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1 *Title*

2 **A holocentric twist to chromosomal speciation?**

3

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16

17

18 *Abstract*

19 Chromosomal rearrangements trigger speciation by acting as barriers to gene flow.

20 However, the underlying theory was developed with monocentric chromosomes in

21 mind. Holocentric chromosomes lacking a centromeric region have repeatedly evolved

22 and account for a significant fraction of extant biodiversity. Because chromosomal

23 rearrangements may be more likely retained in holocentric species, holocentricity could

24 provide a twist to chromosomal speciation. Here we discuss how the abundance of

25 chromosome-scale genomes combined with novel analytical tools offer the opportunity

26 to assess the impacts of chromosomal rearrangements on rates of speciation by

27 outlining a phylogenetic framework that aligns with the two major lines of

28 chromosomal speciation theory. We further highlight how holocentric species could

29 help to test for causal roles of chromosomal rearrangements in speciation.

30

32

33 While most taxonomic groups have chromosomes with centromeric regions,
34 **holocentric** (see Glossary) chromosomes that lack such regions have repeatedly
35 evolved in animals and plants [1,2]. Across the tree of life there is moreover a
36 tremendous variation in the number of chromosomes that mono- and holocentric
37 species have, ranging up to three magnitudes of difference within a taxonomic order
38 [3,4]. The evolutionary significance of this variation has gathered much attention over
39 the decades [5–7], and the interest in the evolution of chromosomal changes is currently
40 undergoing a renaissance [8–10]. This is because novel technologies make it possible
41 to obtain chromosome-scale genomes even for non-model organisms (*e.g.* [11,12]).
42 Together with the emergence of new analytical approaches, this allows tackling the
43 evolutionary impact of chromosomal variation, *e.g.* on rates of speciation [13,14] or
44 gene flow [15]. Variation in chromosome numbers may evolve through very different
45 processes. Large-scale changes in chromosome numbers can for example result from
46 hybridization events [16] or genome duplications through **polyploidization**, the latter
47 being particularly common in plants [17,18]. Other common processes include the
48 fusion of two chromosomes into a single one or the fission of a chromosome into two,
49 resulting in **dysploidy** [6].

50

51 Rearrangements that produce variation in chromosome numbers may eventually
52 result in chromosomal speciation, whereby divergent rearrangements directly or
53 indirectly cause reproductive isolation [5,6]. However, intraspecific karyological
54 variation may also persist and result in only limited levels of reproductive isolation [19–
55 22]. Two major lines of theoretical models exist that outline how chromosomal
56 rearrangements could cause chromosomal speciation (reviewed in [7,23]). The first line
57 comprises many of the classic models, which are based on hybrid dysfunction and
58 assume that differentially fixed chromosomal rearrangements between closely related
59 species cause problems during meiosis in hybrids and therefore act as **Dobzhansky-**
60 **Muller incompatibilities** (DMIs, [5,6,24,25]). The problem with these types of models
61 is that they require chromosomal rearrangements to be fixed in order to be of major
62 effect. This is because newly arising chromosomal rearrangements would typically be
63 **underdominant**, *i.e.*, they would lead to reduced fitness of hybrid individuals (Fig. 1),
64 either within or between species or populations. While strong underdominance makes

65 it unlikely that novel chromosomal rearrangements spread to fixation, weak
66 underdominance may allow for fixation, but would ensure that chromosomal
67 rearrangements represent only shallow barriers, and are therefore unlikely to cause
68 speciation [7,26]. Nevertheless, empirical evidence for such chromosomal speciation
69 exists, and has been primarily found in mammals [27], including mice [28] and
70 wallabies [8]. Here, **monobrachial homology**, *i.e.*, multiple chromosomal fusions with
71 one or more common chromosome arms in different fusion arrangements that are fixed
72 between populations or species, has been suggested to result in reproductive isolation
73 [25]. Explanations on how such species may have overcome the underdominance
74 paradox vary, and include **genetic drift**, genetic bottlenecks and founder effects [29,30].
75 Indeed, chromosomal speciation may initially result in a reduction of the effective
76 population size (N_e), which could in turn affect rates of speciation and change the
77 fixation probabilities of new karyotypes in allopatry [31]. This has been suggested for
78 mammals, where families with large geographic distributions but whose species have
79 restricted geographic ranges showed a greater probability for fixing different
80 karyotypes [32]. Shifts in mating system, *e.g.*, from outcrossing to selfing [33] or
81 **meiotic drive**, whereby some alleles or associated rearrangements are more likely to
82 be transmitted [34] have similarly been suggested to overcome underdominance. All
83 these scenarios have received much criticism in the past, however, and formal
84 experiments for a causal association between chromosomal rearrangements and
85 speciation are lacking [7,24].

86 The second major line of theoretical models was developed more recently and
87 has attempted to overcome the underdominance paradox by focusing on changes in
88 recombination associated with chromosomal rearrangements [7,23,26,35]. In essence,
89 under these suppression of recombination type models, rearranged chromosomes can
90 become fixed by drift but also by selection, *e.g.*, when two or more adaptive loci
91 become physically coupled or by locally reducing recombination, both enhancing
92 existing reproductive isolation [26,35]. Such rearranged regions of reduced
93 recombination may act as barrier loci and promote further differentiation, which may
94 eventually lead to postzygotic isolation through the buildup of genetic incompatibilities
95 [7,26,35]. Reproductive isolation associated with chromosomal rearrangements may be
96 further enhanced by sexual selection or **reinforcement** and may thus promote
97 speciation upon secondary contact. If chromosomal rearrangements contain physically

98 linked clusters of genes they may themselves represent genomic islands of
99 differentiation, or **supergenes** [36,37]. Albeit such supergenes have been suggested to
100 promote speciation [38], their actual contribution towards reproductive isolation
101 remains controversial [39].

102 The current theory on chromosomal speciation has an important gap – it is based
103 on the assumption that chromosomes are monocentric and have a centromeric region
104 that concentrates all **kinetochores** for the attachment of the spindle tubules during
105 mitosis and meiosis [5–7] (Fig. 2). However, holocentric chromosomes that lack a
106 centromeric region have evolved in very distinct taxonomic groups (Fig. 3), comprising
107 some of the most diverse branches of the tree of life such as the sedge family
108 Cyperaceae with ~5’500 species [40], the order Lepidoptera with ~160’000 butterfly
109 and moth species [41], as well as the nematode model organism *Caenorhabditis elegans*
110 [2]. In contrast to monocentric chromosomes, holocentric chromosomes have
111 molecular features that allow kinetochore proteins to bind along the entire chromosome,
112 permitting microtubules to attach broadly [1] (Fig. 2). As a consequence, rearranged
113 parts of the genome may not cause segregation problems during cell divisions.
114 Holocentricity could therefore provide a twist to chromosomal speciation theory.
115 Indeed White already highlighted in his classic work on chromosomal speciation [5]
116 that “*The laws and principles of chromosomal rearrangements in these [holocentric]*
117 *organisms are not yet fully understood, but certainly they differ in some respects from*
118 *those governing chromosomal rearrangements in species with the more usual*
119 *monocentric chromosome.”* However, despite a recent increase in interest in the
120 evolutionary implications of holocentric chromosomes, the potential effects of
121 holocentricity on chromosomal speciation have remained unclear [2,9,10].
122 Holocentricity may for example help to overcome the initial underdominance paradox
123 of the classic chromosomal speciation theory (Fig 1). This is because large-scale
124 rearrangements through chromosomal fusions as well as fissions may be more likely to
125 be retained as rearranged chromosomes maintain kinetochore function [1]. This
126 contrasts to most scenarios in monocentric species, where fission events result in
127 chromosomal segments that are not attached to a centromere and may therefore be lost
128 during meiosis (Fig. 2) or where fusion events result in dicentric chromosomes with
129 two centromeres and similarly cause problems during meiosis [42]. Monocentric
130 chromosomal fusions may not always result in segregation problems though, *e.g.* when

131 two chromosomes with terminal centromeres are involved and both chromosomal arms
132 are retain in the fused chromosome [25]. This scenario applies, however, only when
133 nearly complete chromosomes become rearranged and excludes fission events. In
134 addition, intraspecific crosses between holocentric chromosomal races or closely
135 related species may not necessarily cause a significant immediate reduction in offspring
136 fitness [19–22], suggesting that suppression of recombination could also be an
137 important driver of chromosome associated speciation in taxa with holocentric
138 chromosomes.

139

140 Mono- and holocentric species further differ in several aspects of their meiotic
141 cell division that may affect the potential for chromosomal speciation. In holocentric
142 species the recombination and segregation functions interfere during meiosis,
143 restricting the potential number of **chiasmata** in bivalents [43]. In this way, some
144 holocentric groups have evolved an inverted meiosis, where, opposite to monocentric
145 groups, the first meiotic division separates the sister chromatids and the second division
146 the chromosomal homologs [19,21]. This inverted meiosis has been suggested to
147 promote the evolution of new karyotypes and possibly chromosomal speciation by
148 facilitating a correct chromosome segregation in hybrids between populations or
149 species that differ in their karyotype [21]. Other holocentric groups, like the nematode
150 *C. elegans*, have evolved a monokinetic-like meiosis as they only keep kinetochore
151 activity in the telomeres [44] avoiding potential interference between chiasmata and
152 spindles. While these mechanisms may help to establish novel karyotypes, their impact
153 on meiotic recombination remains unclear [45], also because comparatively few
154 recombination maps exist so far for holocentric species [45–47]. Importantly, because
155 recombination is often, but not always [47], reduced close to centromeres in
156 monocentrics [46], patterns of recombination are likely to differ across holocentric
157 chromosomes. Also, while holocentric chromosomes lack a centromere, their
158 kinetochores may not be equally distributed [48]. The latter is true for *C. elegans* [44],
159 where recombination increases towards the telomeric regions in contrast to the postman
160 butterfly *Heliconius melpomene*, where recombination is similar across chromosomes
161 [47]. Processes similar to meiotic drive in monocentric species may consequently be at
162 play for holocentric species as has been found for sedges, rushes (*Juncus sp.*) and other
163 holocentric lineages [49]. However, given the repeated evolution of holocentricity, it

164 remains to show to which degree such **holokinetic drive** may be common among
165 holocentric groups.

166

167 The causality between chromosomal rearrangements and species diversification
168 has remained contentious [7]. Phylogenetic inferences suggest that rates of
169 chromosome evolution might be similar between holo- and monocentric species in
170 insects [9], and that rates of diversification are similar between holo- and monocentric
171 clades when comparing sister holo- and monocentric taxonomic orders across
172 eukaryotes [50]. However, there is often substantial variation in chromosome numbers
173 between genera or families within orders, that are moreover associated with different
174 rates of speciation [8,10,51]. The relative contribution of chromosomal fusion and
175 fission on phylogenetic species diversification varies similarly among taxonomic
176 groups and thus likely impacts rates of diversification differently [10,51]. Empirical
177 evidence for chromosomal speciation is rare, either because speciation is already
178 complete or not, often precluding causal implications of one or multiple rearrangements
179 [7,19]. The few examples for holocentric species suggest that intrinsic postzygotic
180 reproductive isolation between species with different karyotype seems to be limited for
181 Lepidoptera [16,19,22,52] and sedges [53,54]. Experimental hybrids between
182 cytogenetic races of the same sedge species showed that hybrid dysfunction is very
183 limited between populations that differ in few chromosome rearrangements but
184 increases as the number of chromosome rearrangements increase [53,54]. These few
185 empirical examples contrast the vast diversity of the taxonomic groups that have
186 evolved holocentric chromosomes and karyotypic diversity [1,2,9]. Here, novel
187 phylogenetic approaches [13,14] could help to assess the macroevolutionary
188 implications of changes in chromosome numbers more generally and provide a
189 framework for comparative analyses between holo- and monocentric groups.

190

191 *A phylogenetic framework of chromosomal speciation*

192 Recent advances allow to disentangle models of chromosomal evolution in a
193 phylogenetic framework and to distinguish if a phylogenetic event is rather associated
194 with **ana-** or **cladogenesis** [13,14] (Fig. 4). Under cladogenesis, karyotype evolution
195 occurs at a speciation event, while under anagenesis karyotypes evolve along a branch
196 and speciation happens later. Ana- and cladogenesis are compatible with the two
197 aforementioned major lines of chromosomal speciation models, where cladogenesis

198 resembles the classic hybrid dysfunction type models and anagenesis the recombination
199 suppression type models. Importantly, ana- and cladogenetic processes may not be
200 exclusive and may similarly result in a phylogenetic event when they occur together.

201

202 The phylogenetic framework outlined in (Fig. 4) allows to quantify how
203 common changes in karyotype numbers might be associated with speciation events at
204 a macroevolutionary scale and to compare between mono- and holocentric clades [9].
205 Current limitations are primarily given by the availability of dense phylogenies
206 associated with associated karyotype data, often only allowing to study chromosomal
207 speciation at a lower taxonomic level [10]. As this framework allows identifying
208 branching events that are more likely to have resulted in ana- or cladogenetic events
209 respectively, such species pairs could be used to perform in-depth comparative genomic
210 analyses to identify which rearrangements are more likely to result in one or the other
211 phylogenetic event.

212

213 Because a correct segregation of chromosomes may initially be often possible
214 in hybrids of holocentric parental species with different karyotypes [21], holocentricity
215 provides an excellent system to experimentally study chromosomal speciation. The
216 outlined phylogenetic analyses (Fig. 4) combined with crossing experiments could for
217 example quantify the impact of chromosomal rearrangements on reproductive isolation
218 in relation to *e.g.*, the respective evolutionary distance among distinct species pairs.
219 Irradiation based experiments on holocentric plants moreover suggest that
220 holocentricity and a fast formation of new telomeres at breakpoints enables rapid
221 karyotype evolution in holocentric species, though the impact on reproductive isolation
222 was not tested [55]. As direct experimental manipulations of individual chromosomes
223 become technically feasible through novel laser nanosurgery approaches [56] or by
224 generating **artificial chromosomes** [57], the outcome of specific artificial fusion or
225 fission events can now be experimentally studied, enabling to recreate karyotypic
226 changes between sibling species and to assess their direct impact on reproductive
227 isolation.

228

229 *Concluding remarks*

230 Although some of the most diverse taxonomic groups of animals and plants have
231 evolved holocentric chromosomes [1], the potential evolutionary implications of

232 holocentricity remain elusive. As we outlined, holocentricity could provide a new twist
233 to chromosomal speciation but further research is required. Studying holocentric
234 species could help to advance our understanding on chromosomal speciation (see
235 Outstanding Questions). In addition, we suggest future theoretical explorations, for
236 example, to assess the potential for chromosomal speciation in holocentric taxa, where
237 novel rearrangements may not immediately result in hybrid dysfunction but include a
238 lag time during which heterozygous rearrangements may be tolerated [19]. While
239 chromosomal rearrangements could be an important driver of speciation in cases where
240 they contribute to reproductive isolation, additional pre-zygotic barriers may need to
241 subsequently evolve to complete the speciation process [7]. Comparisons between
242 evolutionary young sibling species that coexist or form zones of secondary contact are
243 thus needed to assess the contribution of chromosomal rearrangements on reproductive
244 isolation in relation to other barriers [58]. To further gain a better understanding on the
245 macroevolutionary impact of karyotype evolution and holocentricity more in-depth and
246 comparative analyses are required to first identify the genomic mechanisms underlying
247 chromosomal fusion and fission sites and their (non-)parallelism across holocentric
248 groups. This would then allow us to identify why for example in Lepidoptera only some
249 genera show tremendous karyotypic variation whereas other genera show none [10].
250 Lastly, the increased availability of genomic resources for non-model species combined
251 with recently developed models for chromosome evolution [13,14] allow for large-
252 scale macroevolutionary studies both within and across taxonomic orders to decipher
253 the evolutionary consequences of holocentricity.

254

255

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412 in Croatia, at the north-western edge of its distribution. *Nat. Slov.* 17, 47–57.

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416
417

418 **Figure legends**

419 Fig. 1: The underdominance paradox. Depicted are hypothetical fitness landscapes for
420 species with different karyotypes and their F1 hybrids with the effects of strong and
421 weak underdominance as predicted by theory for monocentric species [7,26]. While no
422 such theory exists for holocentric species, predictions are given based on empirical
423 findings, which suggest that F1 hybrids in holocentric species may not necessarily
424 suffer from the underdominance paradox [19,21].

425

426 Fig. 2: Comparison of the outcomes of fission events during cell division for mono- and
427 holocentric species. If fission occurs during anaphase, the fragment that is not attached
428 to a centromere is lost for monocentric species. In contrast, fragmented chromosome
429 sections of holocentric species can maintain kinetochore function due to the distribution
430 of centromere-like structures along the chromosome, and may so be retained.

431

432 Fig. 3: Examples of the diversity of holocentric species and their haploid karyotypes.
433 A – *Carex esenbeckiana* (n = 13). B – *C. fischeri* (n = 36). C – *Polyommatus atlantica*
434 (n = 224), adapted from the Natural History Museum London & [59]. D – *P.*
435 *aroaniensis* (n = 47), adapted from [60], [61]. Pictures in A, B – courtesy of Modesto
436 Luceño Garces. Scale bars represent a length of 10 μ m.

437

438 Fig. 4: Contrasting phylogenetic models of karyotype evolution with their putative
439 counterparts of major lines of chromosomal speciation models. The outcome of the
440 different models of karyotype evolution are outlined along a hypothetical phylogeny,
441 with clado- and/or anagenetic karyotypical changes being indicated. Colors of branches
442 indicate changes in haploid chromosome numbers, while color gradients indicate that
443 the process of karyotype fixation may occur more slowly after anagenetic changes.

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451 **Glossary**

452

453 **Anagenesis:** Type of speciation in which an ancestral species gradually evolves
454 into another by accumulating changes within a single lineage over time.

455

456 **Artificial chromosomes:** Artificially created chromosomes that have the
457 necessary properties (e.g. centromeres, telomeres and origins of replication) to be self-
replicating and stable.

458

459 **Dobzhansky-Muller incompatibilities:** Negative epistatic interactions or
460 incompatibilities that occur between loci with different evolutionary histories.
461 Populations may diverge in allopatry and accumulate such incompatibilities through
drift and/or through mutations that prevent hybridization upon secondary contact.

462

463 **Dysploidy:** Process that increases or decreases the number of chromosomes
464 within a species through chromosomal rearrangements with no significant changes in
DNA content.

465

466 **Chiasma:** Point of contact between chromatids from two homologous
467 chromosomes during meiotic divisions that allows recombination through
chromosomal crossovers between both chromatids.

468

469 **Cladogenesis:** Type of speciation in which an ancestral species splits into two
or more species.

470

471 **Genetic drift:** A stochastic evolutionary process that results in changes of allele
frequencies by sampling a finite number of individuals each generation.

472

473 **Holocentric / holokinetic chromosome:** Chromosomes with non-localized
474 centromere-like structures. The kinetochore activity is distributed along the whole
chromosome.

475

476 **Holokinetic drive:** Perturbation of the normal meiotic process so that a
477 particular allele is preferentially transmitted to the progeny over another allele caused
478 by variation in kinetochore distribution along the holocentric chromosomes or the size
of holocentric chromosomes.

479

480 **Kinetochores:** Protein structures located on the chromosomes. Microtubules of
the mito- or meiotic spindles are anchored to this structure during cell division. For

481 monocentric species, kinetochores are located in the centromere whereas for
482 holocentric species, they occur throughout the chromosomes.

483 **Meiotic drive:** Perturbation of the normal meiotic process so that a particular
484 allele is preferentially transmitted to the progeny over another allele. The centromere,
485 its location and size are factors that can result in meiotic drive.

486 **Monobrachial homology:** Homology between two bi-armed chromosomes that
487 is restricted to only one of the two chromosome arms.

488 **Polyploidy:** Chromosome multiplication entailing the addition of complete
489 chromosome sets.

490 **Reinforcement:** Evolutionary process whereby pre- or postzygotic mechanisms
491 increase reproductive isolation between two closely related lineages upon secondary
492 contact.

493 **Supergene:** A set of genes in strong linkage that segregate together during
494 meiotic divisions because there is a mechanism that impedes recombination within the
495 supergene, such as chromosomal rearrangements, like inversions.

496 **Underdominance:** Strong selection against heterozygotes. For chromosomally
497 diverging populations, chromosomal hybrids have low fitness and there is a strong
498 selection against them.

499

500

501

Highlights

Chromosomal speciation, whereby major chromosomal rearrangements trigger reproductive isolation, is a classic evolutionary concept.

The underlying theory was developed for chromosomes with centromeres when holocentric chromosomes that lack centromeres have repeatedly evolved across the tree of life.

We argue that holocentricity may help to overcome problems associated with classic chromosomal speciation theory and that the special characteristics of holocentric chromosomes vastly expand the potential for experimental research on chromosomal speciation.

We outline how new approaches allow to quantify the macroevolutionary impact of chromosomal speciation and to distinguish the associated evolutionary mechanisms.

Outstanding questions

- What are the genomic features underlying chromosomal fusion and fission sites and did they evolve repeatedly across the tree of life? Are there common rearrangement hotspots?
- How do chromosomal rearrangements affect gene flow and does it differ between mono- and holocentric species?
- If rearranged chromosomes act as barrier loci, how does reproductive isolation buildup in the rest of the genome? Are rearranged regions enriched for functional genes?
- How does recombination differ between mono- and holocentric species and what are the implications of fusion and fission on recombination?
- To which degree do ana- and cladogenic phylogenetic events reflect the two lines of chromosomal speciation theory?
- What is the macroevolutionary impact of chromosomal rearrangements between mono- and holocentric species and what are the predominant underlying mechanisms? (see Box 1)
- Is chromosomal speciation more likely to occur in holocentric species?

Figure 1 (PDF)

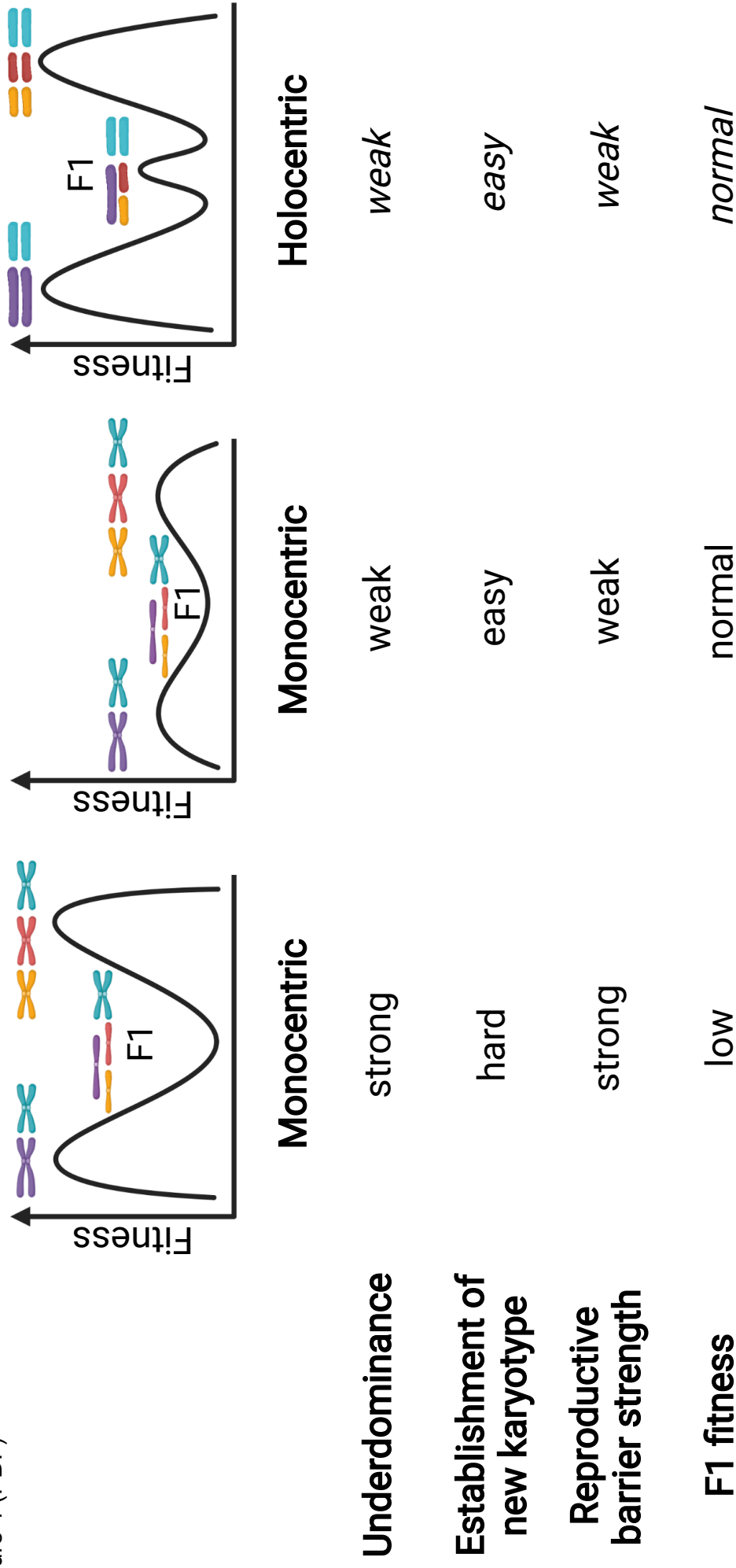
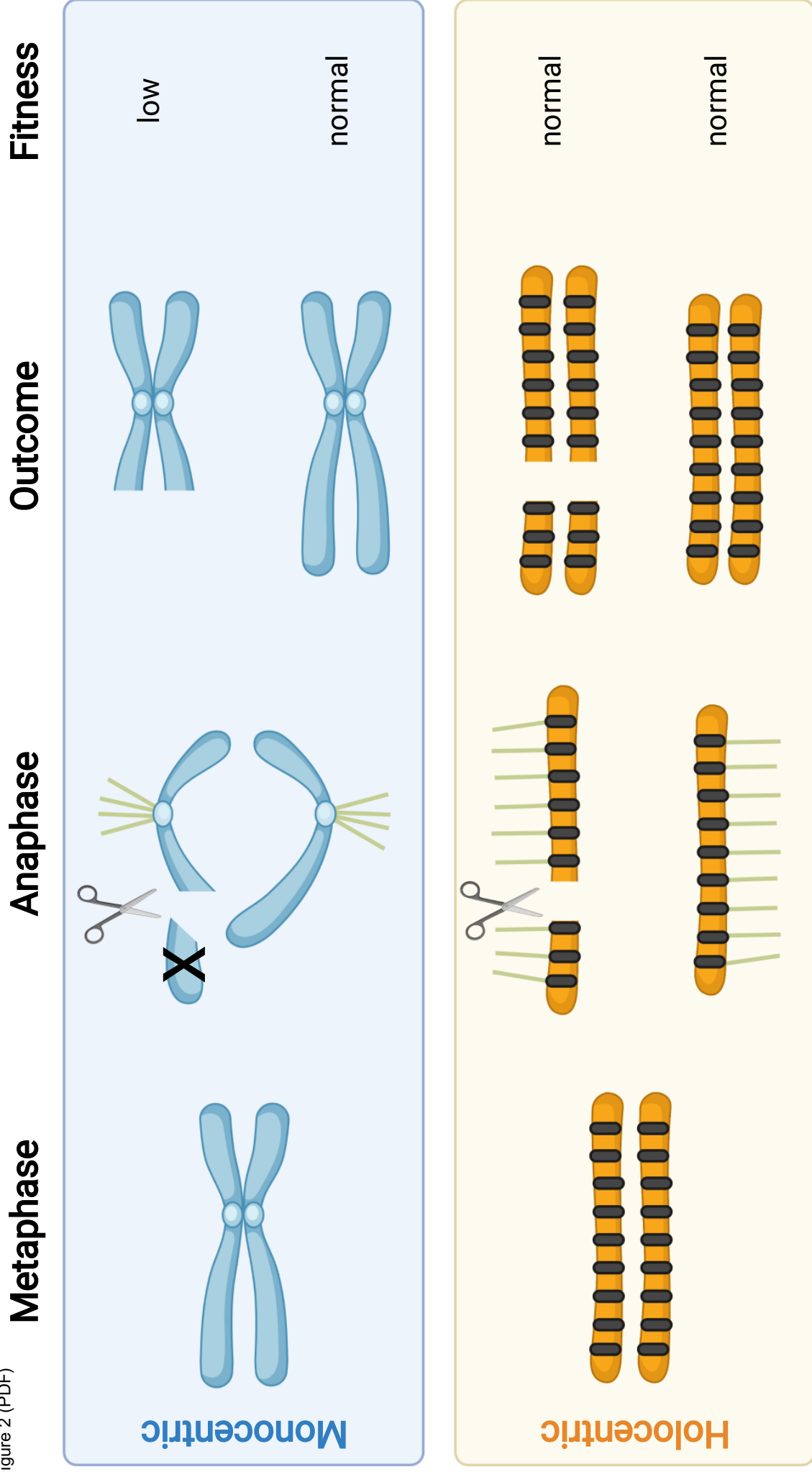


Figure 2 (PDF)



Fission — Microtubuli ● Centromere

● Centromere-like structure

X Loss of chromosome fragment

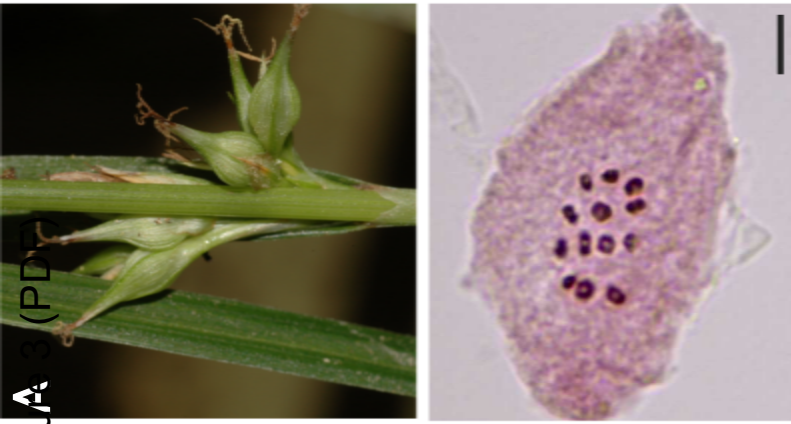
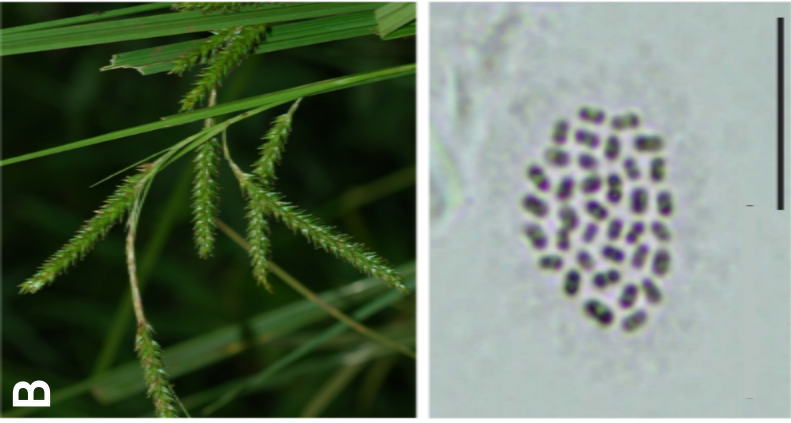
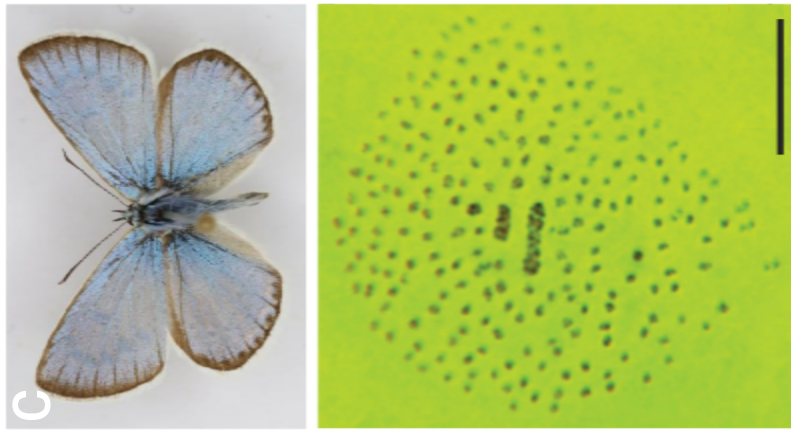
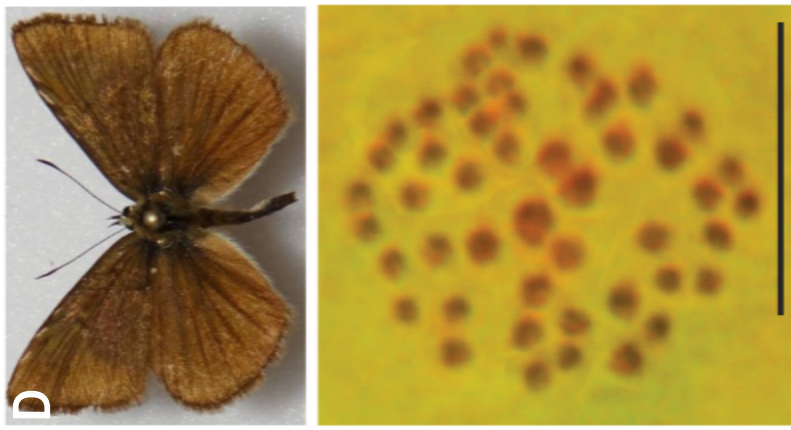


Figure 3 (PDA)

Figure 4 (PDF)

