Int.* 2020, 69, 284–286. [CrossRef] [PubMed]
- 104. Badina, L.; Belluzzi, B.; Contorno, S.; Bossini, B.; Benelli, E.; Barbi, E.; Berti, I. Omalizumab effectiveness in patients with a previously failed oral immunotherapy for severe milk allergy. *Immun. Inflamm. Dis.* **2022**, *10*, 117–120. [CrossRef]
- 105. Andorf, S.; Purington, N.; Kumar, D.; Long, A.; O'Laughlin, K.L.; Sicherer, S.; Sampson, H.; Cianferoni, A.; Whitehorn, T.B.; Petroni, D.; et al. A Phase 2 Randomized Controlled Multisite Study Using Omalizumab-facilitated Rapid Desensitization to Test Continued vs Discontinued Dosing in Multifood Allergic Individuals. *EClinicalMedicine* 2019, 7, 27–38. [CrossRef] [PubMed]
- 106. Yee, C.S.K.; Albuhairi, S.; Noh, E.; El-Khoury, K.; Rezaei, S.; Abdel-Gadir, A.; Umetsu, D.T.; Burke-Roberts, E.; LeBovidge, J.; Schneider, L.; et al. Long-Term Outcome of Peanut Oral Immunotherapy Facilitated Initially by Omalizumab. *J. Allergy Clin. Immunol. Pract.* 2019, 7, 451–461. [CrossRef]
- Brandstrom, J.; Vetander, M.; Sundqvist, A.C.; Lilja, G.; Johansson, S.G.O.; Melen, E.; Sverremark-Ekstrom, E.; Nopp, A.; Nilsson, C. Individually dosed omalizumab facilitates peanut oral immunotherapy in peanut allergic adolescents. *Clin. Exp. Allergy* 2019, 49, 1328–1341. [CrossRef]
- 108. Azzano, P.; Paquin, M.; Langlois, A.; Morin, C.; Parizeault, G.; Lacombe-Barrios, J.; Samaan, K.; Graham, F.; Paradis, L.; Des Roches, A.; et al. Determinants of omalizumab dose-related efficacy in oral immunotherapy: Evidence from a cohort of 181 patients. J. Allergy Clin. Immunol. 2021, 147, 233–243. [CrossRef]
- Yang, H.; Qu, Y.; Gao, Y.; Sun, S.; Wu, R.; Wu, J. Research Progress on the Correlation between the Intestinal Microbiota and Food Allergy. *Foods* 2022, 11, 2913. [CrossRef]
- 110. Wu, Y.; Zhang, G.; Wang, Y.; Wei, X.; Liu, H.; Zhang, L.; Zhang, L. A Review on Maternal and Infant Microbiota and Their Implications for the Prevention and Treatment of Allergic Diseases. *Nutrients* **2023**, *15*, 2483. [CrossRef]
- 111. Zhang, J.; Su, H.; Li, Q.; Wu, H.; Liu, M.; Huang, J.; Zeng, M.; Zheng, Y.; Sun, X. Oral administration of Clostridium butyricum CGMCC0313-1 inhibits beta-lactoglobulin-induced intestinal anaphylaxis in a mouse model of food allergy. *Gut Pathog.* 2017, 9, 11. [CrossRef]
- Aoki-Yoshida, A.; Yamada, K.; Hachimura, S.; Sashihara, T.; Ikegami, S.; Shimizu, M.; Totsuka, M. Enhancement of Oral Tolerance Induction in DO11.10 Mice by Lactobacillus gasseri OLL2809 via Increase of Effector Regulatory T Cells. *PLoS ONE* 2016, 11, e0158643. [CrossRef]

- 113. Yang, B.; Xiao, L.; Liu, S.; Liu, X.; Luo, Y.; Ji, Q.; Yang, P.; Liu, Z. Exploration of the effect of probiotics supplementation on intestinal microbiota of food allergic mice. *Am. J. Transl. Res.* **2017**, *9*, 376–385.
- 114. Tian, X.; Liang, X.; He, H.; Cui, Q.; Liu, Q.; Fan, R.; Liu, T.; Yi, H.; Gong, P.; Wang, Q.; et al. Probiotics Alleviate Food Protein Allergy in Mice by Activating TLR4 Signaling Pathway. *Mol. Nutr. Food Res.* **2023**, *67*, e2200579. [CrossRef] [PubMed]
- Neau, E.; Delannoy, J.; Marion, C.; Cottart, C.H.; Labellie, C.; Holowacz, S.; Butel, M.J.; Kapel, N.; Waligora-Dupriet, A.J. Three Novel Candidate Probiotic Strains with Prophylactic Properties in a Murine Model of Cow's Milk Allergy. *Appl. Environ. Microbiol.* 2016, 82, 1722–1733. [CrossRef]
- 116. Miranda, V.C.; Souza, R.O.; Quintanilha, M.F.; Gallotti, B.; Assis, H.C.; Faria, A.M.C.; Nicoli, J.R.; Cara, D.C.; Martins, F.S. A Next-Generation Bacteria (Akkermansia muciniphila BAA-835) Presents Probiotic Potential Against Ovalbumin-Induced Food Allergy in Mice. *Probiotics Antimicrob. Proteins* 2023. [CrossRef] [PubMed]
- 117. Tang, M.L.; Ponsonby, A.L.; Orsini, F.; Tey, D.; Robinson, M.; Su, E.L.; Licciardi, P.; Burks, W.; Donath, S. Administration of a probiotic with peanut oral immunotherapy: A randomized trial. *J. Allergy Clin. Immunol.* 2015, 135, 737–744.e8. [CrossRef] [PubMed]
- 118. Dunn Galvin, A.; Lloyd, M.; Hsiao, K.C.; Tang, M.L.K. Long-term benefit of probiotic peanut oral immunotherapy on quality of life in a randomized trial. *J. Allergy Clin. Immunol. Pract.* **2021**, *9*, 4493–4495.E1. [CrossRef]
- 119. Loke, P.; Orsini, F.; Lozinsky, A.C.; Gold, M.; O'Sullivan, M.D.; Quinn, P.; Lloyd, M.; Ashley, S.E.; Pitkin, S.; Axelrad, C.; et al. Probiotic peanut oral immunotherapy versus oral immunotherapy and placebo in children with peanut allergy in Australia (PPOIT-003): A multicentre, randomised, phase 2b trial. *Lancet Child Adolesc. Health* 2022, 6, 171–184. [CrossRef]
- 120. Yamamoto-Hanada, K.; Sato, M.; Toyokuni, K.; Irahara, M.; Hiraide-Kotaki, E.; Harima-Mizusawa, N.; Morita, H.; Matsumoto, K.; Ohya, Y. Combination of heat-killed Lactiplantibacillus plantarum YIT 0132 (LP0132) and oral immunotherapy in cow's milk allergy: A randomised controlled trial. *Benef. Microbes* 2023, 14, 17–29. [CrossRef]
- 121. Fox, A.T.; Wopereis, H.; Van Ampting, M.T.J.; Oude Nijhuis, M.M.; Butt, A.M.; Peroni, D.G.; Vandenplas, Y.; Candy, D.C.A.; Shah, N.; West, C.E.; et al. A specific synbiotic-containing amino acid-based formula in dietary management of cow's milk allergy: A randomized controlled trial. *Clin. Transl. Allergy* **2019**, *9*, 5. [CrossRef]
- 122. Candy, D.C.A.; Van Ampting, M.T.J.; Oude Nijhuis, M.M.; Wopereis, H.; Butt, A.M.; Peroni, D.G.; Vandenplas, Y.; Fox, A.T.; Shah, N.; West, C.E.; et al. A synbiotic-containing amino-acid-based formula improves gut microbiota in non-IgE-mediated allergic infants. *Pediatr. Res.* 2018, *83*, 677–686. [CrossRef]
- 123. Hubbard, G.P.; Atwal, K.; Graham, L.; Narayanan, S.; Cooke, L.; Casewell, C.; Denton, S.A.; Gavin, J.; Browne, R.M.; Kinnear, F.J.; et al. Synbiotic containing extensively hydrolyzed formula improves gastrointestinal and atopic symptom severity, growth, caregiver quality of life, and hospital-related healthcare use in infants with cow's milk allergy. *Immun. Inflamm. Dis.* 2022, 10, e636. [CrossRef]
- 124. Wang, J.; Wood, R.A.; Raymond, S.; Suarez-Farinas, M.; Yang, N.; Sicherer, S.H.; Sampson, H.A.; Li, X.M. Double-Blind, Placebo-Controlled Study of E-B-FAHF-2 in Combination with Omalizumab-Facilitated Multiallergen Oral Immunotherapy. J. Allergy Clin. Immunol. Pract. 2023, 11, 2208–2216. [CrossRef] [PubMed]
- 125. Wang, Z.; Wang, Z.Z.; Geliebter, J.; Tiwari, R.; Li, X.M. Traditional Chinese medicine for food allergy and eczema. *Ann. Allergy Asthma Immunol.* **2021**, *126*, 639–654. [CrossRef] [PubMed]
- 126. Yang, N.; Maskey, A.R.; Srivastava, K.; Kim, M.; Wang, Z.; Musa, I.; Shi, Y.; Gong, Y.; Fidan, O.; Wang, J.; et al. Inhibition of pathologic immunoglobulin E in food allergy by EBF-2 and active compound berberine associated with immunometabolism regulation. *Front. Immunol.* 2023, 14, 1081121. [CrossRef]
- 127. Cao, T.W.; Xie, C.L.; Chen, C.Q.; He, Z.H.; Yan, Q.X.; Xu, G.; Yang, X.W. Anti-Food Allergic Alkaloids from the Lotus Seed Pot. *Chem. Biodivers.* **2021**, *18*, e2100770. [CrossRef]
- 128. Guo, Y.; Ma, Y.; Ma, L.; Guo, Z.; Xiao, Y.; Liu, Y.; Li, J.; Wang, S.; Liu, Y. Oleuropein Prevents OVA-Induced Food Allergy in Mice by Enhancing the Intestinal Epithelial Barrier and Remodeling the Intestinal Flora. *Mol. Nutr. Food Res.* 2022, 66, e2200455. [CrossRef] [PubMed]
- 129. Yamamoto, T.; Nagata, Y.; Hayashi, S.; Kadowaki, M. Isoflavones Suppress Cyp26b1 Expression in the Murine Colonic Lamina Propria. *Biol. Pharm. Bull.* 2020, 43, 1945–1949. [CrossRef]
- Vlieg-Boerstra, B.; Groetch, M.; Vassilopoulou, E.; Meyer, R.; Laitinen, K.; Swain, A.; Durban, R.; Benjamin, O.; Bottse, R.; Grimshaw, K.; et al. The immune-supportive diet in allergy management: A narrative review and proposal. *Allergy* 2023, 78, 1441–1458. [CrossRef]
- 131. Nagata, Y.; Yamamoto, T.; Kadowaki, M. Ginger Increases ALDH1A1 Expression and Enhances Retinoic Acid Signaling in a Human Colonic Epithelial Cell Line. J. Nutr. Sci. Vitaminol. 2020, 66, 462–467. [CrossRef] [PubMed]
- 132. Ma, Y.; Li, J.; Guo, Y.; Ma, L.; Liu, Y.; Kuang, H.; Han, B.; Xiao, Y.; Wang, Y. Dietary olive oil enhances the oral tolerance of the food allergen ovalbumin in mice by regulating intestinal microecological homeostasis. *J. Food Biochem.* **2022**, *46*, e14297. [CrossRef]
- 133. Patel, D.; Goruk, S.; Richard, C.; Field, C.J. Combined Supplementation with Arachidonic and Docosahexaenoic Acids in T Helper Type-2 Skewed Brown Norway Rat Offspring is Beneficial in the Induction of Oral Tolerance toward Ovalbumin and Immune System Development. J. Nutr. 2022, 152, 2165–2178. [CrossRef] [PubMed]
- Huynh, L.B.P.; Nguyen, N.N.; Fan, H.Y.; Huang, S.Y.; Huang, C.H.; Chen, Y.C. Maternal Omega-3 Supplementation during Pregnancy, but not Childhood Supplementation, Reduces the Risk of Food Allergy Diseases in Offspring. J. Allergy Clin. Immunol. Pract. 2023, in press. [CrossRef]

- 135. Deng, M.; Wu, X.; Duan, X.; Xu, J.; Yang, X.; Sheng, X.; Lou, P.; Shao, C.; Lv, C.; Yu, Z. Lactobacillus paracasei L9 improves colitis by expanding butyrate-producing bacteria that inhibit the IL-6/STAT3 signaling pathway. *Food Funct.* 2021, *12*, 10700–10713. [CrossRef] [PubMed]
- 136. Domingos-Lopes, M.F.P.; Lamosa, P.; Stanton, C.; Ross, R.P.; Silva, C.C.G. Isolation and characterization of an exopolysaccharideproducing Leuconostoc citreum strain from artisanal cheese. *Lett. Appl. Microbiol.* **2018**, *67*, 570–578. [CrossRef]

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