A rapid-Soxhlet & mini-SPE method for analysis of polycyclic aromatic hydrocarbons in atmospheric particles

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Abstract

An analytical method was validated with two reference materials of polycyclic aromatic hydrocarbons in atmospheric particles. SRMs were incorporated into the matrix of unexposed cut quartz filters. The methodology was previously designed and extraction of PAHs from fortified filters was based on a rapid low-cost method, for a low consumption of volume and time. The optimisation combined a low-volume Soxhlet apparatus performed at hot Soxhlet mode with a quick clean-up by solid phase extraction with special cartridges. The quantification of target compounds was performed by gas chromatography/mass spectroscopy in SIM mode. Temperatures of injector and oven program of the GC-MS were also optimised. Experimental variables of both systems were successfully optimised and validated, achieving a robustness analytical methodology.

Keywords Polycyclic aromatic hydrocarbons · Atmospheric particles · Reference Materials · Gas chromatography · Clean-up · Validation

Introduction

Polycyclic aromatic hydrocarbons (PAHs) are lipophilic compounds with important carcinogenic risk. These hazardous hydrophobic organic compounds (HOCs) led international organisms to establish policies for the environment [1] and indoor air [2].

The European Commission exposed a wide toxicological guidance since the 2001 position paper [3] to the Europe 2020 strategy [4]. Similar measures were taken by other European organizations [5]. In the atmospheric environment, PAHs emissions are well known, as they originated from car exhausts [6], domestic combustion [7] industrial activities [8], agriculture activities [9] and natural sources [10], such as biomass burning [11]. Determination of low levels of PAHs in complexes matrices such as atmospheric particles leads to search accuracy analytical methodologies, which needs to be optimised and validated with recognized material references.

Traditional extraction procedures for PAHs involve many techniques, such as direct extraction with organic solvents, sonication, Soxhlet extraction, etc. [12]. In order to speed up sample preparation, new methodologies such as accelerated solvent extraction [13] or microwave-assisted solvent extraction [14] or ultrasound-assisted solvent extraction [15] have been developed, saving processing time and solvent consumption. However, these new techniques are expensive. Soxhlet extraction represents an inexpensive method for solid samples, nevertheless is rather time-consuming and requires high solvent consumption. An improved Soxhlet technique, called Hot Soxhlet, heat the extractor body at lower temperature than the boiling point of solvents to keep it in the liquid state [16]. Additionally, after extraction step it was needed purification steps by solid phase extraction techniques like packed columns of silica, Florisil®, alumina or mixtures [17].

Therefore, the aim of this paper is to optimise and validate a low-cost and rapid extraction methodology with low time and solvent consumption. The method was based on an improved and more rapid Soxhlet extraction plus an improved, short and more efficient SPE technique for the 16 PAHs included in the US EPA priority pollutants list [18] in airborne particles.

Regarding to the validation process, suitable certified reference materials should be used, but much better with two references [19] to confirm the suitability of the method. Additionally, the reference material was mixed with small pieces of unexposed filter, taking the mixture as a new reference for the same matrix of real samples of particles, which are collected on filters during the sampling. This spiking technique constituted an innovative idea in validation techniques. The relevance of the work is the fact that many modern techniques that reduce solvent volume and time consumption are expensive when there is currently a global crisis due to SARS-CoV-2.

Material and methods

All solvents used in the present study were of analytical and chromatographic grade.

Optimisation of the new procedure: extraction, purification and quantification

We validated in 2005 a five-stage analytical procedure for PAHs in airborne particles using the NIST 1649a (National Institute of Standard and Technology, Gaithersburg, MD, USA) [20] where extraction was performed by Soxhlet with 250 mL of solvent for 10 hours. Also, we optimised in 2016 a four-stage procedure for fat in olive fruits [21] using 100 mL of solvent for 2.5 hours. The proposed methodology reduced the method to only two stages before GC quantification, reducing time of extraction and volume consumption. Thus, optimisation of this new method was structured as follow:

Optimisation of the extraction stage

Optimisation of PAHs extraction was done by intercomparing three experiments using three Soxhlet apparatus, with solvent volumes of 250 mL [21], 100 mL [22] and 25 mL. The lower the volume of extractor body, the lower the time of extraction.

For these experiments, 100 mg of SRM 1649a was added to each extractor body over one quarter of unexposed QM/A quartz filters of 4 x 5 inches (Whatman International, Maidstone, England) cutted in small pieces of 0.25 cm^2 (0.04 $inch^2$). Previously, quartz filters were heated in a muffle furnace at 500ºC for 2 h to remove residual organic traces. The mixture employed was a 5:1 acetone/methylene chloride (Merck, Darmstadt, Germany) mixture including *pyrene-d10* (Dr. Ehrenstorfer GmbH, Augsburg, Germany) as deuterated surrogate standard. Volumes of organic extracts (around 250 mL and 100 mL) were reduced by rotary evaporation and then until 2 mL inside test tubes by slowly nitrogen concentration. Purification and quantification stages were carried out for the time being as in the 2005 validation [21]. Extracts of 25 mL extractor body was nitrogen-concentrated directly. After this volume optimisation, the time of extraction was then minimized on the best Soxhlet system obtained from 250, 100 and 25 mL.

Optimisation of the purification stage

The process of analytes isolation requires a sample clean-up stage to remove interferences. The extracts of filter from the extraction stage were purified comparing the OCC technique (open-column adsorption liquid chromatography) and the SPE technique in normal-phase. In the OCC alumina/silica (from Merck) was used as adsorbent [21], in the SPE miniSpe-ed *Plus* silica-gel cartridges (Applied Separations, Allentown, PA, USA) were proposed. SPE allows isolating PAHs from a sample through the same chemical principles of column chromatography but with low consumption of solvent and quickly.

The OCC technique was performed in glass columns (30 cm x 1 cm) filled with 1 g of each alumina/silica-gel (top/bottom), conditioning it with 30 ml of n-hexane (from Merck). The extract (2 mL) was transferred to the top and the non-polar fraction was eluted with other 2 mL n-hexane, 4 mL were discarded. The aromatic fraction was

eluted with 7 mL of 20% methylene chloride in n-hexane and nitrogen-concentrating < 1 mL –adding here the deuterated standard mixture–, and making up to 2 mL into a chromatographic vial.

SPE technique with miniSpe-ed *Plus* silica-gel cartridges was performed in a Varian vacuum manifold (Varian Inc., scientific instruments; Palo Alto, CA, USA). Each cartridge (450 mg/1 mL) was conditioned with 1.5 mL of n-hexane and 1.5 mL methylene chloride. Then, for the aliphatic fraction, 2 mL of extract plus a few microlitres of deuterated internal standards (ISs mix) were loaded and eluted with 3 mL of n-hexane at a flow of 0.3-0.5 mL min^{-1} (higher elution speeds lead to low retention volume), and then it was discarded. Then 3 mL of methylene chloride was used for eluting PAHs into a 2 mL-vial. The chromatographic vial was located inside the vacuum manifold assisting it with a flow of nitrogen. So, a slow nitrogen-concentration stage was not necessary.

Optimisation of the quantification. GC/MS experimental conditions

The sixteen PAHs listed as priority air pollutants by the US EPA [22] were identified and quantified using gas chromatography (Agilent Serie 6890A, Santa Clara, CA, USA) with mass-spectrometry (Agilent Serie 5973N). A capillary column of (5%-phenyl)-methylpolysiloxane (low polarity) was used (HP-5ms, 30 m x 0.25 mm i.d. x 0.25 μm film thickness) from Agilent Technologies. Helium (99.9995% purity) as carrier gas was operated at constant pressure.

Chromatograms obtained at different temperatures of the injector and different optimised oven programs were compared. These temperatures were systematically optimised by using an orthogonal design approach. First, the inlet temperature was tested at 250, 260, 270 and 280 ºC with the rest instrumental conditions fixed. Second,

the oven temperature program was optimised in order to obtain the best resolution and separation of chromatographic peaks. Three temperature programs were tested according the following conditions:

PrA: The initial temperature of 60 ºC was kept for 1 min, then it rose to 210 ºC at 15 ºC min^{-1}, where kept for 1 min, and finally to 280 ºC at 15 ºC min^{-1}, kept for 25 min. Total time: 41.7 minutes.

PrB: The initial temperature of 60 ºC was kept for 2 min, then it rose to 200 ºC at 7 ºC min^{-1}, where kept for 2 min, and finally to 290 ºC at 10 ºC min^{-1}, keep for 35 min. Total time: 68.0 minutes.

PrC: The initial temperature of 60 ºC was kept for 1 min, then it rose to 175 ºC at 20 ºC min^{-1}, where kept for 3 min, then it rose to 300 ºC at 5 ºC min^{-1}, kept for 20 min. Total time: 54.8 minutes.

Once temperatures were optimised, the instrument quantification method [23] required a calibration curve 4-1,000 μgL^{-1} [ppb] for each. The chromatographic signal of each PAH was relative to a deuterated PAH of the *acenaphtene-d_{10}*, *phenanthrene-d_{10}*, *chrysene-d_{12}* and *perylene-d_{10}* internal standards mixture at 200 ppb (from Dr. Ehrenstorfer). Each peak was identified by the absolute and the relative retention times, and by comparison with the mass spectral library of the instrument [24] using a target ion, primary ion (T) and a qualifier molecular ion (Q) (see Supplementary Information (ESM), Table S1).

Validation of the methodology

In order to study the accuracy (trueness and precision according ISO [25]), two NIST Certified Reference Materials for PAHs were used, to cover two levels of

concentrations: SRM 1649a-Urban Dust and SRM 1648a-Urban Particulate Matter (from NIST) [19,26]. These studies were developed on SRMs 'with/without' filters. Studies to demonstrate the accuracy of the method included (see ESM): a) recovery study, b) t-tests and *F*-assays, c) precision study by Horwitz ratios (HorRat), d) linearity study, e) sensitivity, f) selectivity/specificity determinations, g) limits of detection and quantification, h) ruggedness.

For a), b) and c) studies, twelve replicates of 100 mg of the two SRMs were analysed. Other nine calibration series, which employed PAHs standards and internal standards, constituted the base of the d) to g) validation studies. All experiments of ruggedness were performed with 100 mg of SRM. All statistics were done in agreement with guidelines for validation of the AOAC (Association of Official Analytical Chemists) [27], the IUPAC (International Union of Pure and Applied Chemistry) [28] and the SANCO-DG (Directorate General for Health and consumer Affairs at the European Commission) [29].

a) Recovery study: Recoveries were calculated for n = 12 replicates analysed in different days and weeks according Equation (1, see ESM). Each recovery value was compared with the AOAC acceptable recoveries.

b) F-assays and t-tests: Hypothesis *F*-tests for the precision and *t*-tests for the trueness were done at p = 0.05 for the n = 12 replicates by comparing variances and mean values of the measurements with those certified by the NIST SRMs according Equations (2, 3, and 4, see ESM)).

c) Precision study: The study of the intra-laboratory precision was done under reproducibility conditions (RSD_R), according a twelve replicates experiment based on two daily sessions with duplicates (j =1 and j =2, morning/afternoon sessions), for n = 1,

2, 3 days of different weeks (between-days). The expected AOAC values of RSD for reproducibility ($ERSD_R$), the predicted RSD_R values ($PRSD_R$) and the Horwitz Ratio, *HorRat*, see Equation (5, see ESM), were used as indicator of precision according the Horwitz or Thompson theories [30,31].

d) Linearity R^2, r, L and CV: The linearity study was performed from data of different calibration curves during the validation processes. Coefficient of determination (R^2), the Pearson coefficient of correlation (*r*), the 'Goodness' (*t*-significance), the percentage (*L*) of linearity [32] or the 'online linearity' (*CV*) were calculated according Equations (6 to 10, see ESM)).

e) Sensitivity γ and δ: The PAH sensitivity was determined as the 'sensitivity of calibration' (*gamma*, γ) and as the 'analytical sensitivity' (*delta*, δ). γ is defined as the slope of the regression curve (Equation 11, see ESM). δ is the ratio of calibration sensitivity to standard deviation of the slope (Equation 12, see ESM).

f) Selectivity: Interferences can be introduced through the sample matrix, the sampling system or the instrument system. To reduce interferences we evaluated the use of pure solvents, the sample handling with laminar air-flow cabinets and the use of blanks. Specifically, clean-up procedures were employed to remove most of these substances. In the case of co-eluting compounds, the mass spectrum can be easier interpreted working in SIM mode.

g) Limits of detection and quantification, criteria comparison: Limits of detection were essentials and they evaluated by various criteria. First, both limits were determined based on 1978-IUPAC criteria (Equation 13, see ESM) from the mean of procedural blanks (LOD_{Bl}, *x3* and LOQ_{Bl}, *x10,* n = 8), i.e. matrices containing only 1 µL of 50 pg μL^{-1} of the ISs mix [27].

Second, limits of detection and quantification were also estimated as instrument detection limit (LOD_I, *x3* and LOQ_I, *x10,* n =12) from the standard deviations (Equation 14, see ESM) of calibration curves (ISO 11843, [33]).

For chromatography, standards with concentration close to LOD_I were required (n =12). It is the *noise detection limit*, LOD_N (*x3)* and LOQ_N (*x10*) prepared from spiking blank samples: we spiked unexposed cutted quartz filters with 1 µL of 100 pg μL^{-1} (100 ng of the 16-PAHs mix standard and 50 ng of the ISs mix). Calculation was from the signal-to-noise ratio (*S/N*) [34] according Equation 15, see ESM).

Also, the limits were determined according to the method used by the ISO-17025 accredited laboratories. The limit of quantification was estimated from a standard at a near *Zero Concentration* (*NZC*). Thus, three calibration curves were studied by quadruple at low concentrations < 25 μgL^{-1} [ppb], preparing at high range (HR), 5 – 25 pg μL^{-1}, medium range (MR), 0.5 – 2.5 pg μL^{-1} and low range (LR), 0.05 – 0.25 pg μL^{-1}. Then, the standard with the lowest concentration (determined with less than 3% error) is selected as the reference for the limit of quantification (LOQ_{ZC}). The limit of detection (LOD_{ZC}) was estimated as one half of the quantification limit. Finally, we expressed the method detection limits (*MDLs*) and method quantification limits (*MQLs*) as those expressed in pg m^{-3} and ng g^{-1}.

h) Ruggedness study: The method performance was evaluated using a 'ruggedness test' [35,36] by the Youden method. It was based on the Plackett-Burman orthogonal design of eight experiments and seven factors (L8, $2^{(7-4)}$). They included small changes in seven (A-G) chromatographic factors at two levels for each factor (over-default or high-low) with regard to nominal conditions, affecting to the four following experimental variables:

1. Conditions of the low-volume Soxhlet apparatus: assessment of extraction time (A) and solvent volume (B) used.
2. Conditions of the Solid Phase Extraction technique: assessment of volume (C) and flow rate (D) of elution.
3. Conditions of the solvent evaporation: assessment of the gas flow strength (E) of the nitrogen-assisted solvent evaporation.
4. Conditions of the the chromatographic system: assessment of the injected volume (F) of the sample extract and the final injector temperature (G).

Ruggedness was determined by triplicate with 100 mg of both SRMs according the Hadamard matrix. We then evaluated the variables that best and worst adapted to the small "accidental" changes.

Results and discussion

The results for searching the best procedure are shown below. Statistical techniques were: ANOVA for intercomparison of the three experiments of extraction. Snedecor-Fisher (*F*-test) for precision and Student's (*t*-test) for accuracy and intercomparison of the two purification techniques.

Results of the optimisation

Extraction stage

In the intercomparison of the three Soxhlet systems, the extraction time reached 10 hours for the 250 mL extractor body, and 2.5 hours for those of 100 mL. In the case of extraction at 25 mL, a special mini-Soxhlet system was required: the Quickfit® Soxhlet extractor body with only 20 mL of siphoning volume. The time of extraction was also

optimised here. Procedure for 250 mL, 100 mL and 25 mL were satisfactorily compared (n = 5) of NIST SRM 1649a.

Firstly, *F* and *t*-tests (p = 0.05) applied to both 250 mL 100 mL series showed no significant difference between them for the 13 PAHs of the SRM (all $F_{calc} < 1.37$ for $F_{crit} = 9.60$ and all $t_{calc} < 1.02$ for $t_{crit} = 2.31$, $p > 0.05$). However, *F* and *t*-tests applied to both 250 mL and 25 mL series and to both 100 mL and 25 mL series showed significant differences between 250 mL or 100 mL and 25 mL series (all $F_{calc} > 15.42$ for $F_{crit} = 9.60$ and $t_{calc} > 5.13$ for $t_{crit} = 2.31$, $p < 0.05$). Secondly, ANOVA showed significant differences along the three series of solvent volumes ($F_{calc} = 13.31 > F_{crit} = 3.26$, p = 0.008). In addition, recoveries of the 25 mL procedure were higher (+ 4.3-4.8 %) than those of 250 mL and 100 mL ($t_{calc} < 0.96$ for $t_{crit} = 2.07$, $p > 0.05$) (Fig. 1). As a result, the Quickfit® Soxhlet reduces the required volume of solvent to 25 mL and it allows a high number of cycles (67-83) and shorter (65-80 seconds), reducing the total time of extraction to 60 minutes (see ESM, Fig. S1).

The low volume (24-25 mL) of the final extract implies a short duration of the NASE (Nitrogen-Assisted Solvent Evaporation), which, additionally, were done inside the vacuum manifold. On the other hand, to further increase the efficiency of the rapid extraction, this was performed under the 'hot Soxhlet' conditions [16].

Purification stage

After extraction 24-25 mL of the extract were concentrated to 2 mL inside the vacuum manifold before the purification stage. In the clean-up optimisation both OCC and SPE series were compared (n = 5) using SRM 1649a (Fig. 2), showing no significant difference between them (all $F_{calc} < 2.06$ for $F_{crit} = 9.60$ and $t_{calc} < 1.18$ for $t_{calc} = 2.31$, $p > 0.05$). However, recoveries obtained with SPE were higher (+ 2.0 %) than for OCC ($t_{calc} < 0.81$ for $t_{crit} = 2.07$, $p > 0.05$).

In conclusion, according the best recoveries obtained with Quickfit® Soxhlet and SPE, the methodology proposed combines the Accelerated mini-Soxhlet extraction (AmSE) assisted by hot Soxhlet and the mini-solid phase extraction with miniSpe-ed (mSPE) assisted with simultaneous nitrogen evaporation, as the Rapid Soxhlet & Solid Phase Extraction (RSE-mSPE) method.

The final RSE-mSPE method was performed in two steps:

First, accelerated mini-Soxhlet extraction on atmospheric filters for 60 min with 25 mL of solvent, using the Quickfit® Soxhlet, and second, a mini-solid phase extraction with silica-gel miniSpe-ed on filter extracts for the PAHs isolation before GC injection.

Quantification

GC/MS experimental conditions

Measurements in the chromatographic system were done using helium at a constant pressure of 20 psi, operating in pulsed splitless mode, and injecting 2 μL of all the samples, split opened after 30 s. Optimisation of the inlet temperature was tested at 250, 260, 270 and 280 ºC, fixing the other parameters. All PAHs of 1649a SRM were observed, studying especially the representative compound, benzo[*a*]pyrene [*BaP*] as the first marker of total PAHs in scientific reports [37]. When the injector temperature increases from 250 to 280 ºC (see ESM, Fig. S2), the maximum peak area of the *BaP* was given to 260 ºC, therefore, the temperature of 260 ºC was adopted as the optimal temperature of the injector.

As for the *PrA*, *PrB* and *PrC* oven temperature programs, the three were obtained with good resolution between PAHs. The responses of chromatographic peaks were similar for the three programs tested. However, because of the second is too long (68 min) and although the first is shorter (42 min), the third program has slightly higher

peak resolution for some PAHs than the other two, so the oven temperature program selected was *PrC* (55 min): 60 ºC, 1 min, 20 ºC min^{-1} to 175 ºC, 3 min, 5 ºC min^{-1} to 300 ºC and hold for 20 min.

Finally, the mass selective detector was operated in electron ionization mode with electron energies of 70 eV, being the ion source temperature of 230 ºC. To improve sensitivity, quantitative analysis operated in Selected Ion Monitoring (SIM) mode instead of Full Scan mode. The selected molecular ions of the different PAHs were shown in Table S1 (see ESM).

Results of the validation

Recovery study

Recovery and RSD values obtained for both SRMs (n = 12) were shown in Table 1. Average recovery for SRM1649a+filter was *Rec* = 97.9% (82.6 - 107.2 %) and for SRM1648a+filter was *Rec* = 95.5% (75.7 - 104.8 %), with *RSD* = 2.0% for SRM 1649a and 1.5 % for SRM 1648a. All recoveries were 'acceptable' according to the AOAC ranges, showing excellent agreement between measured and certified values. Differences between experiences with / without pieces of filters were negligible, 97.8 % without filters (SRM only) and 98.9 % with filters (as real samples) for SRM 1649a and a similar difference for SRM 1648a.

The aromatic compound with the lowest recovery in SRM 1649a was fluorene (3 rings) with a value of 82.6 % [38]. This was probably due to its volatility (*MW* = 166 uma and *Pv* = 0.09 Pa). Similar results in SRM 1648a were for acenaphthene and acenaphthylene (76 %, 3 rings, *MW* = 152-154 uma and *Pv* = 0.3-0.9 Pa) and naphthalene (82 %, 2 rings, *MW* = 128 uma and *Pv* = 8.6). On the other hand the

recovery of the extraction method from pyrene-d_{10} was high with a value of 98.1 ± 2.8%.

Precision study

Results of *F*-tests (Table 2) revealed that, for all PAHs of both SRMs, differences between variances were not significant ($F_{crit} > F_{calc}$, $p = 0.05$), except for indeno[*1,2,3-cd*]pyrene and benzo[*ghi*]perylene in SRM 1649a, also, RSD_R values for *naphthalene* and dibenzo[*ah*]anthracene were higher than those acceptable by the AOAC. Afterwards, *t*-tests showed good results, but not for *fluorene* in SRM 1649a and *acenaphthene*, *acenaphthylene*, benzo[*b*]fluoranthene and dibenzo[*a,h*]anthracene in SRM 1648a, which showed significant differences against true values ($t_{crit} < t_{calc}$, $p = 0.05$). As a result, from the recoveries (a) and the *t*-tests and *F*-assays (b), we can affirm that the proposed new analytical methodology was traceable to both NIST SRMs without any important systematic error.

The intra-laboratory study (Table 2) showed that values of RSD_R were lower than AOAC–$ERSD_R$ values in all PAHs of SRM 1649a, so they were acceptable and similar than other works [39]. For SRM 1648a, only the RSD_R values for *naphthalene* and dibenzo[*a,h*]anthracene were higher (41.6% and 47.7%) than the acceptable (22% and 14.6%).

For Horwitz criteria, only *naphthalene* and dibenzo[*a,h*]anthracene have values of Horwitz ratios (HorRat) higher (2.6) than the acceptable values (0.3-1.3) in SRM 1648a. This fact occurred precisely for the two PAHs whose certified RSD values were high, indicating good precision for repeatability but not for reproducibility.

Linearity study

The linearity of the different calibration curves (4 – 1,000 ppb) can be represented by the goodness or *t*-significance (t_{calc}) and by other parameters (r, R^2 and L). In Table S2 (see ESM), the parameters showed excellent results at $p = 0.05$. Coefficients of correlation *r* were over 0.997 (0.997–0.9997) and significantly different from zero ($t_{calc} > t_{crit}$) and greater than the critical value 0.707 for a bad linearity. Coefficients of determination R^2 were over 0.993 (0.993–0.9994) [38]. Linearity *L(%)* were over 95% (96.1–98.9%) and coefficients of variation of the slope $CV_b(\%)$ were less than 5% (1.12–3.91%).

Sensitivity and selectivity studies

The values obtained for *gamma* γ sensitivity (ESM Table S2) ranged from the less sensitive, such as 2.5 x 10^{-3} µL pg^{-1} for *pyrene* and dibenzo[*a,h*]anthracene, or 2.7-3.5 x 10^{-3} µL pg^{-1} for *fluoranthene*, *chrisene* and benzo[*ghi*]perilene, to the more sensitive benzo[*b*] and benzo[*k*]fluoranthene and naphthalene (1.3-1.4 x 10^{-2} µL pg^{-1}). In terms of *delta* δ sensitivity, the most sensitive were *naphthalene*, benzo[*b*] and benzo[*k*]fluoranthene (7.7-8.5 x 10^{-3}) against the less sensitive *pyrene* and *fluoranthene* (9-10 x 10^{-4}), or 1.8-2.1 x 10^{-3} for *acenaphthene*, *fluorene*, *chrysene*, *anthracene* and dibenzo[*a,h*]anthracene.

The *selectivity* study is necessary in complex matrices, such as filters of airborne particles, thus, it must to reduce interferences from real samples and contamination from blanks. Current analysis used selective chemicals that removed interferences and contamination, such as specific cartridges (miniSpe-ed) and solvents of chromatographic grade, and selective instruments, such a gas chromatograph with capillary columns, and a mass-spectrometry detector. In the case of co-eluting compounds the MS detector was set in the Selected Ion Monitoring mode (SIM). The

clean-up procedure used with miniSpe-ed removes aliphatic hydrocarbons and polar compounds. Besides, analysis of blanks also proved that analytical determinations were free from contaminants. In addition, possible contaminants were reduced by handling all format of samples inside the laminar air-flow cabinet INDELAB®, Model IDL-48 V with a HEPA filter plus a charcoal layer, which ensure a clean air inside the cabinet. Consequently, the methodology resulted free from chemical interferences or at least with interferences controlled.

Limits of detection and quantification

Limits of detection from blanks (LOD_{Bl}) were lower (4-7 times) than the instrument detection limits (LOD_I) and the others (Table 3) due to the purity of the chemicals used. Therefore, a more realistic alternative is a detection limit based on the signal-to-noise ratio (3 x *S/N*). As a result, values of the noise detection limits (LOD_N) were similar to the instrumental detection limits, that is, $LOD_{Bl} << LOD_I \cong LOD_N$.

Moreover, as the HR and MR ranges gave good linearity and all NZC standards were determined with accuracy (< 3% error), we focused on the low range (LR). Thus, at the lowest concentration obtained we injected 12 replicates of the reference and quantification and detection limits (LOD_{ZC}, one third of LOQ_{ZC}) were estimated.

In brief, final comparison resulted in $LOD_{Bl} < LOD_{ZC} < LOD_N \cong LOD_I$ and the selected as method detection limit (*MDL*) was the instrumental detection limit (LOD_I), expressed as pg μL^{-1} or as pg m^{-3} and ng g^{-1}. Values of Table 3 were lower than other study [39]. However, Piñeiro-Iglesias et al. [19] reported limits similar for anthracene but not for fluoranthene and pyrene, which were higher than those reported here.

Ruggedness study

The ruggedness of a procedure must be established for 'in−house' developed methods [34]. Seven factors (A - G) were tested along four chromatographic conditions:

1. Conditions of the AmSE (Accelerated mini-Soxhlet Extractor): extraction time of 45 and 75 minutes (A) were employed with a solvent volume of 20 and 30 mL of the solvent mixture (B). Nominal conditions were at 60 minutes for 25 mL.

2. Conditions of the SPE technique with miniSpe-ed cartridges: fraction volumes of 2.5 mL and 3.5 mL of methylene chloride (C) were collected at flow rates of elution of 0.35 mL min^{-1} and 0.65 mL min^{-1} (D). Nominal conditions were at 3 mL and 0.50 mL min^{-1}.

3. Conditions of the gas flow strength for the NASE (Nitrogen-Assisted Solvent Evaporation): nitrogen flows of 25 mL min^{-1} and 75 mL min^{-1} (E) were explored. Nominal conditions were at 50 mL min^{-1}.

4. Conditions of the chromatographic system: injection volume of 1.0 µL and 3.0 µL (F) of the sample extract were tested, performing analytes separation at a final injector temperature of 250 ºC and 270 ºC (G). Nominal conditions were injecting 2 µL and oven temperature of 260 ºC.

The factor variations are shown in Fig. 3. The deviations of chromatographic parameters were below 12.5% for negative deviations and 5% for positive ones. Therefore, SRM recoveries were always within the range of AOAC percentages of recovery. The only factor that most affects results is Factor A (Soxhlet extraction time) when is less than 60 minutes. Therefore, there were no significant differences in performance of the method as a result of the deliberate changes, implying that our developed method is robust in terms of reliability.

The complete optimised and validated method was (Fig. 4): 100 mg of one-half cutted quartz filter of airborne particles was extracted in the AmSE system with 25 mL of 5:1 acetone/methylene chloride for 60 minutes. After quick evaporation until 2 mL with NASE, the organic extract was purified by mSPE using a miniSpe-ed cartridge, eluting PAHs simultaneously to nitrogen evaporation with 3 mL of methylene chloride.

Conclusions

The proposed RSE-mSPE method for extraction of PAHs in atmospheric particles helps in minimizing solvent volume and time extraction. The two main stages optimised AmSE+mSPE improved the recovery and accuracy of PAHs determination. Accelerated mini-Soxhlet Extraction system (AmSE) reduced volume consuming in 90% (10 times), time consuming in 90% (10 times) and increased recoveries of SRM in 9%. Mini-Solid Phase Extraction (mSPE) also reduced time consuming in 90% (9 times) and increased recoveries of SRM in 3%. The other parameters of validation were successfully assessed with good agreement between certified and reference values, thereby no interferences being by the presence of quartz filters. Thus it can be stated that the proposed RSE-mSPE methodology is a simple, fast and low-cost method that can be used, combining with GC/MS-SIM, for the determination of PAHs in airborne particulate matter.

Supplementary Information The online version contains supplementary material available at https://doi.org/

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Compliance with ethical standards

Conflict of Interest The authors declare that they have no conflict of interest.

References

1. World Health Organization, WHO (2000) Air quality guidelines. 2nd edition. Chap. 5.9. pp. 92-96. Regional Office for Europe. WHO regional publications, European series, No. 91. Copenhagen (Denmark).
2. World Health Organization, WHO (2010) Polynuclear aromatic hydrocarbons. In: WHO Guidelines for indoor air quality. Selected Pollutants. Chap. 6. pp. 289-346. Regional Office for Europe. Copenhagen (Denmark).
3. European Commission, EC (2001) Ambient air pollution by Polycyclic Aromatic Hydrocarbons (PAH). Position Paper. Chap. 5. pp. 47-49. July 2001. Office for Official Publications of the European Communities, Luxembourg.
4. HBM4EU (2018) Scoping document on Prioritized substance group: PAHs and air pollutants. 1st edition. Chap. 2. pp. 12-16. December 2018. AUTH Chemical group leader. HBM4EU Project of the European Union's Horizon 2020 programme: Science and policy for a health future. German Environment Agency, Section II 1.2 Toxicology, Health Related Environmental Monitoring.
5. OSPAR Commission (2009) Background Document on Polycyclic Aromatic Hydrocarbons (PAHs), update 2009. Hazardous Substances Series. The Executive Secretary. London, United Kingdom.
6. CONCAWE (2001) Automotive emissions of polycyclic aromatic hydrocarbons. After Report 98/55. Vol. 10, No 1, April 2001. Environmental Science for European Refining. Division of the European Petroleum Refiners Association. CONCAWE, Brussels.
7. OSPAR Commission (2001) Best Environmental Practice (BEP) for the Reduction or Prevention of Emissions of Polycyclic Aromatic Hydrocarbons (PAHs) from

Domestic Combustion Appliances. OSPAR Background Document. The Executive Secretary. London, United Kingdom.

8. Nguyen TNT, Kwon GL, Jung KS, Lee SJ, Choi SD. Spatially high-resolved monitoring and risk assessment of polycyclic aromatic hydrocarbons in an industrial city. J. Haz. Mat. 2020;393:122409 (1-11).

9. Lee BK. Sources, Distribution and Toxicity of Polyaromatic Hydrocarbons (PAHs) in Particulate Matter, Air Pollution, Vanda Villanyi (Ed.), InTech. 2010.

10. Maliszewska B. Sources, Concentrations, Fate and Effects of Polycyclic Aromatic Hydrocarbons (PAHs) in the Environment, Review. Part A: PAHs in Air. Polish J. Environ. Stud. 1999;8:131-136.

11. Luo J, Han Y, Zhao Y, Huang Y, Liu X, Tao S, Liu J, Huang T, Wang L, Chen K, Ma J. Effect of northen boreal forest fires on PAH fluctuations across the artic. Environ. Poll. 2020;261:114186.

12. Galvao ES, Santos JM, Lima AT, Reis Jr, NC, D'Azeredo MT, Stuetz RM. Trends in analytical techniques applied to particulate matter characterization: A critical review of fundaments and applications, Chemosphere 2018;199:546-568.

13. Maurice AS, Ocampo R, Alleman L, Coddeville P. Tenax-TA Spiking Approach of Thermal Desorption Coupled to GC–MSMS for the Quantification of PAHs in Indoor Air and Dust, Pol. Arom. Comp. 2017;37:170-177.

14. Yuan, X., You, F., Yong, L., Yang, C., Zhu, L., Hu, B., Liu, T. Rapid determination of 16 polycyclic aromatic hydrocarbons in PM2.5 by microwave assisted extraction-high performance liquid chromatography, Microch. J. 2019;144:391–396.

15. Rodríguez P, Moreda A, Bermejo A, Bermejo P. Ultrasound-assisted solvent extraction (UASE) of total polycyclic aromatic hydrocarbons from mussels followed by spectrofluorimetric determination, Talanta 2005;66:683-690.

16. Oukebdane K, Portet F, Machour N, Dionnet F, Desbène PL. Comparison of hot Soxhlet and accelerated solvent extractions with microwave and supercritical fluid extractions for the determination of polycyclic aromatic hydrocarbons and nitrated derivatives strongly adsorbed on soot collected inside a diesel particulate filter. Talanta. 2010;82:227–236.

17. Walgraeve, C., Demeestere, K., Dewulf, J., Zimmermann, R., Van Langenhov, H Oxygenated polycyclic aromatic hydrocarbons in atmospheric particulate matter: Molecular characterization and occurrence. Atmos. Environ. 2010;44:1831-1846.

18. EPA, 2014. Priority pollutant list. From Toxic and Priority Pollutants Under the Clean Water Act, Washington, DC.

19. Piñeiro, M., López, P., Vázquez, E., Muniategui, S., Prada, D., Fernández, E. Microwave assisted extraction of polycyclic aromatic hydrocarbons from atmospheric particulate samples, Anal. Bioanal. Chem. 2000;367:29-34.

20. Gutiérrez, A., Fernández, A.J., Ternero, M., Fernández, F. Particle-size distribution of polycyclic aromatic hydrocarbons in the urban air in Southern of Spain. Anal. Bioanal, Chem. 2005;381:721-736.

21. Fernández-Espinosa AJ. Combining PLS regression with portable NIR spectroscopy to on-line monitor quality parameters in intact olives for determining optimal harvesting time, Talanta 2016;148:216–228.

22. EPA, 2014. Priority pollutant list. From Toxic and Priority Pollutants Under the Clean Water Act, Washington, DC.

23. EPA. 2018. Method 8270E (SW-846): Semivolatile Organic Compounds by Gas Chromatography/ Mass Spectrometry (GC/MS), Rev. 6, June 2018, Washington, DC.

24. Wiley/NIST (2017) Wiley Registry®, 11th Edition/NIST 2017, Mass Spectral Library, Mass Spectrometry Data, Mass Spectrometry Data Center. September 2017.

25. ISO (1994) 5725-1:1994. Accuracy (trueness and precision) of measurement methods and results — Part 1: General principles and definitions, ISO Geneva.

26. Karthikeyan S, Balasubramanian R, Wei S. Optimisation and validation of a low temperature microwave-assisted extraction method for analysis of polycyclic aromatic hydrocarbons in airborne particulate matter, Talanta. 2006;69:79-86.

27. AOAC International (2016) Guidelines for Standard method performance requirements. AOAC Official methods of analysis, Appendix F. Gaithersburg, MD.

28. Thompson M, Ellison SLR, Wood R. Harmonized Guidelines for Single-Laboratory Validation of Methods of Analysis, Pure Appl. Chem. 2002;74:835-855.

29. European Commission, EC (2019) SANCO/3030/99 rev. 5. Technical Active Substance and Plant protection products: Guidance for generating and reporting methods of analysis in support of pre- and post-registration data requirements for Annex II (Section 4) of Regulation (EU) No 283/2013 and Annex (Section 5) of Regulation (EU) No 284/2013. Guidance document. Directorate General Health and Consumer Protection. 22 March.

30. Thompson L. The Horwitz Function Revisited, J. AOAC Int. 1997;80:676-679.

31. Horwitz, W., Albert, R., Deutsch, M.J. Precision parameters of methods of analysis required for nutrition labeling, Part I. Major nutrients. J. AOAC Int. 1990;73:661-680.

32. Cuadros L, García AM, Bosque JM. Statistical estimation of linear calibration range, Anal. Lett. 1996;29:1231-1239.

33. ISO (2007) ISO11843-2:2007 Capability of detection–Part 2: Methodology in the linear calibration case, ISO Geneva.

34. EPA (2017) Title 40. Protection of Environment. Chapter I. EPA. Subchapter D. Water programs. Part 136 – Guidelines establishing test procedures for the analysis of pollutants. Appendix B to Part 136–Definition and procedure for the determination of the method detection limit, Revision 2, 28 August. Washington, DC.

35. Magnusson, B., Ornemark, U. Eurachem Guide: The Fitness for Purpose of Analytical methods–A Laboratory Guide to Method Validation and Related Topics. 2nd Edition, Eurachem. 2014.

36. Youden, W.J., Steiner, E.H. Statistical Manual of the AOAC- Statistical techniques for collaborative tests (by) W.J. Youden. Planning and analysis of results of collaborative tests (by) E.H. Steiner. In: AOAC International (Ed.). Washington D.C., U.S.A., 1975;33–41.

37. Boström CE, Gerde P, Hanberg A, Jernström B, Johansson C, Kyrklund T, Rannug A, Törnqvist M, Victorin K, Westerholm R. Cancer risk assessment, indicators and guidelines for polycyclic aromatic hydrocarbons in the ambient air. Special Report. Environ. Health Perspect. 2002; 110:451-488.

38. Li, T., Wang, Y., Hou, J., Zheng, D., Wang, G., Hu, C., Xu, T., Cheng, J., Yin, W., Mao, X., Wang, L., He, Z., Yuan, J. Associations between inhaled doses of PM2.5-

bound polycyclic aromatic hydrocarbons and fractional exhaled nitric oxide. Chemosphere 2019;218:992-1001.

39. Resende, R., de Lourdes, Z., Helvécio, C., Menezes, C. Phase distribution of polycyclic aromatic hydrocarbons and their oxygenated and nitrated derivatives in the ambient air of a Brazilian urban area. Chemosphere 2020;250:126223 (1-11).

Quartz filters + 100 mg of SRM

Recovery standard (Pyrene-d10)

Accelerated Mini-Soxhlet Extraction (AMSE)-60 min.

Acetone:Methylene chloride 5:1 v/v, 25 mL

Solvent reduction (2 mL) Vaccum manifold + N_2 (NASE)

Silica-gel cartridges (Mini*Spe-ed*)

Solid Phase Extraction (SPE)

4 deuterated Internal Standards

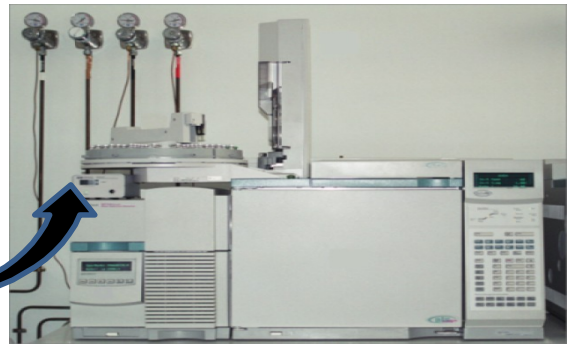
n-hexane 3 mL

Methylene chloride 3 mL

Aliphatic fraction

Aromatic fraction

Quantification of PAHs by GC-MS (SIM mode)



Captions of figures

Fig. 1. Recoveries obtained during the extraction stage. Comparison of the 250/100 mL Soxhlet apparatus with the 25 mL Quickfit® System using SRM 1649a.

Fig. 2. Comparison of the recoveries of SRM 1649a between open-column adsorption liquid chromatography and solid phase extraction techniques.

Fig. 3. Results of the ruggedness study. Changes on SRMs recoveries by variation of factors. A: AMSE extraction time; B: solvent extraction volume; C: volume of methylene chloride in SPE; D: flow rate of elution; E: nitrogen flow of NASE; F: injection volume of the extract; G: injector temperature of GC.

Fig. 4. Analytical methodology validated with two SRMs for PAHs determination.

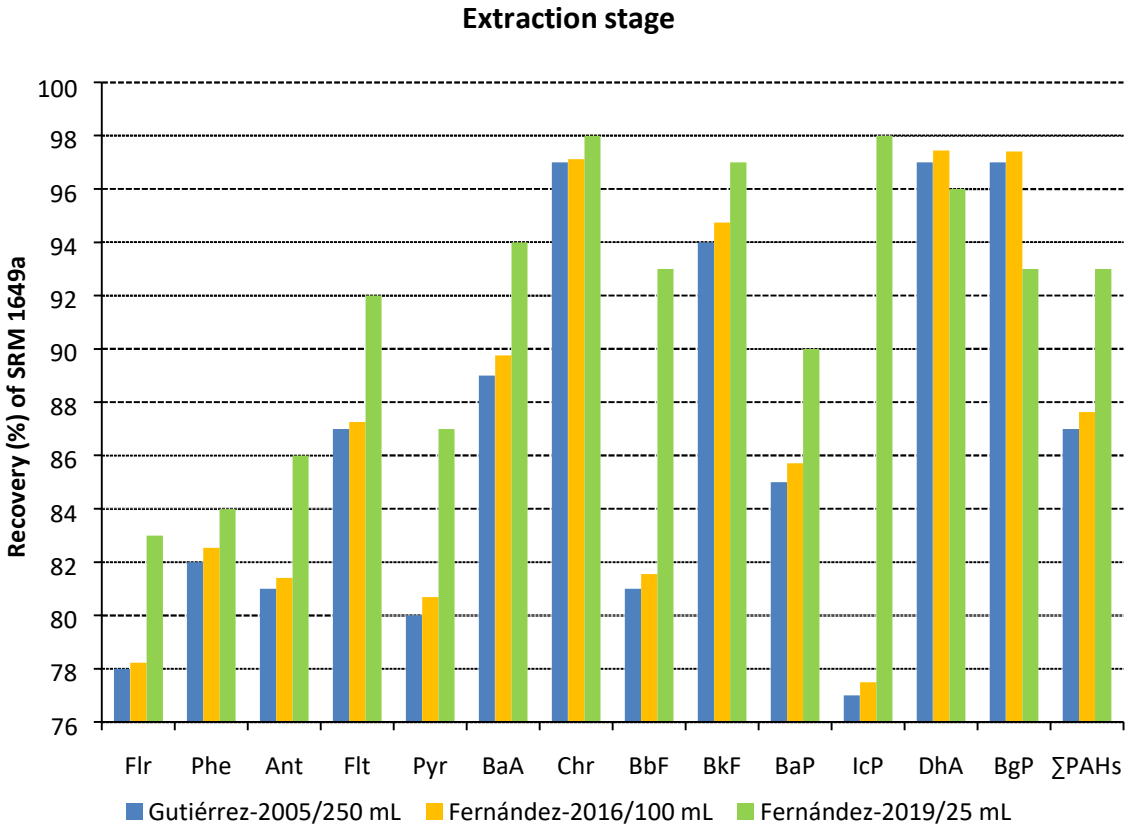

Extraction stage
Recovery (%) of SRM 1649a
100
98
96
94
92
90
88
86
84
82
80
78
76
Flr
Phe
Ant
Flt
Pyr
BaA
Chr
BbF
BkF
BaP
IcP
DhA
BgP
∑PAHs
Gutiérrez-2005/250 mL
Fernández-2016/100 mL
Fernández-2019/25 mL

Fig. 1

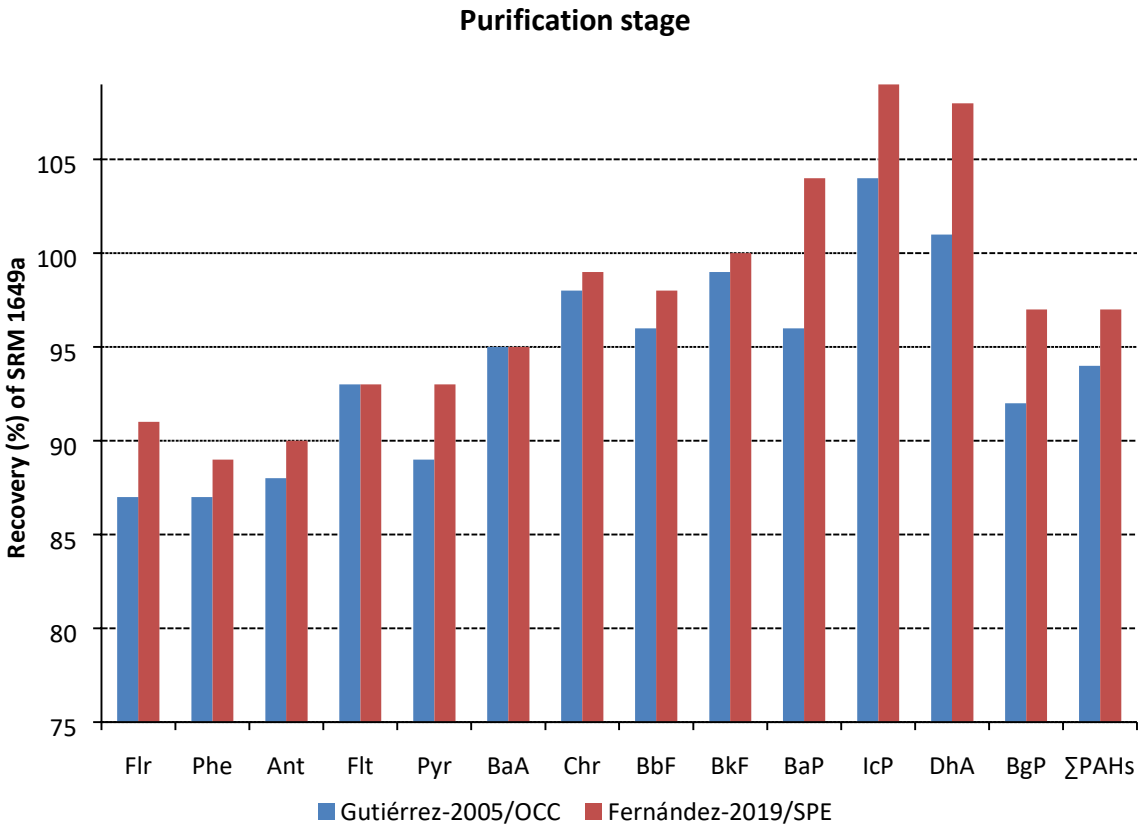

Purification stage
Recovery (%) of SRM 1649a
105
100
95
90
85
80
75
Flr
Phe
Ant
Flt
Pyr
BaA
Chr
BbF
BkF
BaP
IcP
DhA
BgP
∑PAHs
Gutiérrez-2005/OCC
Fernández-2019/SPE

Fig. 2

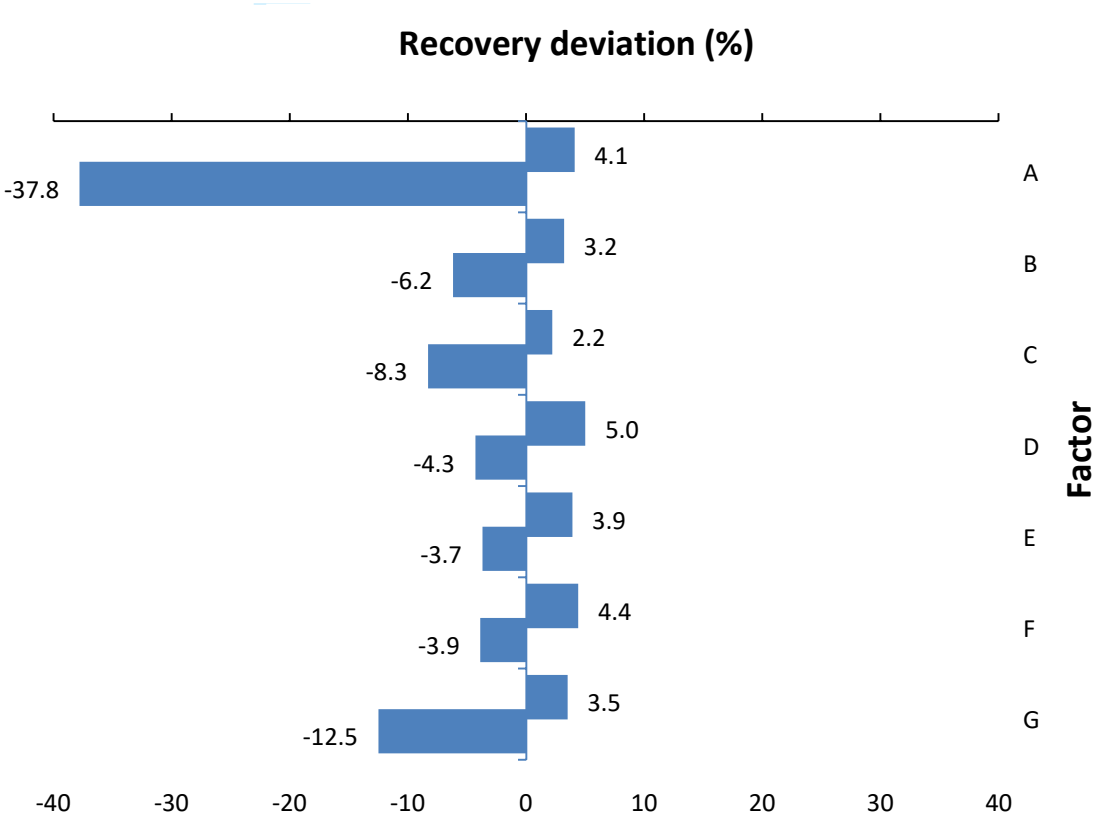

Recovery deviation (%)
4.1
-37.8
3.2
-6.2
2.2
-8.3
5.0
-4.3
3.9
-3.7
4.4
-3.9
3.5
-12.5
A
B
C
D
E
F
G
Factor
-40
-30
-20
-10
0
10
20
30
40

Fig. 3

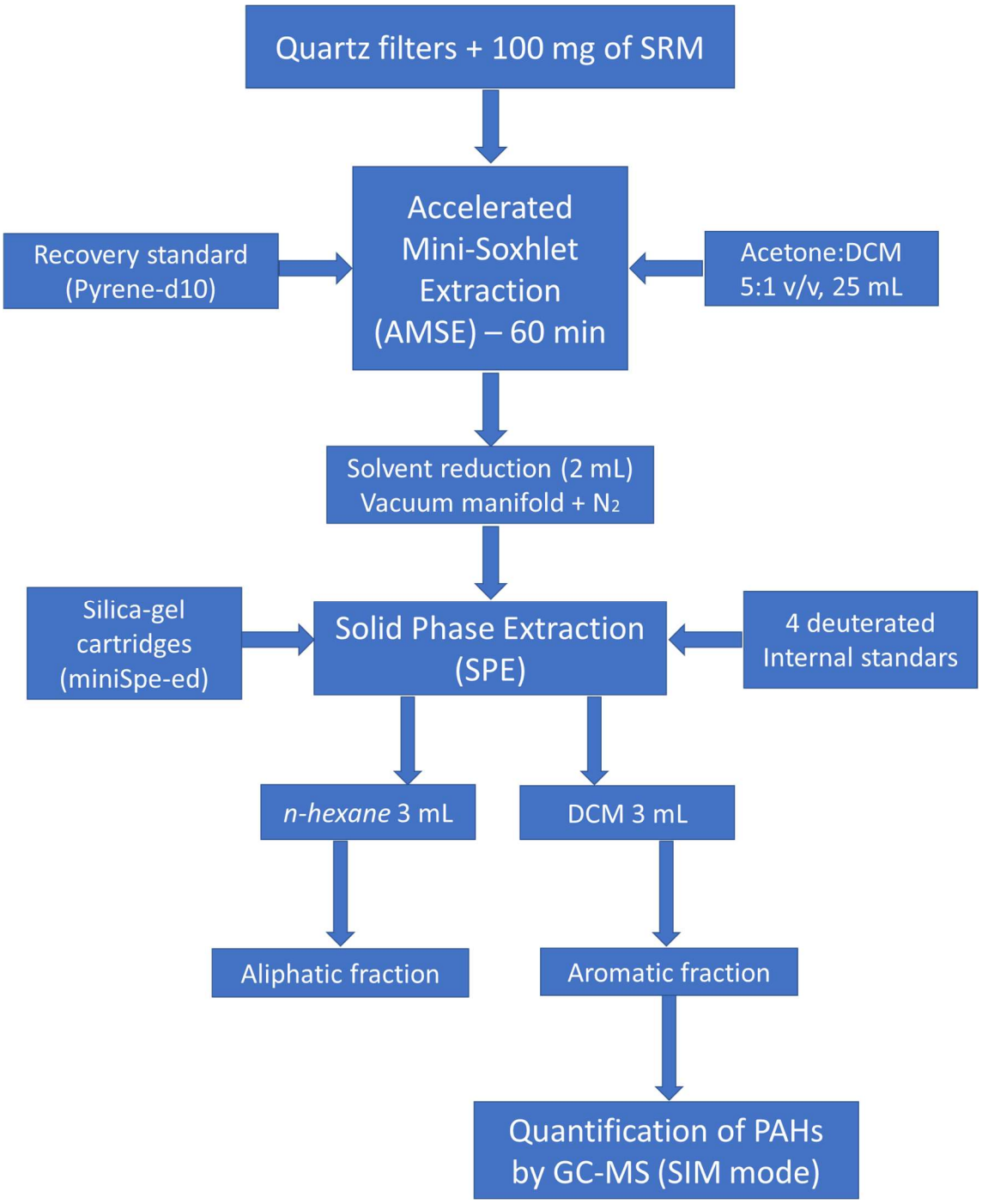

Quartz filters + 100 mg of SRM
Recovery standard (Pyrene-d10)
Accelerated Mini-Soxhlet Extraction (AMSE) – 60 min
Acetone:DCM 5:1 v/v, 25 mL
Solvent reduction (2 mL) Vacuum manifold + N2
Silica-gel cartridges (miniSpe-ed)
Solid Phase Extraction (SPE)
4 deuterated Internal standars
n-hexane 3 mL
DCM 3 mL
Aliphatic fraction
Aromatic fraction
Quantification of PAHs by GC-MS (SIM mode)

Fig. 4

Table 1 Results of the recovery study using both NIST 1649a and 1648a certified values in mg kg^{-1} (mean+standard deviation) for n = 12 replicates.

	Certified values in SRMs		Experimental values (with filter)			Experimental values (without filter)			
	Concentration (mg kg^{-1}+sd)	RSD (%)	Concentration (mg kg^{-1}+sd)	RSD (%)	Rec (%)	Concentration (mg kg^{-1}+sd)	RSD (%)	Rec (%)	AOAC-Rec
PAH NIST 1649a									
Fluorene*	0.23 ± 0.05	21.7	0.19 ± 0.03	15.8	*82.6*	0.21 ± 0.02	9.5	91.3	80-110
Phenanthrene	4.1 ± 0.4	8.9	3.9 ± 0.3	6.6	95.2	4.0 ± 0.2	5.8	96.4	80-110
Anthracene	0.43 ± 0.08	19.0	0.44 ± 0.05	10.7	101.9	0.43 ± 0.06	14.3	98.6	80-110
Fluoranthene	6.4 ± 0.2	2.8	6.3 ± 0.2	3.3	98.1	6.4 ± 0.3	3.9	98.9	80-110
Pyrene	5.3 ± 0.2	4.7	5.2 ± 0.3	6.2	97.4	5.2 ± 0.2	4.0	98.9	80-110
Benzo[*a*]anthracene	2.21 ± 0.07	3.3	2.15 ± 0.11	5.1	97.2	2.1 ± 0.3	11.9	95.2	80-110
Chrysene	3.05 ± 0.06	2.0	2.98 ± 0.10	3.2	98.0	2.90 ± 0.09	3.0	95.0	80-110
Benzo[*b*]fluoranthene	6.4 ± 0.6	9.9	6.1 ± 0.4	6.7	95.2	6.0 ± 0.3	5.7	92.7	80-110
Benzo[*k*]fluoranthene	1.91 ± 0.03	1.6	1.94 ± 0.05	2.5	101.5	1.92 ± 0.07	3.5	100.3	80-110
Benzo[*a*]pyrene	2.51 ± 0.09	3.5	2.46 ± 0.14	5.7	98.0	2.47 ± 0.12	4.9	98.4	80-110
Indeno[1,2,3-*cd*]pyrene	3.2 ± 0.7	22.6	3.4 ± 0.3	8.8	106.9	3.3 ± 0.3	8.6	102.8	80-110
Dibenzo[*a,h*]anthracene	0.29 ± 0.02	8.0	0.27 ± 0.04	13.8	93.1	0.26 ± 0.02	6.8	91.3	80-110
Benzo[*ghi*]perylene	4.0 ± 0.9	22.7	4.3 ± 0.3	7.0	107.2	4.1 ± 0.4	10.0	102.7	80-110
16-PAHs Sum**	40.2 ± 1.4	46.5	39.7 ± 0.8	30.1	97.9	39.3 ± 0.8	28.3	97.1	90-107
PAH NIST 1648a									
Naphthalene*	1.2 ± 0.6	47.2	1.0 ± 0.4	41.6	*82.1*	1.1 ± 0.3	30.4	91.1	80-110
Acenaphthylene*	0.173 ± 0.012	6.9	0.13 ± 0.02	13.0	*75.7*	0.148 ± 0.011	7.4	85.5	80-110
Acenaphthene*	0.25 ± 0.08	33.2	0.19 ± 0.06	28.9	*76.0*	0.23 ± 0.03	14.3	92.0	80-110
Fluorene*	0.25 ± 0.04	13.9	0.23 ± 0.03	11.3	91.6	0.23 ± 0.03	11.3	91.6	80-110
Phenanthrene	4.9 ± 0.2	3.5	5.0 ± 0.2	4.8	101.9	4.9 ± 0.2	4.5	100.6	80-110
Anthracene*	0.459 ± 0.013	2.8	0.47 ± 0.02	3.4	101.3	0.45 ± 0.02	4.7	96.9	80-110
Fluoranthene	8.07 ± 0.14	1.7	8.00 ± 0.2	1.9	99.0	7.25 ± 0.11	1.5	89.8	80-110
Pyrene	5.88 ± 0.07	1.2	5.82 ± 0.10	1.7	99.0	5.91 ± 0.12	2.0	100.5	80-110
Benzo[*a*]anthracene	2.7 ± 0.2	5.5	2.6 ± 0.2	8.5	95.2	2.7 ± 0.3	10.5	98.2	80-110
Chrysene	6.1 ± 0.1	1.0	6.07 ± 0.10	1.5	99.2	6.180 ± 0.011	0.2	101.0	80-110
Benzo[*b*]fluoranthene*	8.89 ± 0.05	0.6	9.25 ± 0.04	0.4	104.0	9.13 ± 0.13	1.4	102.7	80-110
Benzo[*k*]fluoranthene	3.0 ± 0.2	7.9	2.9 ± 0.3	10.2	94.1	2.9 ± 0.2	8.2	96.4	80-110
Benzo[*a*]pyrene	2.57 ± 0.10	3.9	2.49 ± 0.13	5.2	96.9	2.5 ± 0.2	6.0	96.5	80-110
Indeno[1,2,3-*cd*]pyrene	4.2 ± 0.2	4.1	4.3 ± 0.3	5.8	103.4	4.1 ± 0.2	4.2	97.1	80-110
Dibenzo[*a,h*]anthracene	0.4 ± 0.2	35.7	0.4 ± 0.2	47.7	104.8	0.40 ± 0.08	20.0	95.2	80-110
Benzo[*ghi*]perylene	5.0 ± 0.2	3.6	5.2 ± 0.3	5.4	103.2	5.0 ± 0.2	4.4	100.8	80-110
16-PAHs Sum**	54.1 ± 0.8	40.4	54.0 ± 0.8	52.1	96.1	53.0 ± 0.7	28.8	96.9	90-107

*: Reference values, not certified. SRM 1649a and 1648a; **: Values of RSD(%) in the 16-PAHs sum was extended from the individual ones.

Table 2 Results of the *F*-tests and *t*-tests using both NIST SRM 1649a and 1648a for n = 12 replicates. Results of the precision study under reproducibility conditions (R) by Hortwitz and AOAC criteria.

	RSD_R	AOAC-$ERSD_R$	$PRSD_R$	HorRat	F_{calc}	F_{crit}	t_{calc}	t_{crit}
PAH NIST 1649a								
Fluorene*	15.8	30	20.5	0.77	2.78	3.24	*2.50*	2.04
Phenanthrene	6.6	14.6	13.0	0.51	2.03	3.24	1.64	2.04
Anthracene	10.7	22	18.1	0.59	3.04	3.24	0.31	2.04
Fluoranthene	3.3	14.6	12.1	0.27	1.36	2.76	1.72	2.04
Pyrene	6.2	14.6	12.5	0.50	1.64	2.76	1.38	2.04
Benzo[*a*]anthracene	5.1	22	14.3	0.36	2.27	2.76	1.89	2.04
Chrysene	3.2	14.6	13.6	0.24	2.61	2.76	2.00	2.04
Benzo[*b*]fluoranthene	6.7	14.6	12.2	0.55	2.44	3.24	1.50	2.04
Benzo[*k*]fluoranthene	2.5	22	14.5	0.17	2.50	2.76	1.99	2.04
Benzo[*a*]pyrene	5.7	22	14.0	0.41	2.63	2.76	1.22	2.04
Indeno[1,2,3-*cd*]pyrene	8.8	14.6	13.3	0.66	***5.76***	3.24	1.20	2.05
Dibenzo[*a,h*]anthracene	13.8	30	19.5	0.71	2.59	2.76	1.89	2.04
Benzo[*ghi*]perylene	7.0	14.6	12.8	0.54	*9.20*	3.24	1.31	2.06
16-PAHs Sum	2.0	10.6	9.2	0.22	3.24	3.24	1.17	2.05
PAH NIST 1648a								
Naphthalene*	*41.6*	22	16.0	*2.60*	1.91	3.24	1.14	2.04
Acenaphthylene*	13.0	30	21.7	0.60	2.01	2.76	*8.19*	2.04
Acenaphthene*	28.9	30	20.5	*1.41*	2.28	3.24	***2.22***	2.04
Fluorene*	11.3	30	20.0	0.57	1.81	2.76	1.80	2.04
Phenanthrene	4.8	14.6	12.6	0.39	1.99	2.76	1.24	2.04
Anthracene*	3.4	22	18.0	0.19	1.51	2.76	1.16	2.04
Fluoranthene	1.9	14.6	11.7	0.16	1.15	2.76	1.52	2.04
Pyrene	1.7	14.6	12.3	0.14	2.04	2.76	2.00	2.04
Benzo[*a*]anthracene	8.5	22	13.9	0.61	2.15	2.76	1.99	2.04
Chrysene	1.5	14.6	12.2	0.12	2.25	2.76	1.89	2.04
Benzo[*b*]fluoranthene*	0.4	14.6	11.4	0.04	1.56	2.76	*21.16*	2.04
Benzo[*k*]fluoranthene	10.2	22	13.7	0.74	1.46	2.76	1.90	2.04
Benzo[*a*]pyrene	5.2	22	13.9	0.37	1.69	2.76	1.96	2.04
Indeno[1,2,3-*cd*]pyrene	5.8	14.6	12.8	0.45	2.16	2.76	1.89	2.04
Dibenzo[*a,h*]anthracene	*47.7*	14.6	18.1	*2.64*	1.96	2.76	0.31	2.04
Benzo[*ghi*]perylene	5.4	14.6	12.5	0.43	2.42	2.76	1.97	2.04
16-PAHs Sum	1.5	10.6	8.8	0.14	1.85	2.76	0.66	2.04

* Reference values, not certified. SRM 1649a and 1648a; the underlining of some values represents the compounds that exceeded the AOAC limits

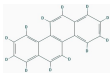
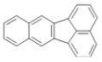
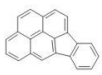
RSD_R: Relative standard deviation under reproducibility conditions; AOAC-$ERSD_R$: Expected AOAC values of RSD_R; $PRSD_R$: Predicted RSD_R values; HorRat: Horwitz Ratio

Table 3. Limits of detection (LODs, pg μL^{-1}) for PAHs. Estimations for instrument detection limit (LOD_I, n =12), blanks detection limit (LOD_{Bl}, n =8), noise detection limit (LOD_N, n =12) zero concentration detection limit (LOD_{ZC}, n =12) and method detection limit (*MDL*) expressed as concentration in the ambient air (MDL_A, pg m^{-3}) and in the solid particulate matter (MDL_B, ng g^{-1}).

HAP	LOD_I	Sd	LOD_{Bl}	sd	LOD_N	Sd	LOD_{ZC}	sd	MDL_A	sd	MDL_B	sd
Naphthalene	0.215	0.011	0.048	0.005	0.189	0.009	0.138	0.005	1.05	0.06	4.3	0.2
Acenaphthylene	0.061	0.004	—		0.036	0.002	0.016	0.003	0.30	0.02	1.22	0.08
Acenaphthene	0.187	0.011	0.029	0.004	0.151	0.004	0.109	0.005	0.92	0.05	3.7	0.2
Fluorene	0.148	0.009	—		0.122	0.007	0.007	0.005	0.72	0.04	2.9	0.2
Phenanthrene	0.222	0.006	0.075	0.002	0.198	0.003	0.145	0.003	1.09	0.03	4.44	0.11
Anthracene	0.152	0.006	0.021	0.002	0.121	0.002	0.075	0.002	0.74	0.03	3.04	0.11
Fluoranthene	0.251	0.006	0.076	0.003	0.228	0.005	0.174	0.003	1.23	0.03	5.02	0.11
Pyrene	0.35	0.02	0.17	0.02	0.31	0.02	0.274	0.009	1.72	0.10	7.0	0.4
Benzo[*a*]anthracene	0.093	0.005	—		0.070	0.003	0.015	0.005	0.45	0.02	1.85	0.11
Chrysene	0.209	0.013	0.076	0.009	0.177	0.003	0.132	0.003	1.02	0.06	4.2	0.2
Benzo[*b*]fluoranthene	0.044	0.003	—		0.017	0.004	0.009	0.004	0.213	0.013	0.87	0.05
Benzo[*k*]fluoranthene	0.056	0.002	—		0.031	0.003	0.021	0.003	0.273	0.010	1.12	0.04
Benzo[*a*]pyrene	0.110	0.007	—		0.071	0.005	0.032	0.002	0.54	0.03	2.19	0.13
Indeno[1,2,3-*cd*]pyrene	0.102	0.006	—		0.061	0.003	0.025	0.002	0.50	0.03	2.04	0.12
Dibenzo[*a,h*]anthracene	0.100	0.006	—		0.062	0.005	0.023	0.003	0.49	0.03	2.00	0.11
Benzo[*ghi*]perylene	0.122	0.005	—		0.099	0.005	0.044	0.002	0.59	0.02	2.43	0.10

—: Values of LOD_{Bl} could not be estimated due to the absence of the compounds in the procedural blanks;

Table S1 Physical properties of the 16-PAHs and chromatographic operative conditions.

PAH (formula)	Rings		Vapour pressure (Pa, 25ºC)	RT* (min)	RRT	Target (T)+ Qualifier (Q)	SIM frequency (seg^{-1})	Internal standard
Naphthalene ($C_{10}H_8$)	2		8.6	9.32	0.77	128.17	8.33	Acenaphthene-d10
Acenaphthylene ($C_{12}H_8$)	3		$9.0x10^{-1}$	11.83	0.98	152.19	1.72	Acenaphthene-d10
Acenaphthene-d10 ($C_{12}H_{10}D_{10}$)	3		N/A	12.10		164.21+162	1.72	
Acenaphthene ($C_{12}H_{10}$)	3		$3.0x10^{-1}$	12.17	1.01	154.21+153	1.72	Acenaphthene-d10
Fluorene ($C_{13}H_{10}$)	3		$9.0x10^{-2}$	13.41	1.11	166.22+165	4.26	Acenaphthene-d10
Phenanthrene-d10 ($C_{14}D_{10}$)	3		N/A	16.30		188.29	4.26	
Phenanthrene ($C_{14}H_{10}$)	3		$2.0x10^{-2}$	16.38	1.01	178.23	4.26	Phenanthrene-d10
Anthracene ($C_{14}H_{10}$)	3		$1.0x10^{-3}$	16.56	1.01	178.23	4.26	Phenanthrene-d10
Fluoranthene ($C_{16}H_{10}$)	4		$1.2x10^{-3}$	21.11	0.75	202.25	4.26	Chrysene-d12
Pyrene-d10 ($C_{16}D_{10}$)	4		N/A	22.05	0.79	212.25	4.26	Chrysene-d12
Pyrene ($C_{16}H_{10}$)	4		$6.0x10^{-4}$	22.13	0.79	202.25	4.26	Chrysene-d12
Benzo[*a*]anthracene ($C_{18}H_{12}$)	4		$2.8x10^{-5}$	27.93	1.00	228.29	4.26	Chrysene-d12
Chrysene-d12 ($C_{18}D_{12}$)	4		N/A	27.98		240.36	4.26	
Chrysene ($C_{18}H_{12}$)	4		$5.7x10^{-7}$	28.13	1.00	228.29	4.26	Chrysene-d12
Benzo[*b*]fluoranthene ($C_{20}H_{12}$)	5		$5.0x10^{-7}$	35.04	0.91	252.31	4.26	Perylene-d12
Benzo[*k*]fluoranthene ($C_{20}H_{12}$)	5		$5.2x10^{-8}$	35.27	0.92	252.31	4.26	Perylene-d12
Benzo[*a*]pyrene ($C_{20}H_{12}$)	5		$5.5x10^{-9}$	37.75	0.98	252.31	4.26	Perylene-d12
Perylene-d12 ($C_{20}D_{12}$)	5		N/A	38.34		264.17	4.26	
Indeno[1,2,3-*cd*]pyrene ($C_{22}H_{12}$)	6		$1.3x10^{-10}$	50.60	1.32	276.33	4.26	Perylene-d12
Dibenzo[*ah*]anthracene ($C_{22}H_{14}$)	5		$3.7x10^{-10}$	51.09	1.33	278.35	4.26	Perylene-d12
Benzo[*ghi*]perylene ($C_{22}H_{12}$)	6		$1.0x10^{-10}$	54.39	1.42	276.33	4.26	Perylene-d12

*: PAHs list was ordered in this table by retention time in the chromatogram

Table S2 Goodness of the linearity (t_{calc}) and other linearity parameters (*r* to *CV*) and sensitivity (γ to s_b) for each PAH measured in the calibration curves*

HAP	r	Crit.Val.	tcalc	tcrit	R^2	L(%)	CV(%)	γ (slope b)	δ	S_b
Naphthalene	0.9996	0.707	86.6	2.45	0.9992	98.8	1.16	0.0136	0.0085	0.00016
Acenaphthylene	0.9996	0.707	86.6	2.45	0.9992	98.8	1.19	0.0088	0.0035	0.00010
Acenaphthene	0.9996	0.707	86.6	2.45	0.9992	98.8	1.19	0.0056	0.0018	0.00007
Fluorene	0.9995	0.707	77.4	2.45	0.9990	98.7	1.33	0.0047	0.0018	0.00006
Phenanthrene	0.9995	0.707	77.4	2.45	0.9990	98.7	1.32	0.0053	0.0024	0.00007
Anthracene*	0.9993	0.707	65.4	2.45	0.9986	98.5	1.50	0.0048	0.0020	0.00007
Fluoranthene	0.9995	0.707	77.4	2.45	0.9990	98.7	1.27	0.0027	0.0010	0.00003
Pyrene	0.9996	0.707	86.6	2.45	0.9992	98.8	1.22	0.0025	0.0009	0.00003
Benzo[*a*]anthracene	0.9993	0.707	65.4	2.45	0.9986	98.5	1.52	0.0050	0.0029	0.00008
Chrysene	0.9996	0.707	86.6	2.45	0.9992	98.9	1.12	0.0029	0.0018	0.00003
Benzo[*b*]fluoranthene*	0.9991	0.707	57.7	2.45	0.9982	97.5	2.49	0.0126	0.0077	0.00031
Benzo[*k*]fluoranthene	0.9996	0.707	86.6	2.45	0.9992	97.8	2.18	0.0142	0.0082	0.00031
Benzo[*a*]pyrene	0.9997	0.707	100.0	2.45	0.9994	97.9	2.12	0.0085	0.0059	0.00018
Indeno[1,2,3-*cd*]pyrene	0.9991	0.707	57.7	2.45	0.9982	97.5	2.54	0.0055	0.0060	0.00014
Dibenzo[*a,h*]anthracene	0.9994	0.707	70.7	2.45	0.9988	96.1	3.91	0.0025	0.0021	0.00010
Benzo[*ghi*]perylene	0.9966	0.707	29.6	2.45	0.9932	97.8	2.19	0.0035	0.0034	0.00008

**: Note that parameters were obtained from the relative analytical signal area analyte/area internal standard*

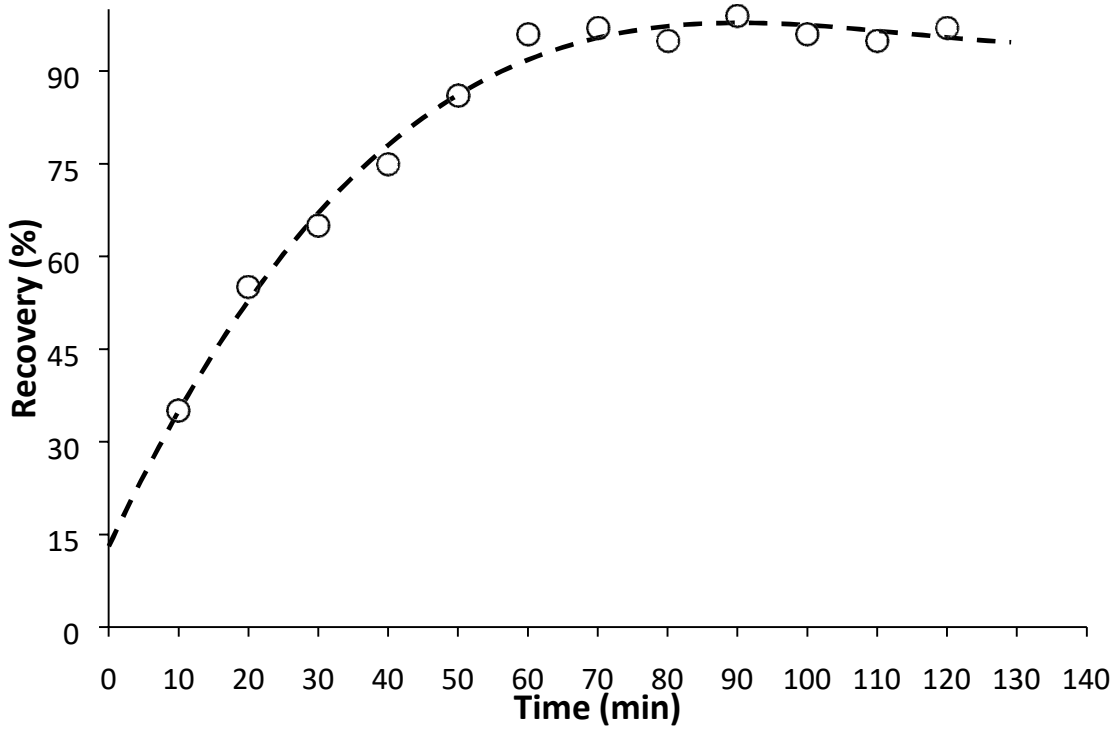

Recovery (%)
0
15
30
45
60
75
90
0
10
20
30
40
50
60
70
80
90
100
110
120
130
140
Time (min)

Fig. S1

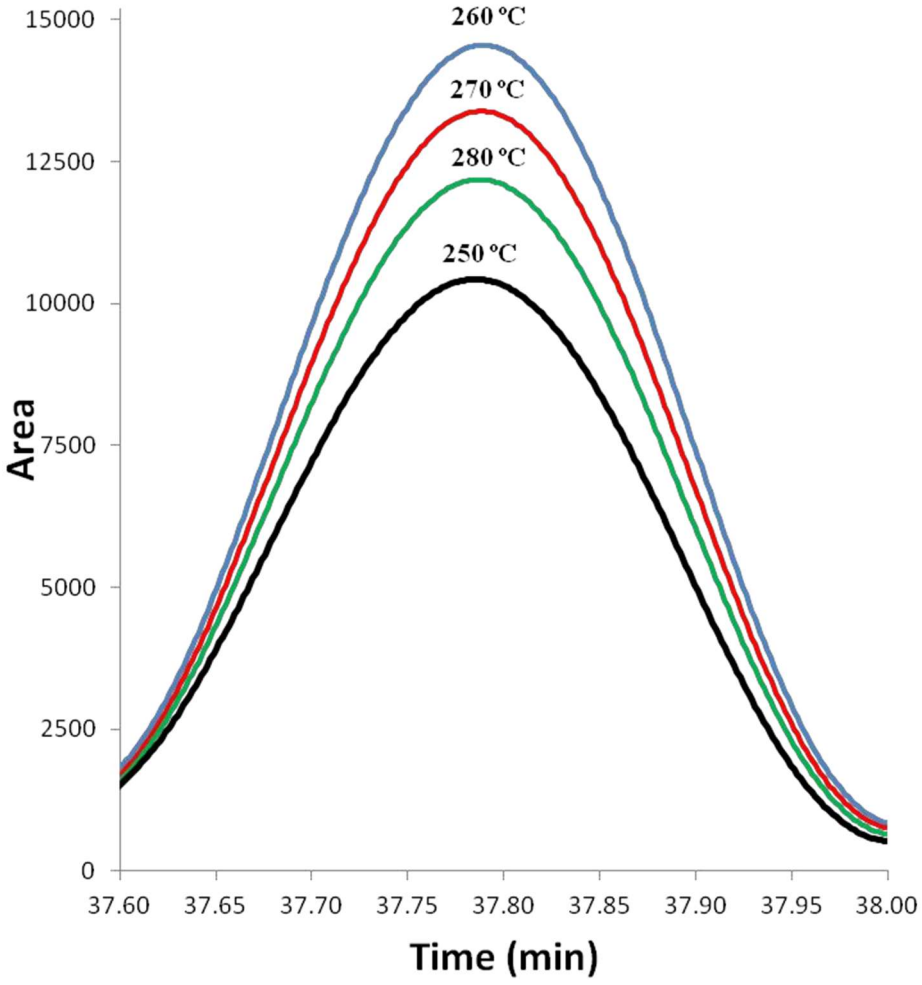

260 °C
270 °C
280 °C
250 °C
15000
12500
10000
7500
5000
2500
0
Area
37.60
37.65
37.70
37.75
37.80
37.85
37.90
37.95
38.00
Time (min)

Fig. S2