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## 1 Glycosidically Bound Aroma Compounds and Impact Odorants of

## 2 Four Strawberry Varieties

- 3 Ubeda, C<sup>1</sup>\*; San Juan, F<sup>2</sup>; Concejero, B<sup>2</sup>; Callejón, R. M<sup>1</sup>; Troncoso, A. M<sup>1</sup>; Morales,
- 4 M.  $L^1$ ; Ferreira,  $V^2$ ; Hernández-Orte,  $P^2$ .
- <sup>5</sup> <sup>1</sup>Área de Nutrición y Bromatología. Facultad de Farmacia. Universidad de Sevilla.
- 6 C/ P. García González nº 2, E- 41012. Sevilla. Spain.
- 7 <sup>2</sup>Laboratorio del Análisis del Aroma y Enología. Departamento de Química Analítica.
- 8 Facultad de Ciencias. Universidad de Zaragoza. 50009, Zaragoza, Spain.
- 9 \*Corresponding author: c\_ubeda@us.es; Tel.: 34-954-556761; Fax.: 34-954-233765

#### 26 Abstract

This paper reports the determination of glycosidically bound aroma compounds and the olfactometric analysis in four strawberry varieties (Fuentepina, Camarosa, Candonga and Sabrina). Different hydrolytic strategies were also studied. The results showed significant differences between acid and enzymatic hydrolysis. In general terms, the greater the duration of acid hydrolysis, the higher was the content of norisoprenoids, volatile phenols, benzenes, lactones, Furaneol, and mesifurane. A total of 51 aglycones were identified, 38 of them unreported in strawberry. Olfactometric analyses revealed that the odorants with higher modified frequencies were Furaneol,  $\gamma$ -decalactone, ethyl butanoate, ethyl hexanoate, ethyl 3-methylbutanoate, diacetyl, hexanoic acid, and (Z)-1,5-octadien-3-one. This last compound, described as geranium/green/pepper/lettuce (linear retention index = 1378), was identified for the first time. Differences with regard to fruity, sweet, floral, and green aroma characters were observed among varieties. In Candonga and Fuentepina, the green character overpowered the sweet. In the other two strawberry varieties sweet attributes were stronger than the rest. Keywords: Glycosides, strawberry, aroma, flavor, olfactometry. 

### 51 **INTRODUCTION**

52 Strawberry is a much appreciated fruit due to its aroma, taste and health properties. It is 53 usually consumed fresh (75% of total production) but is also used in the food industry 54 as an important ingredient in jam, yogurt, syrup, tea, juice, ice cream, and other food 55 products (25% of overall production)<sup>1</sup>. Aroma is one of the most valued attributes of 56 strawberry. The aroma of this fruit includes volatile compounds, both in their free form, 57 and as non-volatile compounds, present mainly as glycoconjugates formed by a sugar 58 and an aglycone.

There are numerous studies concerning free volatile compounds of strawberry, with more than 360 volatile flavor compounds<sup>2</sup> identified. In order to learn more about the volatile composition of strawberry, several olfactometric studies have been undertaken using gas chromatography-olfactometry (GC-O)<sup>3-5</sup>.

Non-volatile compounds are, moreover, potential natural sources of aroma because the hydrolysis of the bonds between the sugar and the aglycone turns this molecule into an aromatic compound. As ripening proceeds, the increase in these soluble sugars results in an increase in the availability of precursors capable of producing aroma compounds<sup>6</sup>.

These non-volatile compounds have been extensively studied in grapes<sup>7–9</sup> and in other 67 fruits such as lychee, acerola, blackberry, pineapple and mango $^{10-14}$ , among others. 68 69 Strawberry precursors have hardly been studied. After the description of the presence of 2,5-dimethyl-4-hydroxy-2H-furan-3-one  $\beta$ -D-glucopyranoside in strawberry<sup>15</sup>, Wintoch 70 et al.<sup>16</sup> analyzed the glycosidical aroma compounds from two strawberry species using 71 72 Amberlite XAD-2. Other research groups have focused their studies on one aglycone, furaneol (2,5-dimethyl-4-hydroxy-2H-furan-3-one)<sup>17</sup>, due to its high influence on the 73 74 overall flavor. In addition, there have been some studies concerning the evolution of 75 these non-aromatic precursors during ripening. These studies show an increase in their

aglycones during the above mentioned stage<sup>18</sup>. Knowledge of the strawberry aromatic 76 77 precursors is important because it enables us to predict the final aroma of new 78 strawberry-based products. As a result, there are several different groups studying the production process of strawberry fermentation products<sup>19,20</sup>. Such of analyses would 79 80 enable us to estimate the aromatic potential and therefore select the best raw material. 81 The aim of this study was to determine the aromatic potential of different strawberry 82 varieties with the aim of selecting the most suitable varieties for producing several 83 fermented strawberry-based food products. Therefore the aroma compounds released by 84 acid hydrolysis of glycosidic precursors isolated from 4 different varieties have been 85 determined. Free aromas were also studied by GC-O analyses to determine the most 86 important compounds, from a sensory point of view, in these varieties.

### 87 MATERIALS AND METHODS

#### 88 **Reagents and standards**

89 Dichloromethane, ethanol and methanol were supplied by Merck (Darmstadt, 90 Germany), ethyl acetate and sodium fluoride by Fluka (Buchs, Switzerland). Sodium 91 dihydrogen-phosphate 1-hydrate, L (+) ascorbic and citric acids were purchased from 92 Panreac (Barcelona, Spain). Pure water was obtained from a Milli-Q purification system 93 (Millipore, U.S.). LiChrolut EN resins were purchased from Merck. An alkane solution 94 (C8–C28), 20 mg/L in dichloromethane, was used to calculate the linear retention index 95 (LRI) of each analyte. The chemical standards used for the identification and 96 quantification of volatile compounds were as follows: (Z)-rose oxide, linalool,  $\alpha$ -97 terpineol, nerol, geraniol, benzaldehyde, β-phenylethanol, 2-phenoxyethanol were 98 purchased from Fluka (Buchs, Switzerland). β-ionone was sourced from Sigma (St. 99 Louis, MO) and guaiacol, m-cresol, eugenol, 4-ethylphenol, 4-vinylguaiacol, methyl 100 vanillate, acetovanillone, zingerone, homovanillyl alcohol, phenylacetaldehyde, benzyl 101 alcohol, ethyl cinnamate,  $\gamma$ -nonalactone,  $\gamma$ -decalactone, (Z)-3-hexen-1-ol from Aldrich 102 (Gillingham, UK). (E)-isoeugenol, 4-vinylphenol,  $\delta$ -octalactone,  $\delta$ -decalactone were 103 purchased from Lancaster (Strasbourg, France). Finally,  $\beta$ -damascenone and vanillin 104 were supplied by Firmenich (Geneva, Switzerland) and Panreac (Barcelona, Spain) 105 respectively.

106 Samples

We employed freshly purchased *Fragaria ananassa, var. camarosa* strawberries in
order to optimize the extraction method and to obtain the aroma precursors extract.
Aromatic precursors were then determined in four different varieties of strawberry: *Fuentepina, Camarosa, Candonga* and *Sabrina*. These strawberries were also employed
for the olfactometric studies.

## 112 Extraction of aroma precursors

113 In order to study the effects of different kinds of hydrolysis we prepared a precursors 114 pool from strawberries of Camarosa variety adquired in the market. The preparation procedure was based on Ibarz et al.<sup>7</sup>. We used an Ultra Turrax T25 Basic mixer (Ika, 115 116 Labortechnik) to crush and homogenize 2 kg of strawberries with 1 L of cold Milli-Q 117 water in the presence of 0.13 M NaF, to prevent microbial growth, and 50 mg/L of 118 ascorbic acid (as an antioxidant). This mixture was then centrifuged and filtered 119 obtaining a strawberry must which was placed in PYREX flasks, adding 2 gr of 120 LiChrolut resins (previously pre-conditioned with dichloromethane, methanol and Milli-121 Q water) per kg of strawberry. The oxygen of the flasks was evacuated using nitrogen. 122 We left the must in contact with the resins for 16h in a Heidolph PROMAX 1020 shaker 123 (Schwabach, Germany) at 90 rpm. We packaged the resin and each cartridge of 500 mg 124 was washed with 50 mL of water. It was then completely vacuum-dried and free aromas 125 were extracted with 50 mL of dichloromethane and discarded. 30 mL of an ethyl

acetate/methanol solution (9:1) were subsequently percolated through the resin. The
solvents were evaporated under vacuum, resuspended in a 50:50 ethanol/water solution
and kept at -20°C.

129 To analyze the 4 different strawberry varieties, we followed the same technique as that 130 utilized for obtaining the precursors pool. In this case we processed 10 gr of strawberry 131 since we obtained the best results in previous studies using that quantity (data not 132 shown). The must was percolated through a 200 mg LiChrolut EN cartridge (previously 133 pre-conditioned with 10 mL dichloromethane, 10 mL methanol and 10 ml Milli-Q 134 water). After that, the column was washed with 20 mL of Milli-Q water and then was 135 completely dried. In order to eliminate all free aromatic compounds, we passed 20 mL 136 of dichloromethane through the cartridge. To recover the precursors from the resin we 137 employed 20 mL of a solution of ethyl acetate/methanol (9:1). This eluate was 138 concentrated to 1 mL under vacuum at 40 °C and then, taken to dryness under a gentle 139 nitrogen stream. Each sample was extracted in duplicate.

### 140 Acid and enzymatic hydrolysis

141 Different hydrolytic conditions were performed in order to study their influence on the 142 aromatic profile of strawberry using the precursors pool previously obtained. The acid 143 hydrolyses assayed were: 15 min, 1 h and 4 h at 100 °C and one week at 45 °C. For this 144 hydrolysis we mixed 8 mL of citric buffer (0.2 M pH 2.5), 1 mL of the precursor extract 145 and 1 mL of an ethanol/water solution (50:50) (to maintain the same concentration of 146 ethanol in all the acid hydrolysis assays in a 20 mL vial. After this, the vial was sealed 147 and placed in the oven. Moreover, an enzymatic hydrolysis was performed during 16 h 148 at 38 °C. In this case we used 8.7 mL of citrate (0.1 M)/ phosphate (0.2 M) buffer 149 solution at pH=5, 1 mL of the precursor extract, which was subjected to vacuum to

remove the ethanol, and 800 μL of a pectinase enzyme solution with 200 mg/mL of AR
2000.

Otherwise, for the analysis of the four varieties of strawberry, the dry extract was reconstituted in 10 mL of citric buffer (0.2 M pH 2.5, 10% EtOH) and was subjected to hydrolysis at 100 °C for one hour. Before any hydrolysis was undertaken, the remaining oxygen was displaced from the vial with nitrogen in order to prevent oxidation of the compounds during the process. Each hydrolysis was done in duplicate.

#### 157 Extraction of volatiles released in the hydrolysis

After the hydrolysis, the solution was percolated through a 50 mg LiChrolut EN cartridge (previously pre-conditioned with 6 mL dichloromethane, 2 mL methanol and 2 mL of citric buffer solution). Then was washed with 1 mL Milli–Q water and dried. To elute the aromatic compounds, 700  $\mu$ L of dichloromethane were passed through the column and collected in a Kuderna Danish Supelco (Bellefonte, PA, USA), adding 14  $\mu$ L of the internal standard 4-methyl-2-pentanol (402.6  $\mu$ g/g). Finally was concentrated to 100  $\mu$ L with a gentle nitrogen stream.

### 165 **Preparation of the olfactometry extract**

166 To obtain a representative extract of each strawberry variety for the olfactometry analyses we followed the method used by Ferreira et al.<sup>21</sup>. 80 g of the fruit were crushed 167 and placed in a purge and trap system<sup>22</sup>. A Lichrolut EN cartridge was placed on the top 168 169 of the bubbler flask. A nitrogen stream of 500 mL/min was applied to the sample for 170 100 min, releasing the free volatile compounds of strawberry in the headspace being 171 trapped by the cartridge. Finally these compounds were eluted with 3.2 mL of 172 dichloromethane containing 5% methanol. The extract was concentrated to a final 173 volume of 200 µL.

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### 175 GC-MS and GC-O analytical conditions

176 Gas chromatographic analysis of the volatiles released in the hydrolysis was performed 177 with a CP-3800 chromatograph coupled to a Saturn 2200 ion trap mass spectrometric 178 detection system from Varian (Sunnyvale, CA, USA). A DB-WAXetr capillary column 179 (J&W Scientific, Folsom, CA, USA) (60 m  $\times$  0.25 mm I.D., film thickness 0.5 µm) 180 preceded by a 3 m  $\times$  0.25 mm uncoated (deactivated, intermediate polarity) precolumn 181 from Supelco (Bellefonte, PA, USA) was used. Helium was the carrier gas at a flow rate 182 of 1 mL/min. The oven temperature program was 3 min at 40 °C, 10 °C/min up to 90 183 °C, 2 °C/min up to 230 °C and finally held at this temperature for 37 min. Initially the 184 injector was kept at 35 °C for 0.3 min and a pressure pulse of 25 psi for 2.60 min was 185 applied. The injector was then heated to 250 °C at rate of 200 °C/min. The splitless time 186 was 2.60 min. The injection volume was 4  $\mu$ L. The global run time was recorded in full 187 scan mode (40-220 m/z mass range). The chromatographic data were analyzed by 188 Varian Saturn GC–MS Version 6.3 software<sup>23</sup>.

189 To carry out the olfactometric analyses we followed the protocol described in Ferreira et 190 al., (2009). The sensory panel was composed of six expert sniffers. Each strawberry 191 extract was smelled once a day by each panelist. Sniffing time was approximately 30 192 min. The experiments were carried out in a Thermo 8000 series GC equipped with a 193 flame ionization detector (FID) and a sniffing port (ODO-1 from SGE) connected by a 194 flow splitter to the column exit. The chromatographic conditions were the same as 195 described in Campo et al., (2005). Tasters were asked to score the intensity of each 196 aromatic stimulus using a 4-point scale (0=not detected, 1=weak, 2=clear but not 197 intense note, 3=intense note). Results were expressed as "modified frequency" (MF). calculated with the formula proposed by Dravnieks<sup>24</sup>. The identification of the odorants 198

199 was done by comparison of their odors, chromatographic retention index and MS200 spectra with those of pure reference compounds.

#### 201 Data treatment

Analysis of variance (ANOVA) was performed using Statistica (version 7.0) software
package (Statsoft, Tulsa, USA). Principal component analysis (PCA) was carried out
using Unscramble vs. 9.7 from Camo (Norway).

#### 205 **RESULTS AND DISCUSSION**

#### 206 Influence of type of hydrolysis

207 In general, the concentrations of the released compounds were very different depending 208 on the type of hydrolysis (Table 1 and Figure 1). Principal component analysis (PCA) 209 was performed in order to observe which conditions were related to the release of the 210 different compounds. As can be seen in Figure 1, PC1 which explains 47% of the 211 variance, clearly separates the acid hydrolyses from the enzymatic ones. Also, PC2 212 (35% of the variance) groups the samples in function of time. As the time of hydrolysis 213 increased, the concentration of norisoprenoids, volatile phenols, benzenes and lactones 214 was higher. The behavior of terpenes was heterogeneous. The amounts of  $\alpha$ -terpinolene, 215 (Z)-rose oxide and neric acid increased during the harsh hydrolysis, reaching the highest 216 amount after four hours of the hydrolytic assay. However, the remaining terpenes 217 reached their maximum concentration between 15 min. and 1 h of hydrolysis. In the 218 case of vanillin derivatives, each compound followed a different trend. With respect to 219 the miscellaneous group, it is important to mention the cases of furaneol and cinnamic 220 acid, which increased during hydrolysis, reaching their maximum after 4 h.

221 Results after leaving the precursors pool one week at 45 °C in citric buffer did not show 222 great differences over the aforementioned hydrolysis. However, hydrolysates from the 223 enzymatic assay were very rich in linalool,  $3-\infty -\alpha$ -ionol and some volatile phenols

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224 such as eugenol, 4-vinylguaiacol and 4-vinylphenol. Vanillin derivatives were also 225 released more effectively. Moreover, this hydrolysis resulted in an extract with high 226 amounts of benzyl alcohol and  $\beta$ -phenylethanol. With regard to furaneol there were no 227 significant differences between 1 h of acid or enzymatic hydrolysis. On the other hand, when applying harsh acid hydrolysis the release of terpenes with the exception of 228 linalool, was greater. These results are in accordance with previous studies<sup>8</sup> in which 229 230 different hydrolytic strategies have been compared. The enzymatic hydrolysis was much 231 more efficient for releasing volatile phenols, vanillin derivatives and benzenes such as 232  $\beta$ -phenylethanol and benzyl alcohol than acid hydrolysis.

Despite these results, we decided to apply acid hydrolysis to perform the assays in each strawberry extract due to its similarity with alcoholic fermentation<sup>8</sup>. This was done in order to compare the results with a hypothetical strawberry fermentation. The time period chosen was one hour as a compromise between compounds which are degraded after 4 h and those which are not formed earlier than this.

## 238 Study of the aglycones released from hydrolysis of four strawberry varieties

Taking into account the results obtained after testing the selected strategies, 1 h of harsh
acid hydrolysis was applied for the analysis of minor aromatic compounds released
from non-volatile precursors of the four strawberry varieties.

As can be observed in Table 2, within the analyzed varieties, *Fuentepina* (Figure 2) proved to have the highest quantity of aromatic compounds present as precursors. After this, *Camarosa* and *Sabrina* varieties presented high levels, the *Candonga* variety being the poorest in these non-aromatic molecules.

In general, among the aglycones quantified, the major ones were linalool,  $\alpha$ -terpineol, geraniol, 4-vinylguaiacol, 4-vinylphenol, benzyl alcohol, benzoic acid,  $\gamma$ -decalactone and cinnamic acid. The presence of 4-vinylphenol in strawberries, especially in 249 *Candonga* variety, is remarkable because reached values between 0.9-9.6 mg/kg of 250 fruit. This is in agreement with the results obtained by Groyne<sup>18</sup>, which observed a great 251 amount and variability of this compound related to the strawberry variety.

The Sabrina variety was characterized by high amounts of terpenes, presenting discrete values for the rest of the aglycones with respect to the other varieties tested.

One of the most important components of strawberry flavor is 2,5-dimethyl-4-hydroxy-2H-furan-3-one (furaneol)<sup>6</sup>, responsible of the sweet, caramel, burnt sugar notes at high concentrations and fruity at lower concentrations. This compound reached the highest levels in *Camarosa* variety. Another important compound of this fruit is mesifurane, which is described with similar descriptors. In this case, *Sabrina* accounted the highest levels of mesifurane as glycosidically bound aroma form.

260 Finally, it is important to remark that XAD-2 Amberlite was the adsorbent employed for 261 the determination of strawberry aromatic precursors in previously published works. In 262 this work we tested the effectiveness of LiChrolut EN cartridges. This resin has been 263 demonstrated as being more efficient that the Amberlite used in previous works by other 264 authors. We identified a total of 51 aglycones with LiChrolut EN resins, 38 of which 265 had previously not been reported in strawberry. Knowing the aromatic potential of the 266 strawberries gives us an idea of the overall final aroma of a product made from this fruit 267 and therefore we could select the best variety as starting substrate.

#### 268 Odor active compounds determined using GC-O

We performed olfactometric analyses of the free aroma compounds of 4 varieties of strawberry. This extraction technique enables us to obtain a more representative extract than other techniques and therefore it provides a more realistic idea of the overall sample flavor. Thirty-four important odor zones were perceived in the headspace extract. Table 3 shows the modified frequency (MF) of all the perceived odorants, only those with MF higher than 30 in at least one sample (odor active compounds) beingincluded. Among these perceived aromatic zones, 6 were not identified.

276 Within the odor zones which had the greatest impact in the majority of the strawberry 277 varieties, we identified furaneol,  $\gamma$ -decalactone, ethyl butanoate, ethyl hexanoate, ethyl-3-methylbutanoate, diacetyl and hexanoic acid, in agreement with other studies<sup>25,3,4</sup>. 278 279 These compounds, therefore, seem to be responsible for the overall impact aroma of 280 strawberries. They provide caramel-like, fruity, buttery and sour notes. Furthermore, 281 other odor zones with high MF were perceived in most of the varieties, with unpleasant 282 notes such as cheese/feet/sweat/milk or burnt hair. We identified them as isovaleric acid 283 and 2-acetylpyrazine. Panelists also perceived an odor zone described as 284 geranium/green/pepper/lettuce (LRI=1378) with an MF higher than 80 in Fuentepina 285 and Candonga varieties, identified as (Z)-1,5-octadien-3-one. This odor zone had been 286 observed by other authors but, to our knowledge, it had not been identified. There are 287 some odor zones that clearly differ one variety from the others. This is the case of the 288 floral/sweet/strawberry (LRI=1346) and floral/lemon (LRI=1563) notes identified as 289 (Z)-rose oxide and (R/S)-linalool and which are only present in *Fuentepina* strawberry. 290 In the *Candonga* variety tasters perceived a tropical/pineapple/citrus/green (LRI=1380) 291 odor zone with a high MF (61), tentatively identified as methyloctanoate, which was not 292 perceived in the other strawberries.

As expected, furaneol reached a high MF ( $\geq$ 80) but mesifurane MF values hovered at 33-45. These compounds, like the rest of the aglycones, are released during the fruit ripening stage, their presence increasing as a free form in ripe strawberry<sup>18</sup>. So, depending on the fruit developmental stage, different aglycones will appear. This explains why some data from the precursors analysis (Table 2) does not match with the olfactometric results. (Z)-Rose oxide is only present as a precursor in the *Fuentepina*  299 variety and was only perceived in this variety during the olfactometric analysis. 300 Additionally, panelists perceived the peach/sweet/strawberry (LRI=2170) odor zone 301 identified as  $\gamma$ -decalactone, with a very high MF ( $\geq 80$ ) in all varieties except for 302 Camarosa. This odor zone reached a low MF (26), a similar situation occurring in the 303 precursors determinations. However, the results obtained in olfactometric and 304 precursors assays for linalool and β-damascenone do not match. As mentioned above, 305 this confirms the staggered release of the aglycones. In conclusion we could say that 306 there were some odor zones that clearly differ among varieties, being only present in 307 one of the varieties.

308 We used spider webs to have a general visual comparison of the four strawberry 309 varieties considering fruity, sweet, floral and green aroma characters (Figure 3). For that 310 purpose, we added the MF of the odor zones of each character type of every strawberry 311 (divided by 10), and then divided by the total of odor zones found of that character 312 during the olfactometric analysis. Differences can be observed among the different 313 strawberry varieties. The Camarosa variety was the least aromatic one since their 314 aromatic zones reached the lowest MF. Green character predominates over sweet in 315 Fuentepina and Candonga; however in the other two varieties the sweet character is 316 stronger than the other attributes.

317 In the case of *Candonga*, the figure shows that the floral character is almost 318 imperceptible compared to the fruity character, which is very high.

In summary, the results suggest that this method is suitable for the determination of glycosidically bound aroma compounds of strawberry. There were several significant differences among varieties with respect to the content in precursors, *Fuentepina* being the variety which had the highest quantity of aromatic compounds present as precursors. A total of 38 aglycones have been described for the first time in strawberry.

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In general, the key odorants were furaneol,  $\gamma$ -decalactone, ethyl butanoate, ethyl hexanoate, ethyl-3-methylbutanoate, diacetyl and hexanoic acid. In addition, we could state that the presence of some odor zones clearly differ among varieties. On the other hand, if we consider fruity, sweet, floral and green aroma characters the overall aroma, of *Fuentepina* and *Candonga* varieties presented mainly green notes, however in the case of *Camarosa* and *Sabrina* varieties the aromatic notes were mainly sweet.

## 330 Abbreviations Used

- 331 Linear retention index (LRI)
- 332 Flame ionization detector (FID)
- 333 Modified frequency" (MF),
- 334 Principal component analysis (PCA)
- 335 Analysis of variance (ANOVA)
- 336 Gas chromatography olfactometry (GC-O)
- 337 Gas chromatography mass spectrometry (GC-MS)
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- 417 **Footnote:** This research was made possible through the financial support from the 418 Spanish Government by means of a predoctoral grant BES-2008-003116 and the 419 research project AGL2007-66417-C02-01 funded by the Ministry of Science and 420 Innovation.
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## 424 FIGURE CAPTIONS

- **Figure 1.** Principal component plot showing the scores for the samples of acid and
- 426 enzymatic hydrolysis.
- **Figure 2.** GC-MS Chromatographic profile of the strawberry variety *Fuentepina*.
- **Figure 3.** Sensory profile plot of Fuentepina, Camarosa, Candonga and Sabrina
- 429 varieties considering fruity, sweet, green and floral characteres.

<b>Table 1.</b> Concentration ( $\mu g/kg$ of Strawberries Except Where Indicated <sup>x</sup> ) of Volatil
Compounds Released After Harsh Acid and Enzymatic Hydrolysis of the Strawberry
Precursors Pool.

	0 min	15 min	1 h	4 h	1 week 45 °C	Enzymatic	
Terpenes							
α-terpinolene	nd	0.62±0.10 <sup>a</sup>	2.94±0.36 <sup>b</sup>	4.51±0.14 °	0.27±0.01 <sup>d</sup>	$1.17 \pm 0.09^{\text{ f}}$	
(Z)-rose oxide	nd	nd	0.02±0.00 <sup>a</sup>	0.25±0.01 b	nd	nd	
(R/S)-linalool	nd	75±2 <sup>a</sup>	3.50±0.30 <sup>b</sup>	nd	5.07±0.13 <sup>c</sup>	105±2 <sup>d</sup>	
α-terpineol	nd	27±1 <sup>a</sup>	111±13 <sup>b</sup>	50±1 °	77±2 <sup>d</sup>	1.28±0.14 e	
nerol	nd	6.20±0.79 <sup>a</sup>	12.72±0.39 <sup>b</sup>	nd	nd	2.18±0.22 <sup>c</sup>	
geraniol	4.46±0.36 <sup>a</sup>	29±1 <sup>b</sup>	4.91±0.63 <sup>a</sup>	nd	3.74±0.35 <sup>a</sup>	5.95±0.56 <sup>a</sup>	
farnesol	nd	12±1	nd	nd	nd	nd	
linalool acetate	nd	0.23±0.04	nd	nd	nd	nd	
terpinen-4-ol <sup>x</sup>	nd	nd	3.01±0.27 <sup>a</sup>	2.84±0.09 <sup>a</sup>	$0.68 \pm 0.04$ <sup>b</sup>	nd	
δ-terpineol <sup>x</sup>	nd	nd	6.74±0.41 <sup>a</sup>	6.05±0.38 <sup>a</sup>	3.93±0.05 <sup>b</sup>	nd	
neric acid	nd	nd	0.20±0.02 <sup>a</sup>	0.50±0.04 <sup>b</sup>	nd	1.01±0.03 °	
Norisoprenoids							
β-damascenone	nd	0.46±0.02 <sup>a</sup>	1.30±0.11 b	2.28±0.08 °	0.59±0.02 <sup>d</sup>	nd	
β-ionone	0.15±0.01 <sup>a</sup>	nd	nd	nd	nd	0.25±0.01 b	
1,1,6-trimethyl-1,2- dihydronaphthalene(TDN) <sup>x</sup>	nd	0.28±0.01 <sup>a</sup>	1.68±0.01 <sup>b</sup>	2.77±0.09 °	0.59±0.02 <sup>d</sup>	1.11±0.01 e	
tert-1-(2,3,6-trimethylphenyl) buta-1,3-diene (TPB) <sup>x</sup>	nd	0.34±0.01 <sup>a</sup>	6.35±0.09 <sup>b</sup>	7.51±0.44 <sup>b</sup>	2.13±0.04 °	$0.50 \pm 0.00^{\ d}$	
3-oxo-β-ionone <sup>x</sup>	nd	1.33±0.03 a	4.51±0.42 <sup>b</sup>	4.65±0.21 <sup>b</sup>	$2.77 \pm 0.14$ <sup>c</sup>	nd	
actinidols <sup>x</sup>	nd	0.24±0.02 <sup>a</sup>	5.81±0.52 <sup>b</sup>	6.62±0.23 <sup>b</sup>	4.09±0.15 °	0.25±0.02 <sup>a</sup>	
norisoprenoid 1 x	nd	nd	2.81±0.22 <sup>a</sup>	4.15±0.10 <sup>b</sup>	0.27±0.03 °	nd	
3-oxo-α-ionol	nd	nd	$0.63 \pm 0.07^{a}$	nd	nd	75±2 <sup>b</sup>	
Volatile phenols							
guaiacol	nd	nd	nd	0.70±0.09 <sup>a</sup>	nd	$0.91 \pm 0.04^{a}$	
m-cresol	nd	nd	nd	nd	nd	0.22±0.01	
eugenol	$0.62{\pm}0.01$ <sup>a</sup>	$0.70{\pm}0.08^{a}$	1.35±0.17 <sup>b</sup>	6.06±0.86 °	1.07±0.02 <sup>b</sup>	18±1 <sup>d</sup>	
4-ethylphenol	nd	$0.08 {\pm} 0.00$ <sup>a</sup>	nd	nd	nd	1.11±0.14 <sup>b</sup>	
4-vinylguaiacol	4.38±0.08 a	5.73±0.79 <sup>a</sup>	116±11 <sup>b</sup>	151±14 <sup>b</sup>	35±2 °	352±8 <sup>d</sup>	
(E)-isoeugenol	$1.79 \pm 0.09^{a}$	$1.38{\pm}0.07$ <sup>a</sup>	0.91±0.14 <sup>b,c</sup>	1.33±0.05 <sup>a,b</sup>	$0.68 \pm 0.01$ <sup>c</sup>	3.70±0.57 <sup>d</sup>	
4-vinylphenol	121±2 <sup>a</sup>	247±16 <sup>b</sup>	12606±1440 °	20904±3263 <sub>c,e</sub>	6231±120 <sup>d</sup>	27863±2764 <sup>e</sup>	
Vanillin derivatives							
vanillin	$0.50\pm0.01^{a}$	$1.08\pm0.01$ <sup>b</sup>	2.22±0.23 °	3.81±0.10	2.36±0.06	8.21±0.68	
methyl vanillate	0.10±0.00 <sup>a</sup>	nd	nd	nd	nd	1.16±0.09 <sup>b</sup>	
acetovanillone	nd	nd	nd	0.56±0.09 <sup>a</sup>	nd	2.19±0.00 <sup>b</sup>	
homovanillyl alcohol	nd	nd	nd	nd	$1.18\pm0.00$	nd	
homovanillinic acid <sup>x</sup>	5.21±0.10 <sup>a</sup>	4.10±0.24 <sup>b</sup>	nd	nd	nd	83±4 °	
Benzenes							
benzaldehyde	$0.74{\pm}0.06$ <sup>a</sup>	$1.86 \pm 0.01$ <sup>b</sup>	3.69±0.35 °	8.13±0.68 <sup>d</sup>	3.07±0.12 °	11±1 <sup>e</sup>	
phenylacetaldehyde	$0.67 \pm 0.01$ <sup>a</sup>	$0.87{\pm}0.10^{a}$	3.24±0.19 <sup>b</sup>	4.35±0.27 °	nd	4.46±0.43 °	
benzyl alcohol	1.69±0.21 <sup>a</sup>	3.14±0.37 <sup>b</sup>	21±1 °	59±1 <sup>d</sup>	10±1 <sup>e</sup>	1361±40 <sup>f</sup>	
β-phenylethanol	nd	$1.90{\pm}0.08$ <sup>a</sup>	4.54±0.05 <sup>b</sup>	9.61±0.27 °	3.21±0.19 <sup>d</sup>	9 <sup>d</sup> 97±4 <sup>e</sup>	
ethyl cinamate	nd	nd	7.09±0.09 <sup>a</sup>	23±1 <sup>b</sup>	18±1 °	3.21±0.19 <sup>d</sup>	
2-phenoxyethanol	1.03±0.04 a	1.38±0.24 a,c	0.64±0.11 <sup>b</sup>	0.96±0.01 <sup>a</sup>	0.54±0.06 <sup>b</sup>	1.85±0.14 °	
benzoic acid	7.10±0.93 <sup>a</sup>	10±1 <sup>b</sup>	113±17 <sup>c</sup>	210±19 <sup>d</sup>	44±3 <sup>e</sup>	240±7 <sup>d</sup>	
dihydromethyl-eugenol x	nd	nd	0.20±0.03 <sup>a</sup>	0.41±0.01 b	0.18±0.01 <sup>a</sup>	3.20±0.17 °	
Lactones							

δ-octalactone <sup>y</sup>	nd	$0.47 {\pm} 0.01$ <sup>a</sup>	1.09±0.01 b,c	1.47±0.01 <sup>b</sup>	1.08±0.00 °	nd
γ-nonalactone <sup>y</sup>	nd	0.68±0.01 <sup>a</sup>	nd	nd	nd	$0.86 \pm 0.00^{b}$
$\gamma$ -decalactone <sup>y</sup>	nd	$0.10{\pm}0.01$ <sup>a</sup>	7.54±0.00 <sup>b</sup>	17±1 °	nd	1.08±0.03 <sup>d</sup>
pantolactone	2.49±0.03 <sup>a</sup>	1.18±0.11 <sup>b</sup>	6.49±0.49 °	$8.47{\pm}0.98$ <sup>c</sup>	nd	nd
Miscellaneous						
(Z)-3-hexen-1-ol	nd	1.16±0.01 <sup>a</sup>	2.03±0.01 b	2.19±0.09 <sup>b</sup>	2.17±0.12 <sup>b</sup>	13±1 °
(E)-2-hexen-1-ol	4.37±0.52 <sup>a</sup>	4.95±0.52 <sup>a</sup>	2.87±0.15 <sup>b</sup>	2.90±0.10 <sup>b</sup>	3.00±0.17 <sup>b</sup>	5.80±0.17 <sup>c</sup>
ethyl decanoate	4.34±0.05 <sup>a</sup>	4.38±0.02 <sup>a</sup>	4.36±0.02 a	nd	4.28±0.00 <sup>a</sup>	nd
2-ethylhexanoic acid	1.30±0.22 <sup>a</sup>	1.20±0.06 <sup>a</sup>	1.13±0.01 <sup>a</sup>	1.07±0.01 <sup>a</sup>	1.87±0.22 <sup>a,b</sup>	2.19±0.09 <sup>b</sup>
4-methoxy-2,5-dimethyl- 3(2H)furanone (mesifurane) <sup>x</sup>	50±5 <sup>a</sup>	338±4 <sup>b</sup>	339±19 <sup>b</sup>	315±23 °	$251\pm6^{d}$	307±9°
4-hydroxy-2,5-dimethyl- 3(2H)furanone (furaneol) <sup>x</sup>	23±1 <sup>a</sup>	58±2 <sup>b</sup>	74±1 °	102±3 <sup>d</sup>	60±1 <sup>b</sup>	74±3 °
cinnamic acid <sup>x</sup>	338±18 <sup>a</sup>	586±61 <sup>b</sup>	2917±226 °	7657±555 <sup>d</sup>	1828±80 <sup>e</sup>	6209±119 <sup>d</sup>

nd: non detected.
<sup>x</sup> Chemical standard not available. Tentatively identified. Data are relative areas (to 4-Methyl-2-pentanol x 1000).
<sup>y</sup> Data are the relative areas (to 4-Methyl-2-pentanol x 1000). Chemical standard available,but the degradation of the products did not allow

quantification. Concentrations of the same compound with different letter show significant differences (p < 0.05).

**Table 2**. Concentration ( $\mu$ g/kg of Strawberries Except Where Indicated <sup>x</sup>) of Volatile Compounds Released After Harsh Acid Hydrolysis of the Precursor Extract from Each Strawberry Variety.

	Fuentepina	Camarosa	Candonga	Sabrina	
Terpenes	_				
α-terpinolene (1)	$0.58{\pm}0.05^{a}$	0.39±0.01 <sup>b</sup>	$0.24\pm0.01^{\circ}$	$0.19 \pm 0.01^{d}$	
(Z)-rose oxide (2)	$0.02 \pm 0.00$	nd	nd	nd	
(Z)-linalool oxide $x(3)$	1.16±0.13 <sup>a</sup>	nd	nd	7.68±0.34 <sup>b</sup>	
(E)-linalool oxide <sup>x</sup> (4)	$1.02\pm0.03^{a}$	nd	nd	4.81±0.46 <sup>b</sup>	
(R/S)-linalool (5)	9.21±0.23 <sup>a</sup>	13±1 <sup>a</sup>	32±3 <sup>b</sup>	48±2 °	
α-terpineol (6)	100±4 <sup>a</sup>	63±6 <sup>b</sup>	89±10 <sup>b</sup>	78±5 <sup>b</sup>	
nerol (7)	0.82±0.09 <sup>a</sup>	0.93±0.13 <sup>a</sup>	3.83±0.43 <sup>b</sup>	6.03±0.42 °	
Geraniol (8)	18±2 <sup>a</sup>	22±1 <sup>a,b</sup>	28±2 <sup>b</sup>	45±5 °	
Farnesol	nd	nd	9±1 <sup>a</sup>	18±2 <sup>b</sup>	
$\delta$ -terpineol <sup>x</sup> (9)	1.19±0.09 <sup>a</sup>	0.48±0.03 <sup>b</sup>	0.59±0.01 <sup>b</sup>	0.34±0.01 °	
Norisoprenoids					
β-damascenone (10)	2.00±0.18 <sup>a</sup>	1.75±0.00 <sup>a</sup>	1.14±0.14 <sup>b</sup>	0.65±0.01 °	
β-ionone	nd	0.92±0.01 <sup>a</sup>	nd	$0.67 \pm 0.04^{b}$	
1,1,6-trimethyl-	1.09+0.08 <sup>a</sup>	0.46+0.03 <sup>b</sup>	0.42+0.01 <sup>b</sup>	0.08+0.01 <sup>c</sup>	
$1,2-dihydronaphthalene (TDN)^{(1)}$	2.84.0.07.8	4.24.0.45.8	0.00 0.00 b	0.59.0.00 °	
2 One R impers & (12)	$3.84\pm0.07$	4.34±0.45	0.96±0.09	0.58±0.02	
	2.01±0.18	1.48±0.03	$0.74\pm0.04$	$0.44\pm0.01$	
Actinidols (14)	2.84±0.32	2.24±0.04	0.91±0.01	$0.75\pm0.01$	
norisoprenoid 1 "(15)	0.69±0.04 "	$0.73\pm0.01$	$0.2/\pm0.01^{-5}$	0.05±0.01	
Volatile phenois	1	0.65.0.00 å	0.47.0.02 <sup>b</sup>	1	
m-cresol	nd	0.65±0.02 <sup>2</sup>	$0.4/\pm0.03^{\circ}$	nd	
Eugenol	nd	0.91±0.01 "	0.1/±0.01 °	$0.2/\pm0.03^{\circ}$	
4-vinylguatacol (16)	/6±1 "	31±2°	31±2°	26±1°	
4-vinylphenol (17)	8565±92 "	994±73°	9602±90°	2426±242 °	
Vanillin derivatives	2.0.5.0.00.3	4.12 0.02 h	1.46.0.10.5	1.55.0.10.6	
Vanillin (18)	2.96±0.08 °	4.12±0.03 °	1.46±0.10°	1.75±0.19°	
Zingerone (19)	0.76±0.01 "	nd	nd	1.0/±0.0/°	
Benzenes	< 0 <b>2</b> 0 11 <sup>3</sup>	4.04.0.17 <sup>h</sup>	a oo o az b	4.54. 0.00 h	
Benzaldehyde (20)	6.82±0.11 °	4.94±0.17 °	3.80±0.35 °	4.74±0.30 °	
Phenylacetaldehyde (21)	3.60±0.28 "	2.66±0.08 °	2.16±0.03 °	2.16±0.03 °	
benzyl alcohol (22)	37±1 "	20±1 °	14±1°	8.45±0.49 °	
β-phenylethanol (23)	9.39±0.62 *	7.55±0.18 "	6.09±0.29 °	6.17±0.37 °	
Ethyl cinamate (24)	8.71±0.69	nd	nd	nd	
2-phenoxyethanol (25)	5.20±0.42 *	7.95±0.67	3.29±0.42 °	5.42±0.42 *	
benzoic acid (26)	131±12 "	80±7 °	129 ±3 ª	116±5 "	
Lactones		e de la de h		=	
δ-octalactone <sup>y</sup> (27)	2.89±0.15 ª	2.10±0.18 <sup>6</sup>	14±0 °	7.65±0.93 <sup>u</sup>	
$\gamma$ -nonalactone <sup>y</sup> (28)	1.89±0.13 ª	1.94±0.16 ª	1.42±0.07 °	1.45±0.16 °	
$\gamma$ -decalactone <sup>y</sup> (29)	12±1 ª	5.55±0.45 °	23±1 °	26±2 °	
Pantolactone (30)	1.66±0.01 <sup>a</sup>	1.28±0.01 <sup>a</sup>	0.93±0.01 °	0.84±0.01 °	
Miscellaneous			<b>b</b>		
(Z)-3-Hexen-1-ol (31)	5.26±0.41 a	5.21±0.08 a	4.85±0.04 °	4.10±0.00 °	
(E)-2-Hexen-1-ol (32)	19±2 <sup>a,b</sup>	24±1 a	17±1 °	18±1 °	
ethyl decanoate (33)	16±1 ª	17±0 ª	16±0 ª	16±0 ª	
2-ethylhexanoic acid (34)	13±1 <sup>a</sup>	14±1 <sup>a,b</sup>	13±1 <sup>a</sup>	15±1 °	
4-methoxy-2,5-dimethyl-3(2H)furanone (mesifurane) <sup>x</sup> (35)	5.07±0.02 <sup>a</sup>	22±1 <sup>b</sup>	34±1 °	42±1 <sup>a</sup>	

4-hydroxy-2,5-dimethyl-3(2H)furanone (furaneol) <sup>x</sup> (36)	8.15±0.01 <sup>a</sup>	39±1 <sup>b</sup>	16±1 °	19±1 °
cinnamic acid <sup>x</sup> (37)	1678±36 <sup>a</sup>	178±24 <sup>b</sup>	850±49 <sup>c</sup>	877±61 <sup>c</sup>

nd: non detected.

(num): peak number in Figure 2.
 <sup>x</sup> Chemical standard not available. Tentatively identified. Data are relative areas (to 4-Methyl-2-pentanol x 1000).
 <sup>y</sup> Data are the relative areas (to 4-Methyl-2-pentanol x 1000). Chemical standard available,but the degradation of the products did not allow quantification.
 Concentrations of the same compound with different letter show significant differences (p < 0.05).</li>

LRI VF5- MSDBWax	Odour descriptor	Identity	% Modified frequency			
			Fue	Cam	Cdo	Sab
918	solvent, gas, glue	n.i	0	0	31	0
972	dairy product, sweet, buttery	diacetyl	55	61	55	78
1007	fruity, strawberry, sweet	isobutyl acetate	0	48	33	24
1033	fruity, strawberry, sweet	ethyl butanoate	69	59	75	73
1052	fruity, sweet, anis, cream	ethyl 2-methylbutanoate	50	46	29	0
1066	fruity, apple, anis, green, metallic	ethyl-3- methylbutanoate	61	33	69	73
1180	rubber, moisture, gas	n.i	0	0	0	34
1191	fruity, anis	methylhexanoate	17	0	0	33
1236	fruity, raspberry, strawberry, anis	ethyl hexanoate	43	33	62	55
1303	mushroom, metallic, chlorine, cucumber	1-octen-3-one	51	33	45	53
1312	spicy, green, barbecue, yeast	2-methyl-3-furanthiol	55	0	50	36
1346	floral, sweet, strawberry	(Z)-rose oxide	38	0	0	0
1378	geranium, green, pepper, lettuce	(Z)-1,5-octadien-3-one	82	51	80	29
1380	tropical, pineapple, citrus, green	methyloctanoate *	0	0	61	0
1458	vinegar	acetic acid	67	38	48	38
1548	green, grass, sweet, cucumber	(E)-2-nonenal	41	0	0	0
1552	garbage, sulfur, peanuts, barbecue	n.i	0	0	49	0
1563	floral, lemon	(R/S)-linalool	33	0	0	0
1570	unpleasant, fatty acid, vomit, vinegar	n.i	0	0	0	40
1597	tropical, sweet, caramel, cotton candy	mesifurane *	43	33	35	45
1609	strawberry	n.i	31	0	0	0
1626	burnt hair	2-acetyl pyrazine	73	61	75	53
1631	cheese, vomit, feet	butyric acid	27	17	35	43
1676	cheese, feet, sweat, milk	isovaleric acid	59	67	61	61
1730	fruity, honey, berry, tropical, sweet, floral	phenyl acetate *	0	31	22	38
1826	sweet, floral, rose	β-damascenone	0	26	0	41
1850	soil, green, spicy, pepper, peanuts, dry grass	hexanoic acid *	65	54	66	58
1865	camphor, barbecue, spicy	guaiacol	45	76	35	59
2052	caramel, strawberry, sweet	furaneol	82	82	80	85
2100	leather, animal, stable	p-m-cresol	31	47	33	36
2170	peach, sweet, strawberry	γ-decalactone	80	26	85	83
2221	animal, spicy, licorice	sotolon	45	76	0	31
2294	latex, spicy, burnt	n.i	0	0	0	53
2420	coconut, vanillin	$\gamma/\delta$ -dodecalactone	0	29	25	33

## **Table 3.** Odour Active Compounds of the Four Strawberry Varieties Analysed.

n.i: not identified

\* Tentatively identified by Lineal Retention Index and odour descriptor. Fue: Fuentepina; Cam: Camarosa; Cdo: Candonga; Sab: Sabrina.

Figure 1.



Figure 2.







# **TOC Graphic**

