1 2 3	This is an Accepted Manuscript of an article published by Elsevier GmbH - Urban und Fischer in Journal of Trace Elements in Medicine and Biology on Epub 2022 Diciembre available at: DOI: 10.1016/j.jtemb.2022.127115
4	
5	
6	
7	Selenium, selenoproteins and cancer of the thyroid
8	
9	Rui Manuel Rua ^a *, Fátima Nogales ^b , Olimpia Carreras ^b , María Luisa Ojeda ^b
10	
11 12	^a Faculty of Health Sciences, University Fernando Pessoa, 4249-004 Porto, Portugal; <u>ruirua@ufp.edu.pt</u>
13 14	^b Department of Physiology, Faculty of Pharmacy, Seville University, 41012 Seville, Spain; <u>fnogales@us.es</u> (F.N); <u>olimpia@us.es</u> (O.C.); <u>ojedamuri11@us.es</u> (M.L.O)
15	
16	*Correspondence: ruirua@ufp.edu.pt Tel.: +351- 22 -507-46 30
17	
 18 19 20 21 22 23 24 25 26 27 28 29 30 31 	
32 33 34 35 36 37 38 39 40 41	Abstract: Selenium is an essential mineral element with important biological functions for the whole body through incorporation into selenoproteins. This element is highly concentrated in the thyroid gland. Selenoproteins provide antioxidant protection for this tissue against the oxidative stress caused by free radicals and contribute, via iodothyronine deiodinases, to the metabolism of thyroid hormones. It is known that oxidative stress plays a major role in carcinogenesis and that in recent decades there has been an increase in the incidence of thyroid cancer. The anti-carcinogenic action of selenium, although not fully understood, is mainly attributable to selenoproteins antioxidant properties, and to the ability to modulate cell proliferation (cell cycle and apoptosis), energy metabolism, and cellular immune response, significantly altered

during tumorigenesis. Researchers have suggested that different forms of seleniumsupplementation may be beneficial in the prevention and treatment of thyroid cancer;

44 however, the studies have several methodological limitations. This review is a summary

45 of the current knowledge on how selenium and selenoproteins related to thyroid cancer.

- 46 **Keywords**: selenium; selenoproteins; thyroid cancer; oxidative stress; supplementation
- 47

48 **1. Introduction**

Selenium (Se) is a critical microelement that was discovered and isolated for the first time in 1817 by Swedish chemist Jöns Jacob Berzelius [1]. While not an essential nutrient for plants, it is an essential nutrient for humans and many other life forms [2, 3]. In tissues, Se forms part of the amino acids selenomethionine and selenocysteine, with the latter being responsible for the main known biological activity of selenoproteins [4].

The thyroid gland is the organ in the human body with the highest Se content per unit of tissue [5,6]. In it, selenoproteins play a crucial role in the cellular defence system against hydrogen peroxide (H_2O_2) and other reactive oxygen species (ROS) [7,8]. The overproduction of free radicals, which triggers oxidative stress (OS), has been associated with several diseases and with cancer in particular [9-11].

60 Thyroid cancer is the most prevalent malignant neoplasm of the endocrine system and 61 its incidence has increased worldwide over the last four decades [12]. Histologically, 62 there are three main types of thyroid cancer: differentiated thyroid carcinoma, anaplastic thyroid carcinoma and medullary thyroid carcinoma. Differentiated thyroid carcinoma 63 accounts for about 95% of thyroid cancers and it originates from follicular thyroid cells, 64 which are responsible for hormone production. This cancer can be subdivided into 65 papillary, follicular and Hurthle cell carcinoma. The first of these is the most common 66 and has the best prognosis [13]. Papillary thyroid cancer invades the lymph nodes, 67 spreading to the cervical lymph nodes and also, less frequently, to other distant sites 68 such as the lungs [14]. This pattern of dissemination is important and can be a 69 presenting symptom of papillary carcinoma because the primary tumour is very small in 70 some cases. When they are less than 1 cm they are often referred to as microcarcinomas 71 [15]. Conversely, in the follicular form, haematogenous metastases are more frequent, 72 mainly affecting the lungs and bones [14]. Hurthle cell carcinoma is follicular in origin, 73 74 with at least 75% of the cells being Hurthle cells and having capsular and/or vascular 75 invasion [16]. The Hurthle cell is characterized cytologically as a large cell with 76 abundant eosinophilic, granular cytoplasm, and a large hyperchromatic nucleus with a prominent nucleolus. Cytoplasmic granularity is due to the presence of numerous 77 mitochondria [17]. Hurthle cell carcinoma is poorly avid to radioiodine and poorly 78 responsive to chemotherapy and radiation [18]. Hurthle cell carcinoma is believed to be 79 more aggressive than common follicular carcinoma [16]. 80

Since the thyroid is specially high in Se, and it plays an important role in this gland, the
relationship of Se with the incidence of thyroid cancer has been extensively studied
[11,19,20, 21, 22]. Thus, this review primarily aims to outline the current knowledge on
the association between Se, selenoproteins and thyroid cancer.

85 2. Selenium, selenoproteins and thyroid homeostasis

Adequate Se nutrition supports the synthesis and metabolism of thyroids hormones (THs) and protects the thyroid gland from damage from overexposure to iodide which increases OS [23]. Se is thus considered to be the second most important element in thyroid metabolism after iodine, which plays a beneficial role by forming part of different antioxidant selenoproteins [19].

91 There are 25 different selenoproteins in the human body with at least one selenocysteine 92 (Sec) amino acid in their structure [24,25]. Their difference in Sec incorporation 93 efficiency leads to a "selenoprotein hierarchy" under selenium deficiency: proteins with higher Sec incorporation efficiency exploit more charged Sec-specific (Sec-tRNASec) 94 and are more rapidly synthesized [26]. The well-studied selenoproteins have antioxidant 95 properties (such as the glutathione peroxidase (GPx) family), are involved in redox 96 regulation (such as the thioredoxin reductases (TXNRD) family), or transport the serum 97 Se to tissues as selenoprotein P (SELENOP). But they also have other biological 98 functions, being regulators of growth, development, and cell differentiation, quality 99 control of protein biosynthesis, inhibitors of non-specific immune responses, 100 101 neutralizers of inflammatory responses, or antiapoptotic function [25, 27]. Many of these selenoproteins are expressed in the thyroid gland and are involved in different 102 processes, such as the formation and regulation of THs (the iodothyronine deiodinases 103 104 (DIO) family) and redox processes linked to gland protection (GPx and TXNRD) (GPx 105 and TXNRD) [24]. These selenoproteins are necessary for the correct functioning of the thyrotropin-releasing hormone (TRH) and thyroid stimulating hormone (TSH). TSH is 106 the major regulator of THs biosynthesis, since it activates a complex signaling network 107 108 across the TSH-receptor in thyrocytes and ends up forming T3 and T4 hormones (Figure 1) [28]. In addition, TSH is involved in the selenoproteins regulation since, 109 through its receptor, it clearly increases the expression of GPx1, GPx3 and 110 TXNRD1[29]. This signaling pathway also stimulate the expression of DIO1 (and DIO2 111 in human) inside the thyrocytes as well as H_2O_2 production [29.30.31]. 112

THs mediate important physiological processes such as development, growth, 113 thermogenesis, and energy metabolism, as well as regulate fatty acid, cholesterol, and 114 carbohydrate homeostasis [32,33]. The synthesis of THs is a complex, multistep process 115 116 that encompasses several redox reactions that need H_2O_2 as an oxidative agent. THs synthesis requires the oxidative iodination of specific tyrosine residues of thyroglobulin. 117 This process is catalyzed by the enzyme thyroid peroxidase (TPO), which requires an 118 appropriate amount of H_2O_2 for oxidation in the colloid (Figure 1). Therefore THs 119 synthesis needs H₂O₂ production. However, this is a disadvantage for thyrocyte, as this 120 large amount of H₂O₂ in the colloid could cross the apical membrane of the thyrocyte 121 and accumulate inside the cell, leading to OS-damage. 122

As it was mentioned, GPxs protect thyroid follicles from excess H_2O_2 that is produced during the synthesis of THs [34]. Cytotoxic ROS are mainly produced in thyroid follicles following activation of TPO as a result of the interaction between H_2O_2 , iodide and heme iron [35]. It has been demonstrated that the role of these selenoproteins, in relation to H_2O_2 , is fundamental to the thyroid, since in severe Se deficiency the lack of GPx activity causes oxidative damage to the thyroid gland, leading to thyroid damage and fibrosis [36, 37]. It has also been shown that pre-incubation of human thyroid follicles with Se (sodium selenite), even at low doses (10 nM) increases GPx activity and decreases cell death induced by high doses of H_2O_2 , iodide or TGF- β [38, 39].

The TXNRDs also play an important role in thyroid metabolism and, together with 132 133 thioredoxin (Trx) and NADPH, form the thioredoxin system, common to nearly all living cells [40]. This system functions in thiol-dependent thiol-disulfide exchange 134 reactions, crucial for controlling the reduced intracellular redox environment, cell 135 proliferation and growth, defence against oxidative stress or control of apoptosis. 136 Moreover, this system participates in the synthesis of deoxyribonucleotides for DNA 137 synthesis and is involved in cancer protection [40]. The two main thioredoxin 138 139 reductases are thioredoxin reductase 1 (TXNRD1), a cytosolic and nuclear form, and thioredoxin reductase 2 (TXNRD2), which is found only as a mitochondrial form [41]. 140 TXNRDs are highly expressed in thyroid cells [8]. 141

Specifically, type 1 and 2 deiodinases (DIO1 and DIO2) activate THs, while type 3 142 deiodinases (DIO3) inactivate both tetraiodothyronine (T4) and 3,5,3'-triiodothyronine 143 (T3) [42]. DIO1 is mainly found in the liver, kidneys and thyroid [43]. In humans, most 144 145 of the circulating T3 is derived from the conversion of T4 to T3 by the actions of DIO1 [44]. Unlike DIO1, the primary function of DIO2 is believed to be the supply of T3 to 146 the nucleus so as to meet intracellular needs, as it is a subcellularly located 147 148 selenoprotein that appears in muscle, brain, heart, bone and brown adipose tissue [45]. DIO2 is important in determining T3 content in developing tissues and the adult brain, 149 and in promoting the process of adaptive thermogenesis in brown adipose tissue. In 150 particular, DIO2 plays a primary role in T4-mediated negative feedback in the pituitary 151 gland and hypothalamus, in which T4 inhibits the expression of thyroid stimulating 152 hormone (TSH) and thyrotropin-releasing hormone (TRH), respectively [33]. DIO3 is 153 the physiological inactivator of THs, which acts by catalysing the deiodination of T4 154 into reverse triiodothyronine (rT3) and converts T3 into 3,3 diiodothyronine (T2) [46]. 155 This enzyme controls the local homeostasis of THs and protects tissues from their 156 157 excess [47]. Deiodinases appear to occupy a special place in the hierarchy in cases of selenium deficiencies thanks to the existence of a selenium accumulation and/or 158 159 redistribution system in the thyroid gland [39]. Initial cell culture and animal 160 experimental studies indicated that adequate nutritional selenium supply appears to limit 161 expression of functional deiodinases during development and in the adult organism. However, the deiodinative turnover of thyroid hormones requires only minimal amounts 162 163 of active enzymes, in contrast to enzymatic pathways acting on abundant metabolic 164 intermediates (e.g. carbohydrate, fatty acid, aminoacido or proteins). This might be one of the reasons why inadequate intake of the essential trace element selenium does not 165 initially manifest as impaired deiodinase activity, but rather affects those metabolic 166 167 pathways, which are catalyzed by more abundant selenoenzymes acting at higher substrate concentrations. These include GPxs and TXNRD involved in celular redox 168 control, several endoplasmatic reticulum-associated selenoproteins as well as 169 170 selenoprotein N (SELENON), all of which contribute to protein biosynthesis or represente structural componentes of cells and tissues [6]. Although small amounts of 171 Se are required for the activity of DIOs, a deficiency of this nutrient decreases THs 172 synthesis and has a major impact on thyroid function [48]. Decreased production of THs 173

leads to stimulation of the TRH-TSH-THs axis, due to lack of control of negativefeedback, increasing the production of TSH [36].

Finally, other selenoproteins, including selenoprotein P, K, S (SELENOP, SELENOK, and SELENOS, respectively) as well as SELENON are actively secreted in the thyrocytes. SELENOP is actively secreted together with GPx3 to protect thyrocytes from H_2O_2 at the colloid in absence of TSH, while the rest of the selenoproteins, within the endoplasmatic reticulum, take part in the quality control pathways [28, 49, 50]. The biosynthesis of these protective selenoproteins is mainly affected by genotype, Se availability, and inflammatory cytokines [28, 49].

183 **3. Selenium and thyroid cancer**

Se is recognised as a nutrient with many health benefits in humans and other mammals 184 such as decreasing the incidence of cancer [51]. Although the specific mechanisms are 185 not fully understood, the chemopreventive effects of Se result from its protective role on 186 cell membranes against OS, its stabilising effect on DNA and its enhancement of 187 cellular immune response [52]. This element also inhibits the proliferation of tumor 188 cells by acting on the expression of the Bcl-2 apoptosis-suppressor gene and p53 tumor 189 suppressor gene, which plays an important role in the processes of control and 190 regulation of cell lifecycle and DNA replication. Furthermore, in vitro and in vivo 191 192 studies have revealed that both Se compounds and selenoproteins act as anti-metastatic agents, inhibiting cell motility, migration and invasion, and reducing angiogenic factors 193 [53]. Nevertheless, it is important to mention that some selenoproteins, like TXNRD1, 194 SELENOF and GPx2 exhibit a split role in preventing and promoting cancer [51]. In 195 addition, Se may exhibits a U-shape relation with cancer risk [54, 55]. 196

197 Various studies have been carried out to examine the relationship between Se and the
198 development of thyroid cancer (Table 1). Overall, the findings suggest a potential
199 association between lower Se concentrations and the development of thyroid cancer.

200 Kucharzewski et al. [56] found that whole blood Se concentrations in a group of 201 patients (n=21) with thyroid cancer were significantly lower (0.57 μ g/g) than in a 202 control group (0.71 μ g/g, p < 0.01). There is no information as to the histological types of cancer included in the research. Moncayo et al. [57], in a study of patients with 203 benign and malignant thyroid pathologies taking thyroid medication found that serum 204 Se levels were lower in patients with papillary (n=73) (0.080 \pm 0.020 µg/ml) and 205 follicular (n=42) (0.077 \pm 0.021 µg/ml) carcinoma than in the control group (0.091 \pm 206 0.021 μ g/ml), p = 0.015 and p = 0.031 respectively). On the other hand, Przybylik-207 Mazurek et al. [58] found no significant changes in Se levels in the serum of patients 208 209 with papillary carcinoma (n=25) and in that of patients with follicular carcinoma (n=13)compared with a control group. The same finding was noted for glutathione peroxidase 210 3 (GPx3) activity. The tumours were diagnosed histologically on routine basis, after 211 surgery. The lag time between surgery and this study examination ranged between 8 and 212 120 months, with mean \pm SD of 42.9 \pm 25.3 months. Patients with carcinomas were 213 receiving thyroid medication. Subsequently, in 2013, Jonklaas et al. [59] conducted a 214 study involving a group of euthyroid patients with indication for thyroidectomy for 215 suspected thyroid cancer or nodular disease. Of the cohort, 48 patients had differentiated 216 thyroid carcinoma and 17 had benign thyroid pathology. 33 of patients with 217

Authors, year	Study design	Description of participants	Blood Se levels	Main results	Reference
------------------	-----------------	--------------------------------	--------------------	--------------	-----------

222

223

²¹⁸ differentiated thyroid carcinoma had papillary carcinoma. Blood samples were obtained 219 two to four weeks before thyroidectomy. In the final analysis, although Se 220 concentrations were not significantly lower in thyroid cancer patients, they were 221 inversely correlated with disease stage (p = 0.011).

Kucharzewski et al., 2003	- Cross - sectional study	 Thyroid cancer (n=21) Control (n= 50) 	- Whole blood (TXRF) - 0.57 ± 0.12 (μg/g) - 0.71 ± 0.06 (μg/g)	- Whole blood Se levels were significantly lower in the group of patients with thyroid cancer vs. control group (p < 0.01)	[56]
Moncayo et al., 2008	- Cross - sectional study	 Papillary carcinoma (n=73) Follicular carcinoma (n= 42) Control (n= 554) 	- Serum (AAS) - 0.080 ± 0.020 (μg/ml) - 0.077 ± 0.021 (μg/ml) - 0.091 ± 0.021 (μg/ml)	- Serum Se levels were significantly lower in patients with papillary and and folicular carcinoma vs. control group (p = 0.015 and p = 0.031 respectively)	[57]
Przybylik- Mazurek et al., 2011	- Cross - sectional study	 Papillary carcinoma (n=25) Follicular carcinoma (n= 13) Control (n=20) 	- Serum (AAS) - 0.78 ± 0.12 (μM/L) - 0.80 ± 0.14 (μM/L) - 0.76 ± 0.12 (μM/L)	 No significant differences among the groups in serum Se levels 	[58]
Jonklaas et al., 2013	- Cross - sectional study	- Differentiated thyroid carcinoma (n=48) - Benign thyroid disease (n= 17)	- Serum (AAS) - 0.116 ± 0.014 (μg/ml) - 0.117 ± 0.010 (μg/ml)	 No significant differences among the groups in serum Se levels Serum Se levels were inversely correlated with thyroid cancer stage (p=0.011) 	[59]
Baltaci et al., 2017	- Cross - sectional study	 Group 1: male thyroid cancer patients group (n = 15) Group 2: female thyroid cancer patients group (n = 15); Group 3: male control group (n = 10) Group 4: female control group (n = 10). 	- Serum (ICP-AES) Pre-operative (μ g/dl) - Group 1: 52.4 ± 5.6 - Group 2: 50.5 ± 4.8 - Group 3: 70.1 ± 6.9 - Group 4: 66.9 ± 7.3 Post- operative (μ g/dl) - Group 1: 54.6 ± 5.5 - Group 2: 51.7 ± 5.2 - Group 3: 69.5 ± 7.1 - Group 4: 67.6 ± 5.9 15 days after the operation (μ g/dl) - Group 1: 70.6 ± 5.9 - Group 2: 70.2 ± 5.5 - Group 3: 72.5 ± 6.5 - Group 4: 68.6 ± 8.0	 Pre- and postoperative serum Se concentrations in patients with thyroid cancer were significantly lower in serum vs. control groups (p < 0.05) 15 days after the operation, insignificant differences were detected in serum Se concentrations among the groups 	[60]
Mehl et al., 2020	- Cross - sectional study	- Thyroid patients (n=323) - Control (n=200)	- Serum (TXRF) - 76.9 ±18.8 (μg/L) - 85.1 ± 17.4 (μg/L)	 A high fraction of patients (37.5%) was classified as Se- deficient (serum Se concentrations <70 µg/L), in particular the patients with thyroid malignancy (59%) 	[61]

224

Table 1. Summary of the most important clinical trials examining the relation between blood Se levels and thyroid cancer. TXRF- total-reflection X-ray fluorescence, AASatomic absorption spectrometry, ICP-AES- inductively coupled plasma - atomic emission spectrometry.

229

In 2017, Baltaci et al. [60] conducted a study to examine the changes in serum Se levels
before, immediately after and fifteen days after thyroidectomy in patients (n=30) with

thyroid cancer (papillary carcinoma). In addition, thyroid tissue samples were taken 232 from all subjects in the postoperative period. Serum Se levels were significantly 233 decreased (p < 0.05) before and immediately after surgery compared with the controls. 234 Fifteen days later Se levels were similar to those found in the control group. Thyroid Se 235 levels postoperatively were significantly higher (p < 0.05) than those of the controls. 236 The fact that the same patients have less Se in their serum indicates, according to the 237 authors, that Se is retained excessively in the thyroid and that changes in the levels of 238 this mineral could be related to the pathogenesis of thyroid cancer. Very recently, Mehl 239 et al. [61] carried out a study to assess the levels of trace elements (iodine, Se, copper 240 and zinc) in patients with thyroid pathologies in a European metropolis. The authors 241 found that patient serum Se values were lower than those in control group participants 242 (p < 0.0001) More importantly, it was found that it was in the group of patients with 243 244 thyroid malignancy (n= 17) that a higher fraction of Se deficient patients were found (59%). 245

It is not yet clear whether the decrease in serum Se levels detected in most studies on thyroid cancer is a consequence or a cause of the disease or if it is simply associated with related pro-inflammatory conditions that alter the expression and secretion of hepatic selenoprotein P, the main contributor to the Se content in serum [6]. A decrease in this protein may be a phenomenon secondary to negative regulation triggered by inflammatory mediators such as tumour necrosis factor α (TNF- α), interleukin 1 β (IL-1 β) and interferon γ (IFN- γ) [62, 63].

253 **4. Selenoproteins and thyroid cancer**

Se is co-translationally inserted in protein as the 21st amino acid, Sec and accounts for a 254 vast majority of the biological activities of Se [64]. Twenty-five selenoproteins have 255 been identified in the human proteome and twenty-four in rat and mouse proteome [65]. 256 The share of selenium in the metabolic pathways associated with the protection of cells 257 258 against oxidative stress causes changes in the activity of selenoproteins. Selenoprotein expression is regulated by the concentration of this element [66, 67]. However, there are 259 260 differences in protein expression. These differences are the result of changes in mRNA translation or the reduction of its stability (increased degradation) [67]. 261

There have also been several studies relating the activity and expression of selenoproteins with thyroid cancer, the most studied being the DIO1 and DIO2 implicated in the control of THs turnover, and GPx1, GPx3 and TXNRD1, which protect thyroid from OS-damage [11, 21, 68, 69, 70, 71, 72, 73, 74].

266

4.a. Selenoproteins implicated in the control of THs turnover

Deiodinase expression patterns in thyroid cancers vary and depend on the type and differentiation of the tumour stage. T3 is known to regulate the expression and/or activity of tumour suppressors genes and oncogenes. Thus, local alterations in the expression and activity of DIOs may have the potential to influence carcinogenesis [75].

Different studies in papillary and follicular carcinomas support this fact. In 2005,
Arnaldi et al. [68] found that DIO1 and DIO2 were underexpressed in papillary
carcinoma following evaluation using cDNA analysis of three thyroid cancer cell lines.
In the same year, Ambroziak et al. [69] identified significantly decreased levels of DIO1

and DIO2 expression (p = 0.017 and p = 0.012, respectively) in papillary carcinoma 275 samples compared to control group samples (thyroid tissue from a non-cancer affected 276 part in human patients) and Meyer et al. [70] found in human patients that DIO1 277 278 expression and activity were decreased in papillary carcinoma samples compared to surrounding normal tissue $(0.25 \pm 0.24 \text{ vs. } 1.09 \pm 0.54 \text{ arbitrary units (AU)}, p < 0.001$ 279 280 and 0.08 ± 0.07 vs. 0.24 ± 0.15 pmol T4/min/mg protein, p = 0.045, respectively). However, in the latter study, the authors found a significant increase in DIO1 expression 281 and activity in tissue samples with follicular carcinoma (1.2 ± 0.46 vs. 0.67 ± 0.18 AU, 282 p = 0.038 and 1.20 ± 0.58 vs. 0.20 ± 0.10 pmol T4/min/mg protein, p < 0.001, 283 respectively). They also detected an increase in DIO2 activity in tissue samples with 284 metastatic follicular carcinoma (5.20 ± 0.81 vs. 0.30 ± 0.27 fmol T4/min/mg protein, p 285 < 0.001) Subsequently, Romitti et al. [76] analysed the expression and activity of DIO3 286 287 in papillary carcinoma human samples. The researchers observed that the augmentations in D3 activity were paralleled by increased DIO3 mRNA levels (approximately 288 fivefold). They also found a positive correlation between tumour size and DIO3 activity 289 290 (r=0.68, p=0.003). Finally, they found that an increase in DIO3 activity in tumour samples was associated with more advanced disease at diagnosis. 291

Taken together, one could posit that the changes found in the expressions of DIOs in 292 papillary carcinoma samples could cause a decrease in intracellular hormones and 293 294 favour tumour proliferation. Increased DIO3 and decreased DIO1 and DIO2, leading to decreased T3 concentrations, could provide an advantage for tumour cell proliferation, 295 as THs can block oncogenic Ras-mediated proliferation, which specifically interferes 296 297 with the activity of the mitogen-activated protein kinase (MAPK) signalling pathway 298 [71]. This pathway has previously been implicated in DIO3 overregulation in other 299 pathological changes [77,78]. Genetic alterations leading to the activation of this 300 pathway are a distinguishing marker of papillary thyroid carcinoma [76]. It is known that DIO3 is upregulated in the papillary thyroid carcinoma-derived cell line, K1, by 301 transforming growth factor β 1 (TGF β 1). Furthermore, it is known that treatment with 302 303 the inhibitors U0126 (ERK pathway) and SB203580 (p38 pathway) leads to blocking of 304 the MAPK pathway and subsequent decrease of DIO3 and inhibition of transcriptional 305 induction of DIO3 through TGF β 1, which clearly suggests that DIO3 is positively 306 regulated through the MAPK signalling pathway [76,79]. In the development of this 307 carcinoma, the BRAF gene is one of those principally affected, with the BRAFV 600 E mutation occurring frequently, through substitution of a valine for a glutamic acid at 308 position 600 [80]. In the study described above, Romitti et al. [76] found that the 309 310 samples in which this mutation was present were those in which there was greater DIO3 activity. Subsequently, Romitti et al. [81] found that activation of the sonic hedgehog 311 (SHH) pathway could also be involved in DIO3 upregulation through a signalling 312 313 cooperation with the MAPK pathway. SHH signalling is critical for embryogenesis and other cellular processes such as proliferation and differentiation. Disruption of SHH 314 signalling leads to several human diseases and appears to contribute to the development 315 of neoplastic processes. Reactivation of SHH occurs in about 25% of human tumours 316 and has been associated with the induction of DIO3 [81-83]. 317

Interestingly, retinoic acid (RA) has been shown to induce DIO1 activity in human thyroid carcinoma cell lines. RA transcriptionally increased the abundance of the p27 subunit of DIO1. RA stimulated DIO1 activity to a greater extent in follicular thyroid carcinoma-133 cells than in follicular thyroid carcinoma-238 cells and had no effect in
 anaplastic thyroid carcinoma. Retinoid induction of DIO1 may thus serve as a parameter
 of functional differentiation of thyroid follicular carcinoma cells [84].

The higher DIO2 activity, in metastatic follicular carcinoma, without significant changes in DIO2 mRNA levels, suggests that DIO2 uprregulation occurs mainly via post-transcriptional regulatory mechanisms [71]. The expression pattern of DIO2 reveals that this selenoprotein is encoded by an cAMP-sensitive gene, so its expression increases in tumoural contexts, such as follicular carcinoma, in which there is an overstimulation of the cAMP pathway [45].

330

4.b. Selenoproteins which protect thyroid from OS-damage

OS plays an important role in carcinogenesis by inducing DNA damage and its effects on intracellular signal transduction pathways. ROS can induce almost all forms of DNA damage that have been described in the dysfunction of genes involved in cancer genesis and play a key role in cancer development by originating and maintaining oncogenic phenotypes [65].

In the thyroid gland, high amounts of H₂O₂ are produced, which triggers high OS, 336 during synthesis of thyroid hormones in follicular cells. On the other hand, as 337 mentioned above, Se deficiency, regardless of the cause, diminished expression and 338 activity of selenoproteins with antioxidant functions such as GPx and TXNRDs. Thus, 339 these selenoproteins cannot fight properly ROS generated during cellular metabolism, 340 increasing OS and cancer genesis [85]. Moreover, thyroid cancer itself can induce OS 341 through inflammation, which is one of its significant features, and this in turn is a 342 classic source of ROS. As a consequence of OS, instability in DNA can be produced 343 and maintained, which are believed to be neoplasia-preceding events in thyroid cells 344 345 [86].

There are also two 'professional' ROS-generating systems in thyroid gland, the NADPH oxidases DUOX1 and NOX4, which cause DNA damage that may promote chromosomal instability, tumourigenesis and anaplasia. Ionising radiation and mutation of oncogenes such as RAS and BRAF positively regulate these NADPH oxidases, playing a key role in thyroid carcinogenesis [87]. In turn, ROS can stimulate MAPK, phosphatidylinositol-3-kinase (PI3K) and NFκB pathways, forming a vicious circle that spurs carcinogenesis [86].

In this context, Young et al. [88] evaluated the levels of DNA damage and lipid pe-353 354 roxidation, measured 8-oxo-7,8-dihydro-2'-deoxyguanosine (8-oxo-dG) and the 4-HNE 355 respectively, in patients with follicular thyroid adenoma (n = 71), papillary thyroid carcinoma (n = 45) and follicular thyroid carcinoma (n = 17). They established that the 356 cytoplasmic expression of 8-oxo-dG and 4-HNE was significantly higher in thyroid 357 tissue samples from patients with follicular adenoma, follicular carcinoma and papillary 358 carcinoma compared to their normal tissue (all p values < 0.001). Similarly, increased 359 nuclear levels of 8-oxo-dG were found in thyroid tissue samples from patients with 360 follicular adenoma, follicular carcinoma and papillary carcinoma compared to their 361 normal tissue (p values < 0.07, p < 0.001 and p < 0.001, respectively). 362

363 Since a correct oxidative balance plays an important role in thyroid carcinogenesis, the 364 main antioxidant selenoproteins expressed in thyroid have been analysed in this context.

GPx1 is distributed throughout the human body and its main activity is antioxidant [1]. It catalyses the reduction of H₂O₂, using reduced glutathione (GSH), transforming it into water. During this process, glutathione is oxidised, subsequently returning to its original state through the action of the enzyme glutathione reductase (GR), so as to maintain GSH levels [89]. GPx1 is one of the selenoproteins that is most sensitive to Se alterations in the body, exhibiting dramatic reductions when this microelement is depleted [90].

- Zagrodzki et al. [21] detected decreased GPx1 activity in anaplastic carcinoma samples
 compared to normal tissue samples from surrounding parts of the gland. Previously,
 Hasegawa et al. [72] found decreased expression of this enzyme in the same type of thyroid cancer. More recently, Metere et al. [11] analysed the expression of GPx1 and
 TXNRD1 in tissue samples with papillary carcinoma. The researchers found a reduction
 in the expression of these enzymes compared to that observed in healthy tissue samples
 taken from the same patients.
- Several studies show a decrease in GPx1 expression in different tumours and suggest a protective role for GPx1 [91]. GPx1 can limit oxidant-induced cell mutagenesis, as well as the inflammatory responses that promote certain cancers. Loss of GPx1 in the early stages of carcinogenesis may contribute to cancer initiation and, in later stages, its deficiency may induce proliferative responses [92].
- With respect to GPx3, this selenoenzyme is the only extracellular enzyme of the GPx family [65]. It has an important extracellular antioxidant role and affords protection to the thyroid against H_2O_2 [36]. It is one of the most highly expressed selenoproteins in follicular cells and, as a result, contributes to the high Se levels of the thyroid [39, 93].
- In situ hybridization revealed to Menth et al. [73] reduced levels of GPx3 in Hurthle cell 388 carcinoma. Subsequently, Schmutzler et al. [94] also used in situ hybridization to study 389 390 the localization of various selenoproteins within the thyroid gland using goiter, 391 autoimmune thyroiditis or thyroid tumor samples. The researchers found that the strongest hybridization signals were obtained from GPx3 mRNA. In thyroid 392 carcinomas, the follicular structure of the normal thyroid was disrupted and GPx3 393 394 signals were evenly dispersed over the whole sample areas or parts of it without any of the thyroid-specific differential distribution patterns observed, e.g., in goiter. More 395 recently, Zhao et al. [74] investigated the expression of GPx3 in patients with primary 396 papillary carcinoma. They found that its expression was often reduced/absent in the 397 398 carcinoma samples compared to surrounding healthy tissue samples. In addition, they found that GPx3 was frequently methylated in the carcinoma samples. These authors 399 also found that the reduced/absent expression was related to hypermethylation of the 400 promoter region and that carcinoma metastasis was suppressed by GPx3 through 401 inhibition of the Wnt / β-catenin signalling pathway. In this context, gene hyper-402 methylation has also been indicated as a cause of down-regulation in various tumour 403 404 tissues [95-97]. Decreased GPx3 expression has been associated with tumour initiation, proliferation and migration as a consequence of increased oxidative stress and pro-405

406 tumourigenic redox signalling. It is currently unclear whether loss of GPx3 leads to407 compensatory increases in other antioxidant enzymes in tumour cells [98].

408 GPx7 is mainly involved in maintaining the redox homeostasis of the body [99]. In functional terms, GPx7 is not selenocysteine-containing peroxidase due to the lack of a 409 GSH-binding domain, but rather a protein disulfide isomerase peroxidase. It is located 410 in the lumen of the endoplasmic reticulum, where it uses the H_2O_2 produced by 411 endoplasmic reticulum oxidoreductase 1 alpha to oxidize protein disulfide isomerase 412 [100]. Recently, Li-Dan Liu et al. [99] investigated the expression of GPx7 in papillary 413 thyroid carcinoma tissues. In this study, GPx7 was found to be expressed at higher 414 levels in papillary thyroid carcinoma tissues and papillary thyroid carcinoma cell lines 415 416 than in other thyroid tissues and related to the size of papillary thyroid tumors. GPx7 was successfully knocked down in K1 cells, and knockdown of GPx7 inhibited cell 417 proliferation and clone formation as well as increased apoptosis and caspase 3/7 activity 418 419 in K1 cells. According to the authors, these results demonstrated that GPx7 promotes the growth of papillary thyroid carcinoma but the mechanisms underlying of action of 420 GPx7 on proliferation and apoptosis are still unclear. 421

422 Overexpression and hyperactivation of TXNRD1 have been described in several cancers

[101]. Moreover, the high expression and activity of TXNRD1 has been directly related
to cellular protection against oxidative stress induced by 4-hydroxynonenal (4- HNE),
one of the end products of lipid peroxidation [102].

426 Metere et al. [11] suggest that the reduction in TXNRD1 and GPx1 expression seen in 427 their study may result from hyperproduction of free radicals, which were not adequately 428 counteracted by the altered antioxidant system in cancer cells, possibly due to increased 429 consumption of antioxidants. Furthermore, the authors detected a significant increase in 430 free radical production in all thyroid tumour tissue samples, compared with healthy 431 tissue from the same patients.

432 Finally, it is important to bear in mind that not just selenoproteins protect thyroid from OS-damage. For instance, J. Maier et al. [103] detected a higher mRNA expression for 433 superoxide dismutase (SOD)-3 isoform and increased total SOD enzyme activity in the 434 thyroid exposed to iodine deficiency compared to normal diet. Especially increased 435 SOD-3 expression, which is the extra cellular SOD isoform, could detoxify superoxide 436 in the follicular lumen and might act as an effective shield against oxidative stress 437 induced by ROS in response to luminal H_2O_2 . Catalase, as well as the peroxired oxins 438 (PRDX), also protect thyroid cells against H₂O₂ [104, 105]. 439

440 5. Selenium supplementation and thyroid cancer

Due to growing evidence suggesting the vulnerability of cancer cells to oxidative stress, 441 the idea of targeting the antioxidant capacity of tumour cells has grown as a therapeutic 442 strategy, leading to the rational design of new anticancer agents. Accordingly, Se has 443 stood out as a redox modulator of cancer cells among compounds with great anti-cancer 444 potential [106]. Different forms of Se have anti-cancer effects on different cancers, such 445 as hepatocarcinoma, breast cancer, oesophageal cancer, prostate cancer and ovarian 446 cancer, among others [25]. However, several studies have shown that Se has a tumor-447 promoting effect. The NPC trial, for example, found that selenium supplementation (as 448

selenized yeast; 200 µg/day) significantly increased the risk of non-melanoma skin 449 cancer and squamous-cell carcinoma [107]. Another study was conducted on the 450 population of the Reggio Emilia municipality in Italy, who were exposed to $7-9 \mu g/liter$ 451 of selenate in tap water from 1975 to 1985. Melanoma incidence was 3.9 times higher in 452 selenium-exposed people than in non-selenium exposed people, according to the 453 454 findings of this study [108]. More recently, Tsuji et al. [109] detected that the deficiency in the 15 kDa selenoprotein inhibits human colon cancer cell growth. Because Sep15 455 expression depends on the selenium status, these results are important in regards to 456 differential intake of, and response to, dietary selenium and potential cancer risk. 457

458 The anti-cancer activity of different forms of Se depends on many factors, such as 459 chemical form, dose, acute vs. long-term nutrition, preventive or pharmacological application, type of cancer cell, bioavailability, and stage of disease [110]. Selenium 460 exhibits chemopreventive activity when used at higher than optimal concentrations or 461 applied for cancer treatment in combination with chemotherapy and radiation [111]. 462 Also, the greatest anticarcinogenic Se effect has been obtained when it is administered 463 before or at an early stage of disease development. It is important to bear in mind that 464 one of the marginal problems of Se use is its narrow range between the toxic dose and 465 the dose necessary for the proper functioning of living organisms [112]. In recent years, 466 due to both their reduced toxicity and their selectivity, Se nanoparticles are considered 467 to be more effective in cancer treatment than other Se compounds. These nanoparticles 468 are prepared by chemical, physical or biological methods, and contain a main inorganic 469 therapeutic core of elemental Se, which presents better antitumor properties than the Se 470 salts [113]. Based on this, Kuršvietienė et al. [110], studying the role of Se and the 471 472 selenoproteins in maintaining cellular redox balance and anticancerogenic function, 473 suggested that nanoparticles are taken up by cancer cells via endocytosis. In these cells, 474 Se nanoparticles act as prooxidants producing endoplasmic reticulum stress, 475 mitochondrial membrane cleavage, apoptosis, DNA fragmentation, and cell cycle arrest. 476 Besides, Khurana et al. [114] trying to understand the various pharmacological activities 477 of Se nanoparticles as well reduction in toxicity of Se upon nanoparticlization, proposed 478 that Se nanoparticles act as antioxidants against high ROS generated by cancer cells. Moreover, their low toxicity, their high bioavailability and biocompatibility are some of 479 480 the properties that also make Se nanoparticles an attractive drug delivery vehicle, 481 reducing the systemic toxicities associated with conventional chemotherapeutic drugs and working synergistically to improve their efficacy [115]. Very recently, beneficial in 482 vivo results have been obtained using a tumor model of mice preinoculated with K1 483 484 cells and treated with a combination of drugs including Se nanoparticles for the 485 treatment of thyroid cancer [116]. However, additional and independent studies are needed to confirm these results in animals. On the other hand, in order to apply the 486 anticancer benefits of Se nanoparticle in clinical studies, extensive studies on its safety 487 488 and synergistic activities with other therapeutic compounds are still needed.

Even though Se supplementation may combat the development of thyroid cancer, the
data that exist so far are not conclusive. The question of whether a Se deficit is a
consequence of thyroid cancer or a predisposing risk factor remains unresolved [20].
Access to mostly retrospective data, relatively small patient groups and short
observation periods are significant limitations of the studies performed [117].

Recent studies have shown beneficial effects of Se supplementation. Nettore et al. [7] 494 conducted a study to characterise the molecular effects determined by Se 495 supplementation (10 nM sodium selenite) on thyroid follicular cells, using the rat 496 497 thyroid follicular cell line FRTL5 as a model. They examined the effect of Se on cell growth, mortality and proliferation, and modulation of pro- and anti-apoptotic 498 pathways, concluding that Se supplementation improved the growth rate of FRTL5. 499 Furthermore, they found that Se reduced cell death and was associated with a 500 501 downregulation of the proapoptotic genes p53 and Bim and an upregulation of the antiapoptotic genes NF-kB and Bcl2. More recently, Ruggeri et al. [118] investigated 502 the effects of Se on oxidative damage in human thyroid follicular cells and thyroid 503 fibroblasts in vitro. Primary cultures were exposed to H_2O_2 in the presence or absence 504 505 of Se, in the form of selenomethionine or selenite. Administration of increasing 506 concentrations of Se, from 0.05 to 20 µM, significantly prevented the genotoxic and cytotoxic effects of H₂O₂ by increasing cell viability, reducing morphological 507 abnormalities, improving cellular DNA integrity and decreasing lipid peroxidation. 508 509 H₂O₂-induced apoptosis was reduced and almost eliminated, as evidenced by reduced 510 caspase-3 activity and modulation of the expression of the antiapoptotic Bcl2/proapoptotic Bax genes. Furthermore, both selenomethione and selenite induce an 511 512 increase in GPx activity which, according to the authors, suggests that these protective 513 effects may be, in part, mediated by these selenoproteins.

514 Animal and human studies have suggested that supplementation with different forms of Se at concentrations higher than those required to maximise selenoprotein expression 515 decreases the incidence of cancer [119]. However, a recent meta-analysis that took as 516 517 one of its research questions "describe the efficacy of Se supplementation for cancer 518 prevention in humans" found no evidence to suggest that increasing Se intake, through 519 diet or by supplementation, prevents cancer in humans [120]. All observational studies 520 and randomised trials appear to be highly conditioned by population characteristics with regard to covariates and confounding factors, which include initial Se intake levels, 521 522 antioxidants cosupplemented, age, gender, diet, lifestyle [65].

523 **6.** Conclusions and future perspectives

524 Se and selenoproteins play a significant role in the development of thyroid cancer. It is 525 generally agreed that oxidative stress plays an important role in cancer genesis and tumour progression. Most studies indicate an association between Se deficiency and the 526 development of thyroid cancer, as well as significant changes in the expression and 527 528 activity of various selenoproteins in different types of thyroid cancer. The mechanisms 529 underlying these changes are not yet fully understood. Although Se supplementation is theoretically beneficial for cancer, in practice the studies are not conclusive, mainly due 530 531 to methodological limitations. In this respect, and specifically for thyroid cancer, the 532 literature is also very scarce. Nevertheless, Se components have already become part of 533 the therapeutic strategy to fight thyroid cancer. It is expected that in the near future there will be a greater knowledge of these components' mechanisms of action, in order to 534 535 improve their use in the prevention and treatment of thyroid cancer.

536

- 537 Author Contributions: Writing—original draft preparation, R.M.R.; writing—review,
- 538 R.M.R.; F.N.; M.L.O.; O.C.; editing, F.N.; M.L.O.; supervision, R.M.R.; F.N.; M.L.O.;
- 539 O.C.; management of reference, R.M.R.; funding acquisition, O.C. All authors have
- read and agreed to the published version of the manuscript.
- Funding: Andalusian Regional Government for its support to CTS-193 research group
 (2017/CTS-193 and 2019/CTS-193).
- 543 **Declarations of interest:** none.
- 544
- 545 **References**
- 546
- 547 [1] Y. Mehdi, J.L. Hornick, L. Istasse, I. Dufrasne, Selenium in the environment,
 548 metabolism and involvement in body functions, Molecules 18 (2013) 3292-3311.
 549 https://doi.org/10.3390/molecules18033292.
- [2] C. Reilly, Selenium in Food and Health, 2nd ed.; Springer Science & BusinessMedia: New York, 2006.
- [3] P.A. Tsuji, C.D. Davis, J.A. Milner, Selenium: Dietary Sources and Human
 Requirements, in: D.L.Hatfield, M.J. Berry, V.N. Gladyshev (Eds.), Selenium its
 Molecular Biology and Role in Human Health, 3rd edn. Springer, New York, 2012.
- [4] R.M. Rua, M.L.Ojeda, F.Nogales, J.M. Rubio, M. Romero-Gómez, J. Funuyet,
 M.L. Murillo, O. Carreras, Serum selenium levels and oxidative balance as differential
 markers in hepatic damage caused by alcohol, Life Sci. 94 (2014) 158-163.
 https://doi.org/10.1016/j.lfs.2013.10.008.
- [5] R.C. Dickson, R.H. Tomlinson (1967). Selenium in blood and human tissues. Clin.
 Chim. Acta. 16 (1967) 311–321. https://doi.org/10.1016/0009-8981(67)90197-0.
- [6] J. Köhrle, Selenium and the thyroid, Curr. Opin. Endocrinol. Diabetes Obes.22
 (2015) 392-401. https://doi.org/10.1097/med.00000000000190.
- [7] I.C. Nettore, E.De Nisco, S. Desiderio, C. Passaro, L. Maione, M. Negri, L. Albano,
 R. Pivonello, C. Pivonello, G. Portella, P.Ungaro, A. Colao, P.E. Macchia, Selenium
 supplementation modulates apoptotic processes in thyroid follicular cells, Biofactors. 43
 (2017) 415-423. https://doi.org/10.1002/biof.1351.
- [8] J. Köhrle, R. Gärtner, Selenium and thyroid, Best Pract. Res. Clin. Endocrinol.
 Metab. 23 (2009) 815-827. https://doi.org/10.1016/j.beem.2009.08.002.
- 569 [9] B.A. Zachara, J. Gromadzińska, W. Wasowicz, Z. Zbróg, Red blood cell and plasma
- 570 glutathione peroxidase activities and selenium concentration in patients with chronic
- kidney disease: a review, Acta. Biochim. Pol. 53 (2006) 663-677.
- 572 https://doi.org/10.18388/abp.2006_3294.
- 573 [10] C. Guerra-Araiza, A.L. Alvarez-Mejía, S. Sánchez-Torres, E. Farfan-García, R.
 574 Mondragón-Lozano, R. Pinto-Almazan, H. Salgado-Ceballos, Effect of natural

- 575 exogenous antioxidants on aging and on neurodegenerative diseases, Free Radic. Res. 47 (2013) 451-462. https://doi.org/10.3109/10715762.2013.795649. 576
- 577 [11] A. Metere, F. Frezzotti, C.E. Graves, M. Vergine, A. De Luca, D. Pietraforte, L. 578 Giacomelli, A possible role for selenoprotein glutathione peroxidase (GPx1) and thioredoxin reductases (TrxR1) in thyroid cancer: our experience in thyroid surgery, 579 Cancer Cell Int. 18 (2018) 7. https://doi.org/10.1186/s12935-018-0504-4. 580
- 581 [12] A. Prete, P. Borges de Souza, S. Censi, M. Muzza, N. Nucci, M. Sponziello, Update on fundamental mechanisms of thyroid cancer, Front. Endocrinol. (Lausanne) 582 583 11 (2020) 102. https://doi.org/10.3389/fendo.2020.00102.
- [13] M.E. Cabanillas, D.G. McFadden, C. Durante, Thyroid cancer, Lancet.388 (2016) 584 2783-2795. https://doi.org/10.1016/S0140-6736(16)30172-6. 585
- 586 [14] L.Lamartina, G.Grani, C.Durante, I. Borget, S.Filetti, M. Schlumberger, Follow-
- 587 up of differentiated thyroid cancer - what should (and what should not) be done, Nat. Rev. Endocrinol.14 (2018) 538-551. https://doi.org/10.1038/s41574-018-0068-3. 588
- [15] T. Carling, R. Udelsman, Thyroid cancer, Annu. Rev. Med. 65 (2014) 125-137. 589 https://doi.org/10.1146/annurev-med-061512-105739. 590
- 591 [16] A.E. Lorea, I.M. Bermejo, E.A. Apiñániz, J.P.Arribas, M.T. García, J.P. Martínez
- 592 de Esteban, A.M. Insausti Serrano, Comparison of clinical characteristics of patients 593 with follicular thyroid carcinoma and Hürthle cell carcinoma, Endocrinol. Diabetes 594 Nutr. (English Ed). 65 (2018) 136-142. https://doi.org/10.1016/j.endinu.2017.12.006.
- 595 [17] S. Kure, R. Ohashi, Thyroid Hürthle Cell Carcinoma: Clinical, Pathological, and 596 Molecular Features, Cancers 13 (2020) 26. https://doi.org/10.3390/cancers13010026.
- 597 [18] H. He, T. Xu, P. Li, G. Jia, X. Li, Q. Song, Anti-PD-1 Immunotherapy Combined
- With Stereotactic Body Radiation Therapy and GM-CSF as Salvage Therapy in a PD-598
- 599 L1-Positive Patient With Refractory Metastatic Thyroid Hürthle Cell Carcinoma: A
- Case Report and Literature Review, Front. Oncol. 11 (2021). 600
- 601 https://doi.org/10.3389/fonc.2021.782646.
- 602 [19] T.J. O'Grady, C.M. Kitahara, A.G. DiRienzo, M.A.Gates, The association between
- selenium and other micronutrients and thyroid cancer incidence in the NIH-AARP Diet 603 604 and Health Study, PLoS One. 9 (2014) e110886.
- https://doi.org/10.1371/journal.pone.0110886. 605
- 606 [20] A. Valea, C.E. Georgescu, Selenoproteins in human body: focus on thyroid
- 607 pathophysiology, Hormones (Athens) 17 (2018) 183-196.
- 608 https://doi.org/10.1007/s42000-018-0033-5.
- 609 [21] P. Zagrodzki, F. Nicol, J.R. Arthur, M. Słowiaczek, S. Walas, H. Mrowiec, R.
- Wietecha-Posłuszny, Selenoenzymes, laboratory parameters, and trace elements in 610
- different types of thyroid tumor, Biol. Trace Elem. Res. 134 (2010) 25-40. 611
- 612 https://doi.org/10.1007/s12011-009-8454-2.
- [22] A.P. Kipp, Selenium in colorectal and differentiated thyroid cancer, Hormones 613
- (Athens, Greece), 19 (2020) 41–46. https://doi.org/10.1007/s42000-019-00118-4. 614

[23] M.B. Zimmermann, J. Köhrle, The impact of iron and selenium deficiencies on
iodine and thyroid metabolism: biochemistry and relevance to public health, Thyroid
(2002) 867-878. https://doi.org/10.1089/105072502761016494.

618 [24] O. Carreras, M.L. Ojeda, F. Nogales, Selenium Dietary Supplementation and
619 Oxidative Balance in Alcoholism, in: V.B. Patel (Ed.), Molecular Aspects of Alcohol
620 and Nutrition, Elsevier, New York, 2016, pp. 133 –142. https://doi.org/10.1016/B978621 0-12-800773-0.00011-2.

- [25] S. Hariharan, S. Dharmaraj, Selenium and selenoproteins: it's role in regulation of
- 623 inflammation, Inflammopharmacology 28 (2020) 667-695.
- 624 https://doi.org/10.1007/s10787-020-00690-x.
- [26] Y. F. Chen, H.C. Lin, K. N. Chuang, C. H. Lin, H. S. Yen, C.H. Yeang, A
- quantitative model for the rate-limiting process of UGA alternative assignments to stopand selenocysteine codons, PLoS Comput. Biol. 13 (2017) e1005367.
- 628 https://doi.org/10.1371/journal.pcbi.1005367.
- 629 [27] M.L.Ojeda, O.Carreras, P.Sobrino, M.L.Murillo, F.Nogales, Biological
- 630 implications of selenium in adolescent rats exposed to binge drinking: Oxidative,
- 631 immunologic and apoptotic balance, Toxicol. Appl. Pharmacol. 329 (2017) 165-172.
- 632 https://doi.org/10.1016/j.taap.2017.05.037.
- [28] L. Schomburg, Selenium, selenoproteins and the thyroid gland: interactions in
- health and disease. Nat. Rev. Endocrinol. 8 (2011) 160–171.
- 635 https://doi.org/10.1038/nrendo.2011.174.
- 636 [29] G.J. Beckett, J.R. Arthur, Selenium and endocrine systems. J. Endocrinol. 184
 637 (2005) 455–465. https://doi.org/10.1677/joe.1.05971.
- [30] J.Köhrle, Thyrotropin (TSH) action on thyroid hormone deiodination and
 secretion: one aspect of thyrotropin regulation of thyroid cell biology, Horm. Metab.
 Res. Suppl. 23 (1990) 18–28.
- [31] S.G.Beech, S.W. Walker, J.R. Arthur, D. Lee, G.J. Beckett, Differential control of
 type-I iodothyronine deiodinase expression by the activation of the cyclic AMP and
- 643 phosphoinositol signalling pathways in cultured human thyrocytes. J. Mol. Endocrinol.
- 644 14 (1995) 171–177. https://doi.org/10.1677/jme.0.0140171.
- [32] R.Mullur, Y.Y.Liu, G.A. Brent, Thyroid hormone regulation of metabolism.
 Physiol. Rev. 94 (2014) 355–382. https://doi.org/10.1152/physrev.00030.2013.
- [33] R. Arrojo E Drigo, T.L. Fonseca, J.P.S.Werneck-de-Castro, A.C. Bianco, Role of
- 648 the type 2 iodothyronine deiodinase (D2) in the control of thyroid hormone signaling,
- 649 Biochim. Biophys. Acta 1830 (2013) 3956-3964.
- 650 https://doi.org/10.1016/j.bbagen.2012.08.019.
- [34] Y. Liu, H. Huang, J. Zeng, C. Sun, Thyroid volume, goiter prevalence, and
- selenium levels in an iodine-sufficient area: a cross-sectional study, BMC Public
- 653 Health. 13 (2013) 1153. https://doi.org/10.1186/1471-2458-13-1153.

- [35] N. Maouche, D. Meskine, B. Alamir, E.A. Koceir, Trace elements profile is
- associated with insulin resistance syndrome and oxidative damage in thyroid disorders:
- 656 Manganese and selenium interest in Algerian participants with dysthyroidism, J. Trace
- 657 Elem. Med. Biol. 32 (2015) 112-121. https://doi.org/10.1016/j.jtemb.2015.07.002.
- [36] M. Ventura, M.Melo, F.Carrilho, Selenium and Thyroid Disease: From
- 659 Pathophysiology to Treatment, Int. J. Endocrinol. 4 (2017)1-9.
- 660 https://doi.org/10.1155/2017/1297658.
- 661 [37] F. Pakdel, R. Ghazavi, R. Heidary, A. Nezamabadi, M. Parvizi, M.H.S.A. Memar,
- 662 R. Gharebaghi, F. Heidary, Effect of Selenium on Thyroid Disorders: Scientometric
- 663Analysis, Iran J. Public Health. 48 (2019) 410-420.
- 664 http://dx.doi.org/10.18502/ijph.v48i3.883.

[38] P. Lehmann, P. Rank, K.L.J. Hallfeldt, B. Krebs, R. Gärtner, Dose-related
influence of sodium selenite on apoptosis in human thyroid follicles in vitro induced by
iodine, EGF, TGF-beta, and H2O2, Biol. Trace Elem. Res. 112 (2006) 119-130.
https://doi.org/10.1385/BTER:112:2:119.

- [39] A. Drutel, F. Archambeaud, P. Caron, Selenium and the thyroid gland: more good
- news for clinicians, Clin. Endocrinol. (Oxf). 78 (2013) 155-164.
- 671 https://doi.org/10.1111/cen.12066.
- [40] E.S.J. Arnér, A. Holmgren, The thioredoxin system in cancer, Semin. Cancer
 Biol. 16 (2006) 420-426. https://doi.org/10.1016/j.semcancer.2006.10.009.
- [41] D.T. Lincoln, F. Al-Yatama, F.M. Mohammed, A.G. Al-Banaw, M. Al-Bader, M.
 Burge, F. Sinowatz, P.K. Singal, Thioredoxin and thioredoxin reductase expression in
 thyroid cancer depends on tumour aggressiveness, Anticancer Res.30 (2010) 767-775.
- [42] A.C. Bianco, R.R. da Conceição, The Deiodinase Trio and Thyroid Hormone
- 678 Signaling, Methods. Mol. Biol. 1801 (2018) 67-83. https://doi.org/10.1007/978-1-4939-679 7902-8_8.
- [43] A.C. Bianco, A. Dumitrescu, B. Gereben, M.O. Ribeiro, T.L. Fonseca, G.W.
 Fernandes, B.M.L.C, Bocco, Paradigms of Dynamic Control of Thyroid Hormone
 Signaling. Endocr. Rev. 40 (2019) 1000-1047. https://doi.org/10.1210/er.2018-00275.
- [44] G.R.Williams, J.H.D. Bassett, Deiodinases: the balance of thyroid hormone: local
 control of thyroid hormone action: role of type 2 deiodinase, J. Endocrinol. 209 (2011)
 261-272. https://doi.org/10.1530/JOE-10-0448.
- [45] A. Nappi, M.A. De Stefano, M. Dentice, D. Salvatore, Deiodinases and Cancer,
 Endocrinology. 162 (2021) https://doi.org/10.1210/endocr/bqab016.
- [46] M.M. AlRasheed, A. AlAnzi, R. AlShalhoub, N. Abanmy, D. Bakheet, A study of
- the role of DIO1 and DIO2 polymorphism in thyroid cancer and drug response to
- therapy in the Saudi population, Saudi Pharm. J. 27 (2019) 841-845.
- 691 https://doi.org/10.1016/j.jsps.2019.05.005.

[47] D. Ciavardelli, M. Bellomo, C. Crescimanno, V. Vella, Type 3 deiodinase: role in
cancer growth, stemness, and metabolism, Front. Endocrinol. (Lausanne). 5 (2014)
215. https://doi.org/10.3389/fendo.2014.00215.

[48] J. Knezevic, C. Starchl, A. Tmava Berisha, K. Amrein, Thyroid-Gut-Axis: How
Does the Microbiota Influence Thyroid Function? Nutrients 12 (2020) 1769.
https://doi.org/10.3390/nu12061769.

[49] K.H.Winther, M.P. Rayman, S.J. Bonnema, L. Hegedüs, Selenium in thyroid
disorders - essential knowledge for clinicians, Nat. Rev. Endocrinol. 16 (2020), 165–
176. https://doi.org/10.1038/s41574-019-0311-6.

- 701 [50] M. Minnetti, V. Sada, T. Feola, E. Giannetta, C. Pozza, D. Gianfrilli, A.M. Isidori,
- A.Cozzolino, Selenium Supplementation in Pregnant Women with Autoimmune
- 703 Thyroiditis: A Practical Approach. Nutrients, 14 (2022) 2234.
- 704 https://doi.org/10.3390/nu14112234.
- [51] D.L. Hatfield, P.A.Tsuji, B.A.Carlson, V. N. Gladyshev, Selenium and
- selenocysteine: roles in cancer, health, and development. Trends Biochem. Sci. 39
- 707 (2014) 112–120. https://doi.org/10.1016/j.tibs.2013.12.007.
- 708 [52] M. Kieliszek, S. Błażejak, Current Knowledge on the Importance of Selenium in
- Food for Living Organisms: A Review, Molecules 21 (2016) 609.
- 710 https://doi.org/10.3390/molecules21050609.
- [53] Y.C. Chen, K.S. Prabhu, A.M. Mastro, Is selenium a potential treatment for cancer
 metastasis? Nutrients 5 (2013) 1149-1168. https://doi.org/10.3390/nu5041149.
- 713 [54] H. Fritz, D.Kennedy, D.Fergusson, R. Fernandes, K.Cooley, A. Seely, S. Sagar,
- R.Wong, D. Seely, Selenium and lung cancer: a systematic review and meta analysis.
- 715 PloS One, 6 (2011) e26259. https://doi.org/10.1371/journal.pone.0026259.
- [55] R. Hurst, L. Hooper, T. Norat, R. Lau, D. Aune, D. C. Greenwood, R.Vieira, R.
 Collings, L.J. Harvey, J.A. Sterne, R. Beynon, J. Savović, S.J. Fairweather-Tait,
 Selenium and prostate cancer: systematic review and meta-analysis. Am. J. Clin. Nutr.
 96 (2012) 111–122. https://doi.org/10.3945/ajcn.111.033373.
- [56] M. Kucharzewski, J. Braziewicz, U. Majewska, S. Gózdz, Copper, zinc, and
 selenium in whole blood and thyroid tissue of people with various thyroid diseases.
 Biol. Trace Elem. Res. 93 (2003) 9-18. https://doi.org/10.1385/BTER:93:1-3:9.
- [57] R. Moncayo, A. Kroiss, M. Oberwinkler, F. Karakolcu, M. Starzinger, K.
- 724 Kapelari, H. Talasz, H. Moncayo, The role of selenium, vitamin C, and zinc in benign
- thyroid diseases and of selenium in malignant thyroid diseases: Low selenium levels are
- found in subacute and silent thyroiditis and in papillary and follicular carcinoma, BMC
- 727 Endocr. Disord. 8 (2008) 2. https://doi.org/10.1186/1472-6823-8-2.
- 728 [58] E. Przybylik-Mazurek, P. Zagrodzki, S. Kuźniarz-Rymarz, A. Hubalewska-
- 729 Dydejczyk, Thyroid disorders-assessments of trace elements, clinical, and laboratory
- 730 parameters, Biol. Trace Elem. Res. 141 (2011) 65-75. https://doi.org/10.1007/s12011-
- 731 010-8719-9.

- [59] J. Jonklaas, M. Danielsen, H. Wang, A pilot study of serum selenium, vitamin D,
 and thyrotropin concentrations in patients with thyroid cancer, Thyroid 23 (2013) 10791086. https://doi.org/10.1089/thy.2012.0548.
- [60] A.K. Baltaci, T.K. Dundar, F. Aksoy, R. Mogulkoc, Changes in the Serum Levels
 of Trace Elements Before and After the Operation in Thyroid Cancer Patients, Biol.
- 737 Trace Elem. Res. 175 (2017) 57-64. https://doi.org/10.1007/s12011-016-0768-2.
- [61] S. Mehl, Q. Sun, C.L. Görlich, J. Hackler, J.F. Kopp, K. Renko, J. Mittag, T.
- 739 Schwerdtle, L. Schomburg, Cross-sectional analysis of trace element status in thyroid
- 740 disease, J. Trace Elem. Med. Biol. 58 (2020) 126430.
- 741 https://doi.org/10.1016/j.jtemb.2019.126430.
- [62] M. W. Angstwurm, R. Gaertner, Practicalities of selenium supplementation in
- rtically ill patients, Curr. Opin. Clin. Nutr. Metab. Care 9 (2006) 233-238.
- 744 https://doi.org/10.1097/01.mco.0000222105.30795.7f.
- [63] M. Geoghegan, D. McAuley, S. Eaton, J. Powell-Tuck, Selenium in critical illness,
- 746 Curr. Opin. Crit. Care 12 (2006) 136-141.
- 747 https://doi.org/10.1097/01.ccx.0000216581.80051.d6.
- [64] J. Fernandes, X. Hu, M. Ryan Smith, Y. M. Go, D.P. Jones, Selenium at the redox
 interface of the genome, metabolome and exposome, Free Radic. Biol. Med.127 (2018)
 215–227. https://doi.org/10.1016/j.freeradbiomed.2018.06.002.
- [65] M. Roman, P. Jitaru, C. Barbante, Selenium biochemistry and its role for human
 health, Metallomics. 6 (2014) 25-54. https://doi.org/10.1039/c3mt00185g.
- [66] M.J. Lammi, C. Qu, Selenium-Related Transcriptional Regulation of Gene
 Expression, Int. J. Mol. Sci. 19 (2018) 2665. https://doi.org/10.3390/ijms19092665.
- [67] M. Kieliszek, Selenium- Fascinating Microelement, Properties and Sources in
 Food. Molecules, 24 (2019) 1298. https://doi.org/10.3390/molecules24071298.
- [68] L.A. Arnaldi, R.C. Borra, R.M. Maciel, J.M. Cerutti, Gene expression profiles
 reveal that DCN, DIO1, and DIO2 are under-expressed in benign and malignant thyroid
 tumors, Thyroid 15 (2005) 210-221. https://doi.org/10.1089/thy.2005.15.210.
- [69] M. Ambroziak, J. Pachucki, E. Stachlewska-Nasfeter, J. Nauman, A. Nauman,
 Disturbed expression of type 1 and type 2 iodothyronine deiodinase as well as
 titf1/nkx2-1 and pax-8 transcription factor genes in papillary thyroid cancer, Thyroid 15
 (2005) 1137-1146. https://doi.org/10.1089/thy.2005.15.1137.
- [70] E.L. De Souza Meyer, J.M. Dora, M.S. Wagner, A.L. Maia, Decreased type 1
 iodothyronine deiodinase expression might be an early and discrete event in thyroid cell
 dedifferentiation towards papillary carcinoma, Clin. Endocrinol. 62 (2005) 672–678.
 https://doi.org/10.1111/j.1365-2265.2005.02277.x.
- [71] I.M. Goemann, V.R. Marczyk, M. Romitti, S.M.Wajner, A.L. Maia, Current concepts and challenges to unravel the role of iodothyronine deiodinases in human neoplasias, Endocr. Relat. Cancer 25 (2018) R625-R645. https://doi.org/10.1530/erc-18-0097.

- [72] Y.Hasegawa, T. Takano, A. Miyauchi, F. Matsuzuka, H. Yoshida, K. Kuma, N.
- Amino, Decreased expression of glutathione peroxidase mRNA in thyroid anaplastic
 carcinoma, Cancer Lett. 182 (2002) 69-74. https://doi.org/10.1016/s0304-
- 775 3835(02)00069-1.
- [73] M. Menth, C. Schmutzler, B. Mentrup, C. Hoang-Vu, K. Takahashi, T. Honjoh, J.
- Köhrle, Selenoprotein expression in Hürthle cell carcinomas and in the human Hürthle
- cell carcinoma line XTC.UC1, Thyroid 15 (2005) 405-416.
- 779 https://doi.org/10.1089/thy.2005.15.405.
- [74] H. Zhao, J. Li, X. Li, C. Han, Y. Zhang, L. Zheng, M. Guo, Silencing GPX3
 Expression Promotes Tumor Metastasis in Human Thyroid Cancer, Curr. Protein. Pept.
 Sci. 16 (2015) 316-321. https://doi.org/10.2174/138920371604150429154840.
- [75] A. Piekiełko-Witkowska, A. Nauman, Iodothyronine deiodinases and cancer, J.
 Endocrinol. Invest. 34 (2011) 716-728. https://doi.org/10.3275/7754.
- [76] M. Romitti, S.M. Wajner, N. Zennig, I.M. Goemann, A.L. Bueno, E.L. Meyer,
 A.L. Maia, Increased type 3 deiodinase ex-pression in papillary thyroid carcinoma,
 Thyroid 22 (2012) 897-904. https://doi.org/10.1089/thy.2012.0031.
- [77] H. Zrouri, C. Le Goascogne, W.W. Li, M. Pierre, F. Courtin, The role of MAP kinases in rapid gene induction after lesioning of the rat sciatic nerve, Eur. J. Neurosci. 20 (2004) 1811-1818. https://doi.org/10.1111/j.1460-9568.2004.03641.x.
- [78] A.Lamirand, S. Pallud-Mothré, M. Ramaugé, M. Pierre, F. Courtin, Oxidative
 stress regulates type 3 deiodinase and type 2 deiodinase in cultured rat astrocytes,
 Endocrinology 149 (2008) 3713-3721. https://doi.org/10.1210/en.2007-1462.
- [79] Y.C. Liu, C.T. Yeh, K.H, Lin, Molecular Functions of Thyroid Hormone Signaling
 in Regulation of Cancer Progression and Anti-Apoptosis, Int. J. Mol. Sci. 20, (2019),
 4986. https://doi.org/10.3390/ijms20204986.
- [80] P. Mondragón-Terán, L.B. López-Hernández, J. Gutiérrez-Salinas, J.A. SuárezCuenca, R.I.Luna-Ceballos, A. Erazo Valle-Solís, Mecanismos de señalización
 intracelular en cáncer de tiroides [Intracellular signaling mechanisms in thyroid cancer],
 Cir. Cir. 84 (2016) 434-443. https://doi.org/10.1016/j.circir.2016.05.017.
- [81] M.Romitti, S.M. Wajner, L. Ceolin, C.V. Ferreira, R.V.P. Ribeiro, H.C.
 Rohenkohl, S.S. Weber, P.L.C. Lopez, C.S. Fuziwara, E.T. Kimura, A.L. Maia,
 MAPK and SHH pathways modulate type 3 deiodinase expression in papillary thyroid
 carcinoma, Endocr. Relat. Cancer 23 (2016) 135-146. https://doi.org/10.1530/erc-150162.
- 806 [82] M. Dentice, C. Luongo, S. Huang, R. Ambrosio, A. Elefante, D. Mirebeau-Prunier,
- 807 A.M. Zavacki, G. Fenzi, M. Grachtchouk, M. Hutchin, A.A. Dlugosz, A.C. Bianco, C.
- 808 Missero, P.R. Larsen, D. Salvatore, Sonic hedgehog-induced type 3 deiodinase blocks
- thyroid hormone action enhancing proliferation of normal and malignant keratinocytes,
- 810 Proc. Natl. Acad. Sci. 104 (2007) 14466–14471.
- 811 https://doi.org/10.1073/pnas.0706754104.

- 812 [83] D.K.L. Aw, R.A. Sinha, H.C. Tan, L.M. Loh, D. Salvatore, P.M. Yen, Studies of
- molecular mechanisms associated with increased deiodinase 3 expression in a case of
 consumptive hypothyroidism, J. Clin. Endocrinol. Metab. 99 (2014) 3965–3971.
 https://doi.org/10.1210/jc.2013-3408.
- [84] H.Y.Lin, Y.T. Chin, Y.C. Yang, H.Y. Lai, J. Wang-Peng, L.F. Liu, H.Y. Tang,
 P.J. Davis, Thyroid Hormone, Cancer, and Apoptosis, Compr. Physiol. 6 (2016) 12211237. https://doi.org/10.1002/cphy.c150035.
- [85] E. Zoidis, I. Seremelis, N. Kontopoulos, G.P. Danezis, Selenium-Dependent
 Antioxidant Enzymes: Actions and Properties of Selenoproteins, Antioxidants (Basel) 7
 (2018) 66. https://doi.org/10.3390/antiox7050066.
- [86] M. Xing, Oxidative stress: a new risk factor for thyroid cancer, Endocr. Relat.
 Cancer 19 (2012) C7-C11. https://doi.org/10.1530/erc-11-0360.
- [87] R.A. El Hassani, C. Buffet, S. Leboulleux, C. Dupuy, Oxidative stress in thyroid
 carcinomas: biological and clinical significance, Endocr. Relat. Cancer 26 (2019)
 R131-R143. https://doi.org/10.1530/erc-18-0476.
- [88] O.Young, T. Crotty, R. O'Connell, J. O'Sullivan, A.J. Curran, Levels of oxidative
 damage and lipid peroxidation in thyroid neoplasia, Head Neck 32 (2010) 750-756.
 https://doi.org/10.1002/hed.21247.
- [89] L.V. Papp, J. Lu, A. Holmgren, K.K. Khanna, From selenium to selenoproteins:
 synthesis, identity, and their role in human health. Antioxid. Redox. Signal 9 (2007)
 775-806. https://doi.org/10.1089/ars.2007.1528.
- [90] R.A. Sunde, A.M. Raines, K.M. Barnes, J.K. Evenson, Selenium status highly
 regulates selenoprotein mRNA levels for only a subset of the selenoproteins in the
 selenoproteome, Biosci. Rep. 29 (2009), 329-338. https://doi.org/10.1042/bsr20080146.
- [91] S.P. Short, C.S. Williams, Selenoproteins in Tumorigenesis and Cancer
- 837 Progression, Adv. Cancer Res. 136 (2017) 49-83.
- 838 https://doi.org/10.1016/bs.acr.2017.08.002.
- [92] E. Lubos, J. Loscalzo, D.E. Handy, Glutathione peroxidase-1 in health and disease:
 from molecular mechanisms to therapeutic opportunities, Antioxid. Redox Signal. 15
 (2011) 1957-1997. https://dx.doi.org/10.1089%2Fars.2010.3586.
- [93] L. Schomburg, J. Köhrle, On the importance of selenium and iodine metabolism
 for thyroid hormone biosynthesis and human health, Mol. Nutr. Food Res. 52 (2008)
 1235-1246. https://doi.org/10.1002/mnfr.200700465.
- 845 [94] C.Schmutzler, B. Mentrup, L.Schomburg, C. Hoang-Vu, V. Herzog, J. Köhrle,
- 846 Selenoproteins of the thyroid gland: expression, localization and possible function of
- 847 glutathione peroxidase 3. Biol. Chem. 388 (2007) 1053–1059.
- 848 https://doi.org/10.1515/BC.2007.122.
- 849 [95] M.M. Mohamed, S. Sabet, D.F. Peng, M.A. Nouh, M. El-Shinawi, W. El-Rifai,
- 850 Promoter hypermethylation and suppression of glutathione peroxidase 3 are associated

- with inflammatory breast carcinogenesis, Oxid. Med. Cell Longev. 2014 (2014)
 787195. https://doi.org/10.1155/2014/787195.
- 853 [96] D.M. Yao, J.D. Zhou, Y.Y. Zhang, L.Yang, X.M.Wen, J. Yang, H. Guo, Q. Chen,
- J. Lin, J. Qian, GPX3 promoter is methylated in chronic myeloid leukemia, Int. J. Clin. Exp. Pathol. 8 (2015) 6450-6457.
- [97] H. Chen, Z. Zheng, K.Y. Kim, X. Jin, M.R. Roh, Z. Jin, Hypermethylation and
 downregulation of glutathione peroxidase 3 are related to pathogenesis of melanoma,
 Oncol. Rep. 36 (2016) 2737-2744. https://doi.org/10.3892/or.2016.5071.
- [98] C. Chang, B.L Worley, R. Phaëton, N. Hempel, Extracellular Glutathione
- Peroxidase GPx3 and Its Role in Cancer, Cancers (Basel) 12 (2020) 2197.
- 861 https://doi.org/10.3390/cancers12082197.
- 862 [99] L.D. Liu, Y.N. Zhang, L.F. Wang, GPX7 promotes the growth of human papillary
- thyroid carcinoma via enhancement of cell proliferation and inhibition of cell apoptosis.
- 864 Transl. Cancer Res. 8 (2019) 2570–2580. https://doi.org/10.21037/tcr.2019.10.14.
- [100] R.Brigelius-Flohé, L.Flohé, Regulatory Phenomena in the Glutathione Peroxidase
 Superfamily, Antioxid. Redox Signal. 33 (2020) 498–516.
- 867 https://doi.org/10.1089/ars.2019.7905.
- [101] M. Jaganjac, L. Milkovic, S.B. Sunjic, N. Zarkovic, The NRF2, Thioredoxin,
 and Glutathione System in Tumorigenesis and Anticancer Therapies, Antioxidants
 (Basel) 9 (2020) 1151. https://doi.org/10.3390/antiox9111151.
- [102] S. G. Leoni, E.T. Kimura, P.Santisteban, A. De la Vieja, Regulation of thyroid
 oxidative state by thioredoxin reductase has a crucial role in thyroid responses to iodide
 excess, Mol. Endocrinol. 25 (2011) 1924-1935. https://doi.org/10.1210/me.2011-0038.
- [103] J. Maier, H. van Steeg, C. van Oostrom, R. Paschke, R.E. Weiss, K. Krohn,
- 875 Iodine deficiency activates antioxidant genes and causes DNA damage in the thyroid
- gland of rats and mice. Biochim. Biophys. Acta. 1773 (2007) 990–999.
- 877 https://doi.org/10.1016/j.bbamcr.2007.03.011.
- 878 [104] H. Kim, T.H. Lee, E.S. Park, J.M. Suh, S.J. Park, H.K. Chung, O.Y. Kwon, Y.K.
- Kim, H. K. Ro, M. Shong, Role of peroxiredoxins in regulating intracellular hydrogen
 peroxide and hydrogen peroxide-induced apoptosis in thyroid cells, J. Biol. Chem. 275
- 881 (2000) 18266–18270. https://doi.org/10.1074/jbc.275.24.18266.
- [105] A. Nicolussi, S. D'Inzeo, C. Capalbo, G. Giannini, A. Coppa, The role of
- peroxiredoxins in cancer, Mol. Clin. Oncol. 6 (2017) 139–153.
- 884 https://doi.org/10.3892/mco.2017.1129.
- [106] A.P Fernandes, V.Gandin, Selenium compounds as therapeutic agents in cancer,
- 886 Biochim. Biophys. Acta 1850 (2015) 1642-1660.
- 887 https://doi.org/10.1016/j.bbagen.2014.10.008.
- 888 [107] A.J. Duffield-Lillico, E.H. Slate, M.E. Reid, B.W. Turnbull, P.A. Wilkins, G.F.
- 889 Combs Jr, H.K. Park, E.G. Gross, G.F. Graham, M.S. Stratton, J.R. Marshall, L.C,
- 890 Clark, Nutritional Prevention of Cancer Study Group, Selenium supplementation and

- secondary prevention of nonmelanoma skin cancer in a randomized trial. J. Natl.Cancer
 Inst. 95 (2003) 1477–1481. https://doi.org/10.1093/jnci/djg061.
- [108] M.Vinceti, K.J. Rothman, M.Bergomi, N. Borciani, L. Serra, G. Vivoli, Excess
 melanoma incidence in a cohort exposed to high levels of environmental selenium.
 Cancer Epidemiol. Biomarkers Prev. 7 (1998) 853–856.
- [109] P.A. Tsuji, S. Naranjo-Suarez, B.A. Carlson, R. Tobe, M.H. Yoo, C.D.Davis,
 Deficiency in the 15 kDa selenoprotein inhibits human colon cancer cell growth,
- 898 Nutrients. 3 (2011) 805–817. https://doi.org/10.3390/nu3090805.
- 899 [110] L. Kuršvietienė, A. Mongirdienė, J. Bernatonienė, J. Šulinskienė, I.
- 900 Stanevičienė, Selenium Anticancer Properties and Impact on Cellular Redox Status,
- 901 Antioxidants (Basel) 2020, 9, 80. https://doi.org/10.3390/antiox9010080.
- 902 [111] S.O. Evans, P.F. Khairuddin, M.B. Jameson, Optimising Selenium for Modulation
- 903 of Cancer Treatments, Anticancer. Res. 37 (2017) 6497-6509.
- 904 https://doi.org/10.21873/anticanres.12106.
- 905 [112] M. Kieliszek, B. Lipinski, S. Blazejak, Application of Sodium Selenite in the
- 906 Prevention and Treatment of Cancers, Cells 6 (2017) 39.
- 907 https://doi.org/10.3390/cells6040039.
- 908 [113] C. Ferro, H.F.Florindo, H.A.Santos, Selenium Nanoparticles for Biomedical
- Applications: From Development and Characterization to Therapeutics, Adv.Healthc.
 Mater 10 (2021) e2100508 https://doi.org/10.1002/edbm.202100508
- 910 Mater. 10 (2021) e2100598. https://doi.org/10.1002/adhm.202100598.
- 911 [114] A. Khurana, S. Tekula, M.A. Saifi, P. Venkatesh, C. Godugu, Therapeutic
- applications of selenium nanoparticles, Biomed. Pharmacother. 111 (2019) 802-812.
 https://doi.org/10.1016/j.biopha.2018.12.146.
- 914 [115] F. Maiyo, M. Singh, Selenium nanoparticles: potential in cancer gene and drug
 915 delivery, Nanomedicine (Lond) 12 (2017) 1075-1089. https://doi.org/10.2217/nnm-
- 916 2017-0024.
- 917 [116] X. Zou, Z. Jiang, L. Li, Z. Huang, Selenium nanoparticles coated with pH
- 918 responsive silk fibroin complex for fingolimod release and enhanced targeting in
- 919 thyroid cancer, Artif. Cells Nanomed. Biotechnol. 49 (2021) 83-95.
- 920 https://doi.org/10.1080/21691401.2021.1871620.
- 921 [117] M. Stuss, M. Michalska-Kasiczak, E. Sewerynek, The role of selenium in thyroid
- gland pathophysiology, Endokrynol. Pol. 68 (2017) 440-465.
- 923 https://doi.org/10.5603/ep.2017.0051.
- 924 [118] R.M. Ruggeri, A. D'Ascola, T.M. Vicchio, S. Campo, F. Gianì, S. Giovinazzo, F.
- 925 Frasca, S. Cannavò, A. Campennì, F. Trimarchi, Selenium exerts protective effects
- 926 against oxidative stress and cell damage in human thyrocytes and fibroblasts,
- 927 Endocrine. 68 (2020) 151-162. https://doi.org/10.1007/s12020-019-02171-w.
- 928 [119] K.S. Prabhu, X.G. Lei, Selenium, Adv. Nutr. 7 (2016) 415-417.
- 929 https://doi.org/10.3945/an.115.010785.

- 930 [120] M. Vinceti, T. Filippini, C. Del Giovane, G. Dennert, M. Zwahlen, M. Brinkman,
- 931 M.P. Zeegers, M. Horneber, R. D'Amico, C.M. Crespi, Selenium for preventing cancer,
- 932 Cochrane Database Syst. Rev. 1 (2018) CD005195.
- 933 https://doi.org/10.1002/14651858.cd005195.pub4.