1 Binge drinking during the adolescence period causes oxidative damage-induced cardiometabolic disorders: a

2 possible ameliorative approach with selenium supplementation.

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- 4 María Luisa Ojeda<sup>1</sup>, Fátima Nogales<sup>1,\*</sup>, María del Carmen Gallego-López<sup>1</sup>, and Olimpia Carreras<sup>1</sup>
- <sup>1</sup>Department of Physiology, Faculty of Pharmacy, Seville University, 41012 Seville, Spain.
- 6 \*Correspondence: <u>fnogales@us.es</u>
- 7
- 8 \*Address: Dra. Fátima Nogales Bueno.
- 9 Department of Physiology.
- 10 Faculty of Pharmacy, Seville University.
- 11 C/ Profesor García González, nº 2.
- 12 41012. Seville. Spain.
- 13 Tel: +34 954556518
- 14 Fax: +34 954233765
- 15 E-mail: <u>fnogales@us.es</u>
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## 30 Abstract:

31 Binge drinking (BD) is the most common alcohol consumption model among adolescents. BD exposure during 32 adolescence disrupts the nervous system function, being involved in the major mortality causes at this age: motor vehicle 33 accidents, homicides and suicides. Recent studies have also shown that BD consumption during adolescence affects liver, 34 renal and cardiovascular physiology, predisposing adolescents to future adult cardiometabolic damage. BD is a 35 particularly pro-oxidant alcohol consumption pattern, because it leads to the production of a great source of reactive 36 oxygen species (ROS) via the microsomal ethanol oxidizing system, also decreasing the antioxidant activity of 37 glutathione peroxidase (GPx). Selenium (Se) is a mineral which plays a pivotal role against oxidation; it forms part of the 38 catalytic center of different antioxidant selenoproteins such as GPxs (GPx1, GPx4, GPx3) and selenoprotein P (SelP). 39 Specifically, GPx4 has an essential role in mitochondria, preventing their oxidation, apoptosis and NFkB-inflamative 40 response, being this function even more relevant in heart's tissue. Se serum levels are decreased in acute and chronic 41 alcoholic adult patients, being correlated to the severity of oxidation, liver damage and metabolic profile. Experimental 42 studies have described that Se supplementation to alcohol exposed mice clearly decreases oxidative and liver damage. 43 However, clinical BD effects on Se homeostasis and selenoproteins' tissue distribution related to oxidation during 44 adolescence are not yet studied. In this narrative review we will describe the use of sodium selenite supplementation as an 45 antioxidant therapy in adolescent BD rats in order to analyze Se homeostasis implication during BD exposure, oxidative 46 balance, apoptosis and inflammation, mainly in liver, kidney, and heart. These biomolecular changes and the 47 cardiovascular function will be analyzed. Se supplementation therapies could be a good strategy to prevent the oxidation, 48 inflammation and apoptosis generated in tissues by BD during adolescence, such as liver, kidney and heart, improving 49 cardiovascular functioning.

50 Keywords: selenium, antioxidant, binge drinking, apoptosis, NFkB, alcohol metabolism.

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### 58 **1. Introduction:**

Ethanol (EtOH) is a major contributor to global disease, being a causal factor in more than 200 diseases and a leading cause of preventable death [1]. According to the World Health Organization (WHO), in 2018 EtOH consumption led to the 5.3 % of all deaths, increasing this value to 13.5 % in the 20–39 years age group. Moreover, EtOH consumption causes death and disability relatively early in life and during adolescence; during both periods, EtOH consumption could lead to disease later in life, including metabolic and cardiovascular diseases [2–4].

64 Relative to the adolescent period, Fagundes et al. reported that those individuals over 15 years of age consume on average 65 6.2 liters of pure alcohol per year [5]. Besides, their pattern of EtOH consumption is particularly dangerous, since adolescents consume high doses of EtOH for a short period of time. According to the National Institute on Alcohol Abuse 66 67 and Alcoholism (NIAAA), binge drinking (BD) is an acute EtOH consumption model, which brings blood alcohol concentration (BAC) to 0.08 % or higher within 2 hours. Before and during the COVID-19 pandemic, this pattern of 68 69 EtOH consumption has been the most widespread among teenagers [6,7], being a public health concern. Adolescence is a 70 stage especially vulnerable to the toxic neural effects of EtOH; to engage in risk-taking behavior, such as initial drug 71 consumption; and to increase a person's propensity for later EtOH problems [8,9]. Finally, hormonal changes during 72 puberty may also affect sensitivity to alcohol, making adolescents less sensitive to the effects of intoxication and 73 increasing its consumption [10]. The BD alcohol consumption model in adolescents has been associated with numerous 74 injuries and is involved in all of the major causes of mortality at this age: motor vehicle accidents, homicides, and suicides 75 [11]. Moreover, recently the adolescent BD exposure has not only been associated with nervous harms [12], but also with 76 long term systemic harms related to hepatic [13], metabolic [14], renal [15] and cardiovascular damage [16].

77 Acute EtOH consumption leads to different biological effects than chronic (Chr-EtOH), since in its metabolism, it 78 produces higher amounts of reactive oxygen species (ROS) by increasing hepatic CYP2E1 activity [17]; and also because 79 EtOH arrives in a higher amount to the extrahepatic tissues. Therefore, BD exposure is an especially potent pro-oxidant 80 that could damage different tissues, compromising future adolescent health later in life. Clinically, it is well documented 81 that Chr-EtOH consumption during adulthood decreases the antioxidant activity of glutathione peroxidase (GPx), a 82 selenoprotein which needs selenium (Se) in its catalytic center; presenting these patients lower serum Se levels [18,19]. 83 Moreover, serum Se levels showed a direct correlation with GPx activity and lipid oxidation, suggesting that serum 84 Se/lipid oxidation ratio could be an indicator of hepatic damage caused by Chr-EtOH consumption; this points to Se as a

possible antioxidant therapy to alcoholic patients with liver disease [20]. Recently, Isobe et al. have found in adult patients that the alcohol intake is associated with serum Se levels and Selenoprotein P [21], a hepatokine related to insulin resistance, lipid profile and cardiovascular function. Clinical acute-EtOH studies in adults exposed to chronic heavy alcohol consumption also found lower serum and erythrocyte Se levels in these patients, which were independent of malnutrition, and returned to normal levels by cessation of EtOH ingestion; these data indicate a clear inverse correlation among acute EtOH intake and serum Se levels [22–25].

91 However, there are no clinical studies related to BD EtOH consumption and serum Se levels during adolescence. These 92 data should be interesting since this depletion could be compromising the oxidative balance in the adolescence even more. 93 Moreover, since plasma levels of metabolites are not a simple reflection of changes in tissue levels of the same 94 metabolites [26], Se tissue deposits should be analyzed. In this context, preclinical animal research strategies are needed 95 to enhance our understanding of the effects of BD consumption during adolescence on oxidative balance. Se and 96 selenoproteins' tissue balance; and its relationship to metabolic and cardiovascular physiology. Considering the fact that 97 there are few studies in adult acute-EtOH exposed mice showing that hepatic Se and SelP deposits are related to EtOH 98 consumption [27], and that Se supplementation by modulating these deposits is effective avoiding oxidative, apoptotic, 99 inflammatory and metabolic hepatic damage [27,28], only our research group has studied these effects in BD exposed 100 adolescent rats [14,15,29-31]. This narrative review will try to summarize these findings.

## 101 **2. BD and Oxidative Stress**

Among the pathophysiological mechanisms that have been identified as causative factors in tissue and organ injuries as a consequence of excessive EtOH consumption, such as BD, are: acetaldehyde generation, adduct formation, mitochondrial injury, cell membrane perturbations, immune modulation, and oxidative stress (OS). Some of these mechanisms can appear as a result of direct alcohol-induced cell perturbations, while the others are the consequence of tissue alcohol metabolism [11]. Moreover, the consumption pattern, amount, frequency, and type of alcoholic beverages all contribute to provoking alcohol-induced tissue injury. For this reason, it is important to analyze EtOH metabolism and OS generation during BD exposition.

EtOH intake damages cells and organs in part due to the OS generated while this drug is being metabolized by the oxidative pathway. OS is an imbalance between the production of free radicals and the body's ability to detoxify or fight against their harmful effects by neutralizing them with antioxidants [32]. Studies with animals and humans indicate that BD can specifically increase, even more than other EtOH consumption patterns, the development of reactive oxygen species (ROS) and decrease the antioxidant body activity [30,33–38]. The main ROS molecules generated by the organisms and by the BD-exposition include the superoxide anion ( $O_2^{-}$ ), hydroxyl radical (•OH) and hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>), these reactive species could lead to lipid, protein and DNA damage, which would compromise cell functioning [11,39]. Therefore, OS causes serious damage in different tissues, including liver and heart, since disturbances in the redox-equilibrium also trigger intracellular inflammatory and apoptotic signaling, which may further progress to different health issues related to cardiovascular function [40–42].

119 The oxidative metabolism of alcohol uses the enzymes alcohol dehydrogenase (ADH), cytochrome P450 2E1 (CYP2E1), 120 and catalase (CAT), all of which generate acetaldehyde [43]. In mitochondria, the enzyme alcetaldehyde dehydrogenase 121 (ALDH) subsequently coverts acetaldehyde into acetate which is then released from the liver into blood and thence exported to other tissues where it enters the Krebs cycle (KC) to be broken down into CO<sub>2</sub> and H<sub>2</sub>O [44]. During this 122 123 metabolism, both ADH and ALDH enzymes reduce NAD<sup>+</sup> to NADH, shifting the cellular redox ratio [45]. High NADH 124 levels lead to an increase in ROS formation and a decrease in eNOS activity [38,46]. The liver is the main organ for 125 metabolizing ingested alcohol – more than 90%; however, when BACs are high, EtOH can arrive to more tissues, where it 126 is also metabolized leading to OS.

ADH is located in the cellular cytosol and plays a significant role in alcohol metabolism, however due to its low Km, it is saturated once the BACs exceed 15-20 mg/dL (2 × Km), as happens during BD [45,47]. Therefore during BD, alcohol is then oxidized by the CYP2E1 located in the endoplasmic reticulum, which has a significantly higher Km. CYP2E1 requires reduced nicotinamide adenine dinucleotide phosphate (NADPH) or NADPH-regenerating systems to act [48,49]. This results in the production of a high amount of ROS [11,50]. Induced CYP2E1 is described as a major contributor of the OS that causes a predisposition to experimental and clinical liver injury [51].

There is another pathway by which EtOH-exposure leads to OS. It has a direct action on the biological membranes of cells and organelles. In high amount, such as BD exposure, it damages the mitochondrial membrane and provokes a dysfunction of these organelles, causing ROS overproduction and reducing ATP production, displaying apoptosis [35,52,53].

In addition, alcohol consumption also affects the enzymatic and non-enzymatic antioxidant systems. BD-exposure
decreases reduced glutathione (GSH), which plays an important pathogenic role in alcoholic liver disease [46,48,54].
Moreover, BD exposure disturbs the hepatic activity of the endogenous antioxidant system: Superoxide Dismutase
(SOD), CAT, Glutathione Peroxidase (GPx), and Glutathione Reductase (GR) [15,29,33,55,56]. The SOD enzyme acts as

the first line of defense against free radicals, since it catalyzes the dismutation of  $O_2$ - to  $H_2O_2$  and oxygen. Then, the  $H_2O_2$ generated or lipid hydroperoxides are reduced by CAT and GPx enzymes to water and oxygen, and to water and alcohol respectively. GPx uses the GSH to reduce its substrate, working together with GR, an enzyme NADPH-dependent that regenerates GSH from its oxidized form, the glutathione disulfide (GSSG). This antioxidant system is unbalanced in adolescent rats exposed to BD, being the selenoprotein GPx the most affected enzyme [15,29,33,55].

BD exposure during adolescence generates not only hepatic oxidative damage, but also systemic and cardiac OS [55,57]. 146 147 ROS generation alters the redox-homeostasis leading to systemic damage and increasing vascular endothelial growth 148 factor (VEGF), which provokes microvascular proliferation-related cellular damage [42]. Besides, increased lipid 149 peroxidation and nitric oxide promotes reduced cell viability, deoxyribonucleic acid damage and apoptosis via activation 150 of caspase 3 in liver and heart [40,42]. In this context, the use of exogenous antioxidants in pro-oxidative situations 151 significantly ameliorates the induced hyperlipidemia, the metabolic disturbances, the inflammatory response (via 152 regulating the NFkB expression), and it avoids the loss of mitochondrial membrane potential decreasing the apoptosis 153 [41].

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#### 3. Selenium and Selenoproteins

Selenium (Se<sup>34</sup><sub>79</sub>) is a metalloid member of the group XVI of the periodic table, closely allied in chemical and physical properties with the elements sulfur and tellurium. It was discovered in 1817 by the Swedish chemist Jöns Jacob Berzelius.
Se is a micronutrient, which plays an essential role in human health since, as part of the catalytic center of different selenoproteins, it is involved in a variety of physiological processes; the best known is its antioxidant capacity.

159 The Recommended Dietary Allowance (RDA) for Se intake is  $20-30 \ \mu g/d$  for children, from 40 to 70  $\mu g/d$  for adult males 160 and, 45–60 µg/d for adult females, rising to 65 µg/day for pregnant women and 75 µg/day for lactating mothers [58,59]. No data is available regarding Se requirements for adolescents; this reference values are based on the values compiled for 161 162 adults and are calculated taking into account body weights and growth factors. The resulting estimated values for Se 163 intake are: for 10 to under 13 years old 45 µg/d, for 13 to under 15 years old 60 µg/d, and for 15 years old and more, like adults, the gender is taken into account [60]. Se intake is, however, considered a mixed blessing because it has been 164 165 associated with negative health effects and it is characterized by a U-shaped relationship [61,62]. An increased risk of 166 mortality, a depressed immune system (a decrease in T lymphocytes and in the activity of Natural killer cells), and 167 cognitive decline (related with depression, anxiety, mental confusion, and hostility, epileptic seizures, and Alzheimer's) 168 have all been associated with low Se levels. Meanwhile, moderate Se levels has antiviral effects, reduces the risk of 169 autoimmune thyroid disease (Hashimoto's thyroiditis) and it is crucial for reproduction, reducing the risk of miscarriage

and improving sperm motility [61]. However, high Se levels are toxic (66 mg Se/Kg b/w) provoking diarrhea, abdominal
pain, hypotension, poor perfusion, prolonged electrocardiogram QT intervals, hypokalemia, and death six hours after
ingestion [63–65]. Urine is the dominant route of excretion of Se in monogastric. It depends on the amount of Se ingested,
the chemical form, the composition of the food, the Se status of the animal and the percentage of the glomerular filtration.
Se is also excreted by feces [66].

Se is present in grain, meat, seafood, and nuts, as well as dairy products. It is also used as a dietary supplement. In Spain, the Se content in food is low, in agreement with the low levels of Se in serum [67]. Rural areas of China, Siberia and parts of Europe are also Se-poor regions [68,69]. Severe and endemic Se deficiency is linked to Keshan disease, a myocardiopathy located in China and Kaschin-Beck disease, an endemic osteoarthritis in adolescents [70,71].

179 Se is present in a pivotal group of proteins called selenoproteins, so called because Se is incorporated in the form of 180 selenocysteine (SeCys), a selenium-containing amino acid. Se in the form of SeCys is specified in the genetic code and is 181 now recognized as proteinogenic amino acid 21 [72,73]. Moreover, it also has its own codon (UGA) which is normally the 182 signal for protein synthesis completion [74]. To date, 25 selenoproteins have been identified in the human proteome [75]. 183 Most selenoproteins have antioxidant effects and cell signaling and redox homeostasis functions like GPx family [76]. 184 There are other selenoproteins which participate in different processes. The thioredoxin reductase (TrxRs), and 185 iodothyronine deiodinase (DIOs) families participate in thyroid hormones regulation; selenoprotein W (SelW), 186 selenoprotein H (SelH), selenoprotein T (SelT), selenoprotein V (SelV) in redox regulation processes; selenoprotein P15 187 (SeP15) in apoptosis regulation. Selenoprotein P (SelP) is the only selenoprotein that contains 10 SeCys residues. It is 188 synthesized mainly in liver, regulates Se transport, storage, OS, immunomodulation and is related to the insulin resistance 189 process.

190 In mammals, the GPx family (GPx1-GPx8) participates in the defense against OS. They reduce lipid hydroperoxides to 191 their corresponding alcohols and free  $H_2O_2$  to water [77]. GPx1 is pervasive in cytosol and participates in the reduction of 192 H<sub>2</sub>O<sub>2</sub> and lipid hydroperoxides [78]. GPx1 is the most sensitive to changes in both oxidative balance and Se status: low Se 193 levels can cause a decrease in its mRNA and protein levels [79]. GPx2 is mainly expressed in the intestinal epithelium, 194 acting as an antioxidant against ingested oxidants. GPx3 is found in the extracellular space and in plasma with a 10-fold 195 lower activity than GPx1 [80,81]. GPx1, GPx2 and GPx3 operate in the aqueous phase, while GPx4 protects the 196 membranes from oxidative damage, therefore reducing hydroperoxides to lipoproteins and complex lipids [77]. GPx4 197 function is also related to the transcriptional factor NFkB and apoptosis. Recently, Ojeda et al. have published that GPx4,

198 specifically, has an indispensable role in mitochondria, preventing their oxidation, apoptosis, and NFkB dysregulation, 199 being crucial in heart function [30]. In summary, GPx4 plays an antioxidant and anti-inflammatory role, reducing 200 peroxided complex lipids integrated in biomembranes from cells and organelles, and could avoid ferroptotic cell death in 201 cardiomyocytes upon metabolic stress [82]. GPx5, containing a cysteine (Cys) instead of SeCys in the active center, is 202 characterized as a protein secreted in the epididymis. GPx6 is a selenoprotein, whose function as yet remains unknown, is 203 found in humans, but not in rats or mice, and is expressed in the olfactory epithelium and embryonic tissues. GPx7 and 204 GPx8 are CysGPxs with low GPx activity, although they are the only selenoproteins that contribute to oxidative protein 205 folding in the endoplasmic reticulum [77,80-82]. The function of fifty percent of the selenoproteome still remains 206 undefined.

207 Depending on the amount of selenium, the tissue and physiological conditions, the synthesis of this Se is altered in 208 different ways [83]. As they are produced according to a certain hierarchy, the production of some selenoproteins 209 terminates when there is a scarcity of Se [84]. An example of this concept was provided by comparing the activities of 210 GPxs: GPx1 and GPx4 in rat liver, kidney, and heart. In a Se-deficient status, GPx1 activity falls dramatically, while 211 GPx4 activity is better maintained. This hierarchy is a tissue phenomenon, since each tissue synthetizes its own 212 selenoproteins; for instance, GPx1, GPx4, and SelP are specifically found in a rodent's hepatic tissue [85], yet GPx4 is 213 found mainly in heart and GPx3 in kidney and serum [86]. The regulation of selenoprotein's synthesis in each tissue, 214 therefore, needs to be researched.

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### 4. Se homeostasis and BD during adolescence

It has recently been found that in rats BD exposure during adolescence profoundly affects Se homeostasis, leading to 216 217 severe Se depletion in serum, liver, kidney, and heart, accompanied by an imbalance in antioxidant selenoproteins' 218 expressions [30]. In this context, to demonstrate the role of Se and selenoproteins during BD exposure, the homeostasis of 219 Se has been studied in adolescent rats with BD exposure administered orally and intraperitoneally (i.p.) [29]. It was 220 demonstrated that BD during adolescence alters Se homeostasis regardless of the administration route employed. Despite 221 the fact that the BD oral group ingested less Se in diet than the i.p. group, leading to a lower Se apparent balance, the 222 changes in Se body distribution were similar in both BD models. BD rats exposed via i.p. intake a higher amount of Se in 223 diet, but this route of administration leads to higher oxidation by increasing cortisol levels, NOX activity and decreasing 224 GSH hepatic levels [33]. Consequently, more Se is needed in order to counteract this oxidation and, therefore, EtOH per 225 se, and not only because of the effect on dietary nutritional status, leads to a general body depletion of Se. Moreover, in 226 both BD models Se retention increased, in line with lower Se excretion via feces and urine. However, this effort was insufficient and a decrease in serum and hepatic Se was found in both models, correlating with a descent in the activity and expression of several antioxidant selenoproteins and with an increase in lipid, protein, and DNA oxidation [29,33]. Se supplementation therapies could be a good strategy to prevent EtOH-oxidative damage. This narrative review is focused on the use of sodium selenite supplementation administered to BD adolescent rats, examining its effect in liver, kidney, heart, and systemic oxidative balance, related in part to the above organs functioning and their cardiovascular implications.

### 233 **5. Selenium supplementation and BD during adolescence**

234 5.1. Effects in liver

235 5.1.1. BD damage in liver function

236 Alcohol is one of the greatest causes of liver-related death worldwide related to its hepatotoxicity [11], leading to the 237 alcoholic liver disease (ALD). ALD covers alcoholic fatty liver (steatosis), inflammation, fibrosis/cirrhosis, and increased 238 risk of hepatocellular carcinoma [87]. However, despite the fact that among teenagers binge-alcohol exposition is one of 239 the most widely-used intoxicating drug [88], and that it is specially pro-oxidant in liver [30] inducing hepatocyte 240 apoptosis via the mitochondrial pathway [89], the effects of BD intoxication on the liver function in teenagers are not well 241 characterized. In a recent clinical trial Binder et al. [13], it has been found that a single event of acute alcohol intoxication 242 in adolescent increases aspartate transaminase (AST) and alanine transaminase (ALT) levels. Most liver diseases course 243 with an increase in serum transaminases, especially ALT, except in ALD, where AST levels are twice those of ALT, with 244 a resulting increase in the AST/ALT ratio, being this index a specific marker of ALD [90]. Similar results of this index 245 were found in more than 90% of these teenagers, indicating damage in hepatocyte, and reflecting the fact that ALD is 246 initiating. One difference among AST and ALT lies in the fact that AST is located within the cell, linked to mitochondria, 247 while the ALT is free in the cytoplasm. This confirms what it was expected, that BD during adolescence affects the 248 hepatocyte function mainly affecting mitochondria. Experimental research have confirmed these effects on transaminases 249 serum levels after BD-exposure during adolescence and hepatocyte damage. Donohue et al. defend that acute EtOH 250 administration during early growth contributes to steatosis development and to important metabolic disturbances related 251 to future cardiovascular function.

In this context, it has been described that during adolescence BD affects mitochondrial functions and energy status by increasing the intramitochondrial NADH/NAD+ ratio, which produces ATP and ROS. The increase in ATP and ROS generated after BD exposure decreases AMP-dependent protein kinase (AMPK) activity (Figure 1a) [14]. AMPK is an important energy metabolism regulator. When a cell's energy state is diminished, AMPK activation restores energy balance by stimulating catabolic processes that generate ATP and by downregulating anabolic processes that consume
ATP [91]. It is also known, however, that the action of AMPK is suppressed by OS [92]. By generating ATP and ROS in
liver, BD in adolescent rats decreases AMPK activity leading, among others, to higher Acetyl CoA carboxylase (ACC)
activity and higher Malonyl CoA levels, increasing lipogenesis and preventing lipolysis, thus contributing to steatosis and
to the development of ALD [93].

The mechanisms by which BD affects liver function are related in part to oxidation, inflammation, apoptosis and to changes in the mitochondrial energetic balance related to lipogenesis and lipolysis, which in turn leads to steatosis. Se supplementation as an antioxidant therapy with capacity to modulate lipid profile could, therefore, be a good strategy to prevent this kind of damage.

# 265 5.1.2. Se Supplementation and antioxidant effects

266 The descent of hepatic Se levels provoked by BD during adolescence is correlated with a decrease in the expression of 267 hepatic GPx1 and GPx4, affecting liver oxidative balance (Figue1a). In the hepatocytes' cytoplasm the rest of the 268 antioxidant enzymes' (GR, CAT and SOD) activities are increased, and a high lipid and protein oxidation appears. This points to GPx1 as a key factor in hepatic cytoplasm oxidative balance [30,45] GPx4 is the only selenoprotein associated 269 270 with protecting biomembranes against oxidative insults, including mitochondria. GPx4 must be playing a crucial role in 271 the hepatic damage caused by BD, since it is at the top of the selenoproteins hierarchy and should be maintained; however, 272 it is decreased. Therefore, another stimulus apart from Se liver deposits is at work, modulating GPx4 expression. As other 273 authors affirm, this factor could be the inflammatory modulator NFkB [94]. After BD GPx4 is related to a NFkB lower 274 expression in the liver according to the greater clinical predisposition to develop an infection after acute EtOH exposition in adolescents and with greater apoptosis [30,95]. SelP P was, however, not affected by acute EtOH [30]. Therefore, its 275 276 action delivering Se to serum and other tissues, and its relationship to AMPK inactivation seems to be unaffected. 277 Surprisingly hepatic AMPK activation decreases after BD exposition, contributing to impair insulin signaling in the liver. 278 In adolescent BD rats, sodium selenite supplementation repletes hepatic Se deposits, improving principally GPx1 279 expression and activity, preventing cytosolic lipid and protein oxidation [30]. Furthermore, CAT, SOD and GR activities 280 were balanced (Figure 1b). Se supplementation increased hepatic GPx4 expression and decreased mitochondrial 281 oxidation, increasing NFkB activation, and decreasing apoptosis [30].

After Se supplementation, AMPK was upregulated. This increased orchestrated by Se supplementation does not seem to be related to SelP expression, since it was unaffected. More studies are needed in order to understand SelP function in insulin resistance. This implies that OS plays a pivotal role in AMPK inactivation by BD. This link is even more important, since it is increasingly clear that AMPK activation also has multiple actions on the inflammatory signalingprocess [96].

287 5.1.3. Physiological implications

The Se supplementation used (0.4 ppm) prevents in part cytosolic and mitochondrial oxidation leading to a better 288 289 apoptotic and inflammatory profile and a better energetic-metabolic rate in hepatocytes. Therefore, the Se 290 supplementation used in BD-exposed rats improves hepatic function; it modifies transaminases' profile by decreasing 291 AST levels and thus compensating AST/ALT ratio [30]. This therapy also increases to normal values the hepatic 292 expression of NFkB p65, in consonance with the increase in serum pro-inflammatory cytokines and chemokines (IL-6, 293 MCP-1 and CINC-1) preventing future infections. Se supplementation avoids, in part, hepatic apoptosis since it leads to a 294 decrease in the pro-apoptotic caspase-3 activation and to an increase in the anti-apoptotic TIMP-1 which protects from 295 apoptosis modulating the Bcl-2 family of proteins and decreasing Bax expression, preventing cyt. c liberation and the 296 intrinsic apoptotic pathway [30,97]. Moreover, Se supplementation increases the hepatic activation of AMPK, 297 contributing to improving both energy and the metabolic state, probably decreasing lipogenesis and increasing lipolysis, 298 thus preventing steatosis [14].

299 5.2. Effects in kidney

### 300 5.2.1. BD damage in kidney function

301 Acute EtOH exposure has deleterious structural and functional effects on kidney intimately related to hypertension (HT) 302 since EtOH consumption leads to a dysregulation of renal water and sodium excretion [98,99]. This imbalance is related 303 in part to an increase in anti-natriuretic hormones such as Aldosterone (Aldo) or Angiotensin II (AGTII) [100]. Aldo and 304 AGTII decrease the glomerular filtration rate (GFR) and constrict the peripheral vasculature, they are also related to the 305 genesis of vascular OS [101], elevating blood pressure values. In the case of repeated binge EtOH administration during 306 adolescence, a disturbance on renal electrolytes excretion, and a decrease in GFR have also been described, implying an 307 increase in water retention and high systolic blood pressure (SBP) [15,102,103]. The main mechanism that leads to all 308 these functional alterations related to oxidation seems to be the lipid peroxidation in kidney cells, the OS generated in 309 mitochondria which affects apoptosis and inflammation, and the high systemic OS generated [15]. Lipid peroxidation of 310 nephron epithelial cells interferes with carrier functions such as Na<sup>+</sup>K<sup>+</sup>-ATPase activity, affecting sodium and potassium 311 excretion [104]. After a large amount of EtOH exposure, such as that which occurs during BD, these oxidative effects are 312 greater, as well as the anti-natriuretic effect provoked. It has been reported that the high systemic OS caused by BD 313 exposure stimulates the Sympathetic Nervous System (SNS), the Hypothalamus-hypophysis-adrenal axis (HHA), and the

Renin-Angiotensin-Aldosterone System (RAAS), contributing to an increase in Aldo and AGTII concentration and to their anti-natriuretic effects, renal vasoconstriction and high SBP [105,106].

#### 316 5.2.2. Se Supplementation and antioxidant effects

317 During BD exposure BACs are high and EtOH arrives in greater amount to kidney, which actively collaborates in 318 EtOH metabolism, leading to high ROS production. The descent of kidney Se levels provoked by BD is correlated with a 319 decrease in the expression of kidney GPx1< GPx4 <GPx3 (Figure 2a). This fact, together with the antioxidant enzyme 320 imbalance that takes place (high SOD, CAT and GR activities), leads to cytosolic lipids and protein oxidation (Figure 2a) 321 [15]. Furthermore, the amount of acetaldehyde generated by ADH and CYP2E1 enters the mitochondria, where it is 322 oxidized to acetate by ALDH. This acetate together with the acetate taken up from blood, highly increases the KC and 323 Ox-Phos pathways leading to mitochondrial OS. This is related to higher caspase-3 activation and a lower NFkB 324 activation, thus increasing apoptosis (Figure 2a). The EtOH-OS-apoptosis relationship occurs mainly in tubular epithelial 325 cells [107]. Recently Li et al. found that in pigs Se deficiency disrupts oxidative balance and activates inflammation in 326 kidneys, leading to inflammatory lesions and renal tubular atrophy by downregulating, among others, selenoproteins 327 GPx1 and GPx3 [108]. However, little is known about BD renal Se deposits and NFkB related to inflammation and/or 328 fibrosis.

329 GPx3 is the main selenoprotein expressed in kidney [109]. It acts both on the proximal tubes of nephrons, reducing  $H_2O_2$ 330 to water, and acts also on plasma being the largest source of plasma GPx [86]. Kidney probably acts as a GPx3 reservoir 331 that can be mobilized when needed to combat oxidative challenges in plasma or in other parts of the body, such as the 332 challenges which occur after BD exposure. Therefore, this decrease in GPx3 expression contributes to inducing OS in 333 plasma and kidney cells after BD exposure. Moreover, kidney is also an important Se reservoir [108], since the excess of 334 Se is mostly excreted into urine or retained in kidney. After BD exposition, Se renal clearance is decreased [15], however 335 serum Se values and kidney deposits are decreased. This confirms the fact that repeated BD exposure consumed Se 336 drastically.

Sodium selenite supplementation in drinking water administered to adolescent BD rats increased GPx1 expression and activity by increasing renal Se deposits and reduced GR activity. Despite the fact that SOD and CAT activity remained high, cytosolic lipid oxidation was prevented completely (Figure 2b) [15]. Sodium selenite supplementation also increased GPx4 expression. This probably decreased mitochondrial oxidation which, in turn, decreased caspase-3 and increased NFkB activation avoiding apoptosis. GPx3 expression is also increased, thus intensifying the activity against ROS in kidney and in plasma [86,110].

#### 343 *5.2.3. Physiological implications*

344 These improvements in oxidative balance and GPxs, NFkB p65 and cleaved caspase-3 expression following Se supplementation are related to better kidney functional parameters in adolescent rats exposed to a repeated BD exposure 345 346 model. In this model, BD leads to a lower GFR, causing hypernatremia, hypokalemia, hyperaldosteronism, and a low 347 relative Aldo clearance value [15]. Therefore, BD consumption during adolescence, when the RAAS system is especially active, leads to a large hydric and electrolyte disturbance that, together with a systemic oxidative process, cause an 348 349 increase in SBP [111]. Supplementation with the antioxidant Se principally improves electrolyte balance-related functions. This is due to the fact that lipid peroxidation interferes with carrier functions such as Na<sup>+</sup>K<sup>+</sup>-ATPase activity, 350 351 increasing Na<sup>+</sup> reabsorption and K<sup>+</sup> excretion [104], especially in the renal papillary collecting duct cells [112]. Moreover, Se supplementation also reduces serum Aldo levels and SBP by decreasing systemic OS and RAAS stimulation and, since 352 353 Se improves hepatic function, by increasing its renal excretion probably by improving hepatic Aldo clearance [30,113]. 354 However, the functions related to filtration processes are not improved by Se supplementation. This could be due to the 355 fact that the GFR reduction is mainly related to the inflammatory process generated in podocytes, these latter being highly 356 OS resistant [114].

357 **5.3.** Effects in heart

#### 358 5.3.1. BD damage in heart function

359 Different studies report a J-shaped curve relationship between the amount of alcohol consumption and CVD, the dose and 360 pattern of alcohol consumption being the greatest modulators of these effects [115]. In this context, repeated BD 361 consumption during adolescence is clinically associated with a higher risk of HT, vascular dysfunction and cardiac 362 arrhythmias [3,4,16]. Acute EtOH induces general oxidation which activates the SNS and RAAS, leading to HT [11,105]. 363 The mechanisms implicated in cardiovascular damage are complex; BD in young adults produce vascular OS, 364 inflammation, activation of the SNS and RAAS, impairment of the baroreceptors, changes in endothelial and smooth cell 365 function and vascular reactivity which affects coronary arteries and heart function. Moreover, BD also directly affects 366 heart function since it provokes OS and apoptosis and increases the vulnerability of the myocardium to the development of arrhythmias by altering myocardial electrophysiological properties [116,117]. Recently Lizhuo et al. [118] have found 367 368 that BD exposition in adolescent rats impedes the normal rapid physiological growth of heart and reorients it towards 369 pathological hypertrophy. These cardiac structural alterations persist through adolescence even after cessation of EtOH 370 exposure. They also concluded that the adolescent heart is substantially more sensitive to EtOH damage than adult ones.

371 5.3.2. Se Supplementation and antioxidant effects

The excess of EtOH in blood after BD exposure could be oxidized in heart (Figure 3a). Due to the lack of ADH in the heart, cardiomyocytes metabolize EtOH mainly through the enzyme CAT, which also acts upon myocardial morphology and hemodynamics [119]. However, depending on EtOH concentration, CYP2E1 could be up-regulated in heart contributing to ROS generation [120]. These ROS generated, together with the antioxidant enzyme imbalance (high CAT and GR and low GPx1 activities) [55], leads to cytosolic oxidation of lipids and proteins.

Like in kidney, the acetate generated by EtOH metabolism together with the acetate taken up from blood in an uncontrolled manner [44], increases mitochondrial ROS overproduction (Figure 3a). This fact together with the extremely lower GPx4 expression found in BD heart, leads to mitochondrial oxidation [55], caspase-3 activation and to a higher NFkB p65 activation. In heart, prolonged and excessive ROS production can activate NFkB, leading to the activation of proinflammatory and proapoptotic pathways, increasing harmful cytokines production [121].

Since expression of GPx3 is not affected after BD exposure, despite it being halfway down the Se-specific hierarchy, it probably has another action in heart [55]. Apart from its ability to serve as an ROS scavenger in extracellular matrix spaces [122], it has been defended that an increase in GPx-3 may play a significant role in protecting cardiomyocytes from OS caused by hyperglycemia. In this context, adolescent BD rats also present hyperglycemia [30]. GPx3 could, therefore, play an important role in the heart of BD animals, preventing OS generated by hyperglycemia in myocytes and for this reason it is either conserved or derived from blood.

388 By increasing heart Se deposits, sodium selenite supplementation in adolescent BD rats increases GPx1 expression and 389 activity; thus, it decreases intracellular myocyte lipid and protein oxidation [55]. It also greatly increases GPx4 expression 390 and protects mitochondria from oxidation [123], decreasing the release of cyt. c owing to its inner membrane location in 391 mitochondria and its ability to repair cardiolipin peroxidation [124]. For these reasons, Se supplementation decreases 392 caspase-3 activation in myocytes, decreasing apoptosis [55]. Moreover, GPx4 also protects from ferroptotic cell death (a lipid ROS-induced cell death programmed) in cardiomyocytes during metabolic stress [125]. The higher GPx4 expression 393 394 could specifically interfere with NFkB activation by interleukin-1, decreasing the synthesis of leukotrienes and 395 prostanoids, modulating inflammatory process [84] (Figure 3b).

### 396 *5.3.3. Physiological implications*

397 It can be concluded that BD is related to cardiac oxidation, apoptosis and inflammation in adolescent rats and that Se 398 supplementation decreases these alterations. Previous studies find a relationship among EtOH consumption and these 399 biomolecular alterations and an increase in stroke volume and SBP [120,126,127]. However, it is also related to 400 myocardial contractile dysfunction which affects heart rate (HR), since OS alters myocardial electrophysiological 401 properties [116,117], increasing the vulnerability of the myocardium to developing arrhythmias. Moreover, it is also 402 recognized that, following BD consumption, mitochondria are the main organelles in heart damaged by OS [128]. This 403 affects their function – intimately linked to cardiovascular biology – since cardiomyocyte mitochondrial damage alters 404 calcium dynamics and leads to cardiac fibrosis [11]. In this context, it is important to point out that Se deficiency is 405 profoundly associated with cardiac pathology [105,121,129,130]. In fact, adolescent BD rats present higher SBP, DBP, 406 MBP and HR [55].

Se supplementation to BD rats protects mitochondria mainly by increasing GPx4 which is especially important for a correct cardiac energy function. These actions have a great effect on heart function by sharply decreasing cardiac HR and preventing tachycardia. Despite these important protective roles in heart, and the fact that Se supplementation improves electrolyte renal imbalance, and decreases serum Aldo values, this therapy only partially decreases SBP. This was the reason for undertaking vascular studies.

412 It has been demonstrated that BD exposition to adolescent rats leads to systemic OS by leading to lipid and protein plasma 413 oxidation and to a dangerous oxidation in lymphocytes DNA [34] together to an unbalanced antioxidant system in which 414 serum GPx (GPx3) is decreased [15]. Systemic OS is intimately related to vascular function and blood pressure (Figure 4) 415 [131]. According to that, it has been found that BD exposition increased serum vascular endothelial markers such as 416 Vascular Endothelial Growth Factor (VEGF), Connective Tissue Growth Factor (CTGF), Caveolin 1 (Cav-1) and 417 Plasminogen Activator Inhibitor-1 (tPAI-1) [55]. These results indicate that BD exposition affects vascular endothelial 418 function contributing to blood pressure dysfunctions. The beneficial actions of Se on vasculature physiology have been 419 previously described. It has anti-atherosclerotic properties possibly by regulating cholesterol (Chol) metabolism and 420 reducing OS through GPx and SelP activities [132]. Sodium selenite supplementation to BD adolescent rats by increasing 421 serum Se levels increased GPx3 activity avoiding serum lipid oxidation and lymphocyte DNA oxidation contributing to 422 lower systemic OS [15,55]. This supplementation also decreased the levels of proangiogenic growth factors VEGF and 423 CTGF, which have deleterious effects in vascular function; however, Se supplementation increased even more serum 424 Cav-1 and tPAI-1 levels in BD rats (Figure 4) [55]. Se is a mineral intimately related to lipid homeostasis [133], in this 425 context, t-PAI-1 secretion is modulated by triacylglycerol in liver [134], maybe Se and tPAI-1 synthesis could be related 426 via lipid homeostasis. In the same line, among other functions, Cav-1 plays an important role in Chol trafficking [135], 427 being crucial for controlling mitochondrial Chol levels avoiding this organelle dysfunction [136]. However, these

428 relationships among Se and t-PAI-1 and Cav-1 should be deeply studied, in order to know if these effects are beneficial or 429 not to vascular endothelial function.

430 **6.** Conclusions

431 In summary, BD is a potent pro-oxidant pattern of alcohol consumption, since during its oxidative metabolism in different 432 tissues great amount of ROS in cytoplasm and organelles -especially mitochondria- are generated. This OS affects organs 433 functions, and increased risk for developing HT and steatosis, which could lead to chronic conditions such as CVD or 434 insulin resistance in later life. Furthermore, in all the tissues studied there is a depletion of Se deposits. However, 435 antioxidant selenoproteins expressions are affected in different ways, in part by their Se-hierarchy, in part by the kind of 436 EtOH-oxidative enzymes presented in each tissue. For instance, in liver where there are high ADH and CYP2E1 437 activities, OS mainly takes place in cytosol and there is a deep decrease in cytosolic GPx1 expression. In kidney, where 438 there is a moderate activity of ADH and CYP2E1 together with an increase in acetate import from blood, GPx3 expression 439 is mainly affected. In heart, where there is a moderate activity of CAT and CYP2E1 along with an increase in acetate 440 import from blood, OS takes place mainly in mitochondria, being GPx4 expression severely impaired. The OS generated 441 is also related to inflammation, apoptosis and an unbalanced energy process in tissues. However, sodium selenite 442 supplementation to BD adolescent rats increases Se deposits in all the tissues studied and the expression of all the 443 selenoproteins affected. Therefore, further animal and human studies should be considered to support the theory that Se 444 supplementation to BD adolescents could be a cheap and efficient therapy for mitigating the adverse effects of alcohol in 445 heart, kidney, and liver functions, while decreasing the possibility of developing CVD during adolescence or later in life.

# 446 Limitations of the revision:

In order to extrapolate to humans, the results exposed above, there are two main limitations, both related to the fact that most of the studies analyzed are obtained by an experimental rat model of BD. The first one is that certain authors defend that some characteristics found in human adolescents are clearly unique, although there are other key characteristics of this developmental stage that are common across species. However, it seems that those related to alcohol intake are mainly common among human and rats [8,137]. The BD i.p. model mainly analyzed in this review assesses the validity of animal models in human psychopathology based on Face validity, Predictive validity, and Construct validity.

The second limitation of the data analyzed in this revision is related to the fact that peak BACs are not similar among humans and rats. Although in both cases BACs are higher than to 0.08 % according to BD definition, these values remain

longer in humans. However, they present similar BACs levels in the first hour after exposition [138].

456 It is also important to point that the history of the BD behavior in humans is difficult to investigate. However, forced 457 rodent models of BD, such as the i.p. model analyzed in this review, allow greater control over environmental parameters,

- 458 such as Se intake. In summary, data obtained from this revision could lead to future clinical BD-exposed teenagers'
- 459 studies, which could analyze hepatic profile and cardiovascular risk factors, related for instance to Se, GPx1 and SelP
- 460 serum levels.
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- 464 **Conflicts of Interest:** The authors declare no conflict of interest.
- 465

# 466 **References**

- K. Witkiewitz, R.Z. Litten, L. Leggio, Advances in the science and treatment of alcohol use disorder,
  Sci Adv. 5 (2019). https://doi.org/10.1126/SCIADV.AAX4043.
- 469 [2] M. Himmelreich, C.J. Lutke, E.T. Hargrove, The lay of the land : Fetal alcohol spectrum disorder
  470 (FASD) as a whole-body diagnosis, The Routledge Handbook of Social Work and Addictive
  471 Behaviors. (2020) 191–215. https://doi.org/10.4324/9780429203121-14.
- 472 [3] E.G. Vaschillo, B. Vaschillo, J.F. Buckman, S. Heiss, G. Singh, M.E. Bates, Early signs of cardiovascular
  473 dysregulation in young adult binge drinkers., Psychophysiology. 55 (2018) e13036.
  474 https://doi.org/10.1111/psyp.13036.
- M.R. Piano, L. Burke, M. Kang, S.A. Phillips, Effects of repeated binge drinking on blood pressure 475 [4] levels and other cardiovascular health metrics in young adults: National health and Nutrition 476 477 Examination Survey, 2011-2014, J Am Heart Assoc. 7 (2018)2011-2014. https://doi.org/10.1161/JAHA.118.008733. 478
- [5] N.C.F. Fagundes, L.M.P. Fernandes, R.S.D.O. Paraense, P.M.A. de Farias-Junior, F.B. Teixeira, S.M.
  Alves, J.D.J.V. Pinheiro, M.E. Crespo-López, C.S.F. Maia, R.R. Lima, Binge Drinking of Ethanol during
  Adolescence Induces Oxidative Damage and Morphological Changes in Salivary Glands of Female
  Rats, Oxid Med Cell Longev. 2016 (2016). https://doi.org/10.1155/2016/7323627.
- E.E. Bonar, M.J. Parks, M. Gunlicks-Stoessel, G.R. Lyden, C.J. Mehus, N. Morrell, M.E. Patrick, Binge
  drinking before and after a COVID-19 campus closure among first-year college students, Addictive
  Behaviors. 118 (2021) 106879. https://doi.org/10.1016/J.ADDBEH.2021.106879.
- 486 [7] S. Muzi, A. Sansò, C.S. Pace, What's Happened to Italian Adolescents During the COVID-19
  487 Pandemic? A Preliminary Study on Symptoms, Problematic Social Media Usage, and Attachment:
  488 Relationships and Differences With Pre-pandemic Peers, Frontiers in Psychiatry. 12 (2021) 590543.
  489 https://doi.org/10.3389/FPSYT.2021.590543.
- 490 [8] L. Spear, Modeling adolescent development and alcohol use in animals., Alcohol Res Health. 24 (2000)
  491 115–23. http://www.ncbi.nlm.nih.gov/pubmed/11199278 (accessed June 19, 2018).
- 492 [9] L.P. Spear, Author Correction: Effects of adolescent alcohol consumption on the brain and behaviour,
  493 Nat Rev Neurosci. 19 (2018) 439. https://doi.org/10.1038/S41583-018-0007-2.
- 494 [10] C. Tapia-Rojas, R.G. Mira, A.K. Torres, C. Jara, M.J. Pérez, E.H. Vergara, W. Cerpa, R.A. Quintanilla,
  495 Alcohol consumption during adolescence: A link between mitochondrial damage and ethanol brain
  496 intoxication, Birth Defects Research. 109 (2017) 1623–1639. https://doi.org/10.1002/bdr2.1172.

- 497 [11] P.E. Molina, S. Nelson, Binge Drinking's Effects on the Body, Alcohol Res. 39 (2018) 99–109.
  498 https://pubmed.ncbi.nlm.nih.gov/30557153/ (accessed February 10, 2021).
- [12] S.A. Jones, J.M. Lueras, B.J. Nagel, Effects of Binge Drinking on the Developing Brain, Alcohol Res. 39
   (2018) 87–96. /pmc/articles/PMC6104956/?report=abstract (accessed August 10, 2020).
- [13] C. Binder, K. Knibbe, A. Kreissl, A. Repa, M. Thanhaeuser, S. Greber-Platzer, A. Berger, B. Jilma, N.
  Haiden, Does acute alcohol intoxication cause transaminase elevations in children and adolescents?,
  Alcohol. 51 (2016) 57–62. https://doi.org/10.1016/j.alcohol.2015.12.001.
- 504 [14] F. Nogales, O. Cebadero, I. Romero-Herrera, R.M. Rua, O. Carreras, M.L. Ojeda, Selenite
  505 supplementation modulates the hepatic metabolic sensors AMPK and SIRT1 in binge drinking
  506 exposed adolescent rats by avoiding oxidative stress, Food Funct. 12 (2021) 3022–3032.
  507 https://doi.org/10.1039/D0FO02831B.
- [15] P. Sobrino, M.L. Ojeda, F. Nogales, M.L. Murillo, O. Carreras, Binge drinking affects kidney function,
  osmotic balance, aldosterone levels, and arterial pressure in adolescent rats: the potential hypotensive
  effect of selenium mediated by improvements in oxidative balance, Hypertension Research. 42 (2019)
  1495–1506. https://doi.org/10.1038/s41440-019-0265-z.
- J. Yan, J.K. Thomson, W. Zhao, X. Gao, F. Huang, B. Chen, Q. Liang, L.S. Song, M. Fill, X. Ai, Role of
  Stress Kinase JNK in Binge Alcohol-Evoked Atrial Arrhythmia, J Am Coll Cardiol. 71 (2018) 1459–
  1470. https://doi.org/10.1016/j.jacc.2018.01.060.
- 515 [17] Y. Lu, A.I. Cederbaum, CYP2E1 and oxidative liver injury by alcohol, Free Radical Biology and
  516 Medicine. 44 (2008) 723–738. https://doi.org/10.1016/j.freeradbiomed.2007.11.004.
- 517 [18] E. González-Reimers, L. Galindo-Martín, F. Santolaria-Fernández, M.J. Sánchez-Pérez, J.
  518 Alvisa-Negrín, E. García-Valdecasas-Campelo, J.M. González-Pérez, M.C. Martín-González,
  519 Prognostic value of serum selenium levels in alcoholics., Biological Trace Element Research. 125 (2008)
  520 22–29. https://doi.org/10.1007/S12011-008-8152-5.
- [19] E. González-Reimers, M.C. Martín-González, M.R. Alemán-Valls, M.J. de La Vega-Prieto, L.
  Galindo-Martín, P. Abreu-González, F. Santolaria-Fernández, Relative and Combined Effects of
  Chronic Alcohol Consumption and HCV Infection on Serum Zinc, Copper, and Selenium, Biological
  Trace Element Research. 1–3 (2009) 75–84. https://doi.org/10.1007/S12011-009-8399-5.
- [20] R.M. Rua, M.L. Ojeda, F. Nogales, J.M. Rubio, M. Romero-Gómez, J. Funuyet, M.L. Murillo, O.
  Carreras, Serum selenium levels and oxidative balance as differential markers in hepatic damage
  caused by alcohol, Life Sciences. 94 (2014) 158–163. https://doi.org/10.1016/j.lfs.2013.10.008.
- Y. Isobe, H. Asakura, H. Tsujiguchi, T. Kannon, H. Takayama, Y. Takeshita, K.A. Ishii, T. Kanamori, A.
  Hara, T. Yamashita, A. Tajima, S. Kaneko, H. Nakamura, T. Takamura, Alcohol Intake Is Associated
  With Elevated Serum Levels of Selenium and Selenoprotein P in Humans, Frontiers in Nutrition. 8
  (2021) 30. https://doi.org/10.3389/FNUT.2021.633703/BIBTEX.
- 532 [22] S.K. Dutta, P.A. Miller, L.B. Greenberg, O.A. Levander, Selenium and acute alcoholism, The American
  533 Journal of Clinical Nutrition. 38 (1983) 713–718. https://doi.org/10.1093/AJCN/38.5.713.
- 534 [23] G.A. Bjørneboe, J. Johnsen, A. Bjørneboe, J. Mørland, C.A. Drevon, Effect of heavy alcohol
  535 consumption on serum concentrations of fat-soluble vitamins and selenium., Alcohol and Alcoholism
  536 (Oxford, Oxfordshire). Supplement. 1 (1987) 533–537. https://europepmc.org/article/med/3426729
  537 (accessed February 20, 2022).
- L. Sher, Role of selenium depletion in the etiopathogenesis of depression in patient with alcoholism,
  Medical Hypotheses. 59 (2002) 330–333. https://doi.org/10.1016/S0306-9877(02)00180-9.

- 540 [25] A. Luty-Frckiewicz, Z. Jethon, L. Januszewska, Effect of smoking and alcohol consumption on the
  541 serum selenium level of Lower Silesian population, Science of The Total Environment. 285 (2002) 89–
  542 95. https://doi.org/10.1016/S0048-9697(01)00898-1.
- 543 [26] T.M. Duncan, M.C. Reed, H. Frederik Nijhou, A Population Model of Folate-Mediated One-Carbon
  544 Metabolism, Nutrients. 5 (2013) 2457. https://doi.org/10.3390/NU5072457.
- [27] Z. Zhang, Y. Guo, C. Qiu, G. Deng, M. Guo, Protective Action of Se-Supplement Against Acute
  Alcoholism Is Regulated by Selenoprotein P (SelP) in the Liver., Biological Trace Element Research.
  175 (2016) 375–387. https://doi.org/10.1007/S12011-016-0780-6.
- L. Gao, J. Yuan, Y. Cheng, M. Chen, G. Zhang, J. Wu, Selenomethionine-Dominated
  Selenium-Enriched Peanut Protein Ameliorates Alcohol-Induced Liver Disease in Mice by
  Suppressing Oxidative Stress, Foods. 10 (2021). https://doi.org/10.3390/FOODS10122979.
- M.L. Ojeda, R.M. Rua, M.L. Murillo, O. Carreras, F. Nogales, Binge drinking during adolescence
  disrupts se homeostasis and its main hepatic selenoprotein expression, Alcoholism: Clinical and
  Experimental Research. 39 (2015). https://doi.org/10.1111/acer.12707.
- [30] M.L. Ojeda, O. Carreras, P. Sobrino, M.L. Murillo, F. Nogales, Biological implications of selenium in
   adolescent rats exposed to binge drinking: Oxidative, immunologic and apoptotic balance, Toxicology
   and Applied Pharmacology. 329 (2017). https://doi.org/10.1016/j.taap.2017.05.037.
- [31] M.L. Ojeda, P. Sobrino, R.M. Rua, M. del C. Gallego-Lopez, F. Nogales, O. Carreras, Selenium, a dietary-antioxidant with cardioprotective effects, prevents the impairments in heart rate and systolic blood pressure in adolescent rats exposed to binge drinking treatment, The American Journal of Drug and Alcohol Abuse . (2021) 1–14. https://doi.org/10.1080/00952990.2021.1973485.
- [32] M.R. Piano, A. Mazzuco, M. Kang, S.A. Phillips, Cardiovascular Consequences of Binge Drinking: An
   Integrative Review with Implications for Advocacy, Policy, and Research, Alcoholism: Clinical and
   Experimental Research. 41 (2017) 487–496. https://doi.org/10.1111/acer.13329.
- 564 [33] F. Nogales, R.M. Rua, M.L. Ojeda, M.L. Murillo, O. Carreras, Oral or intraperitoneal binge drinking
  565 and oxidative balance in adolescent rats, Chemical Research in Toxicology. 27 (2014).
  566 https://doi.org/10.1021/tx5002628.
- [34] M.L. Ojeda, R.M. Rua, F. Nogales, J. Díaz-Castro, M.L. Murillo, O. Carreras, The Benefits of
  Administering Folic Acid in Order to Combat the Oxidative Damage Caused by Binge Drinking in
  Adolescent Rats., Alcohol Alcohol. 51 (2016) 235–41. https://doi.org/10.1093/alcalc/agv111.
- C.P. Nascimento, D.A. Luz, C.C.S. da Silva, C.M.R. Malcher, L.M.P. Fernandes, H.S. Dalla Santa, 570 [35] 571 A.R.Q. Gomes, M.C. Monteiro, C.H.M.A. Ribeiro, E.A. Fontes-Júnior, C.S.F. Maia, Ganoderma 572 lucidum Ameliorates Neurobehavioral Changes and Oxidative Stress Induced by Ethanol Binge 573 Drinking, Oxidative Medicine and Cellular Longevity. 2020 (2020). 574 https://doi.org/10.1155/2020/2497845.
- [36] L.M.P. Fernandes, K.S. Lopes, L.N.S. Santana, E.A. Fontes-Júnior, C.H.M.A. Ribeiro, M.C.F. Silva, R.S.
  de Oliveira Paraense, M.E. Crespo-López, A.R.Q. Gomes, R.R. Lima, M.C. Monteiro, C.S.F. Maia,
  Repeated cycles of binge-like ethanol intake in adolescent female rats induce motor function
  impairment and oxidative damage in motor cortex and liver, but not in blood, Oxidative Medicine
  and Cellular Longevity. 2018 (2018). https://doi.org/10.1155/2018/3467531.
- [37] R. Liu, Q.H. Chen, J.W. Ren, B. Sun, X.X. Cai, D. Li, R.X. Mao, X. Wu, Y. Li, Ginseng (Panax ginseng meyer) oligopeptides protect against binge drinking-induced liver damage through inhibiting oxidative stress and inflammation in rats, Nutrients. 10 (2018). https://doi.org/10.3390/nu10111665.

- [38] Y.E. Cho, B.J. Song, Pomegranate prevents binge alcohol-induced gut leakiness and hepatic
  inflammation by suppressing oxidative and nitrative stress, Redox Biology. 18 (2018) 266–278.
  https://doi.org/10.1016/j.redox.2018.07.012.
- J.H. Song, H.S. Chin, O.W. Kwon, S.J. Lim, H.K. Kim, for the D.R. Group, Effect of sulodexide in 586 [39] patients with non-proliferative diabetic retinopathy: diabetic retinopathy sulodexide study (DRESS), 587 588 Graefe's Archive for Clinical and Experimental Ophthalmology. 253 (2015)829. https://doi.org/10.1007/S00417-014-2746-8. 589
- [40] A. Banerjee, S. Mukherjee, B.K. Maji, Worldwide flavor enhancer monosodium glutamate combined
   with high lipid diet provokes metabolic alterations and systemic anomalies: An overview, Toxicology
   Reports. 8 (2021) 938–961. https://doi.org/10.1016/J.TOXREP.2021.04.009.
- A. Banerjee, D. Das, R. Paul, S. Roy, U. Das, S. Saha, S. Dey, A. Adhikary, S. Mukherjee, B.K. Maji,
  Mechanistic study of attenuation of monosodium glutamate mixed high lipid diet induced systemic
  damage in rats by Coccinia grandis, Sci Rep. 10 (2020). https://doi.org/10.1038/S41598-020-72076-6.
- 596 A. Banerjee, S. Mukherjee, B.K. Maji, Efficacy of Coccinia grandis against monosodium glutamate [42] 597 induced hepato-cardiac anomalies by inhibiting NF-kB and caspase 3 mediated signalling in rat 598 model, Human Experimental Toxicology. 40 (2021)1825-1851. and https://doi.org/10.1177/09603271211010895. 599
- 600 [43] P.E. Molina, J.D. Gardner, F.M. Souza-Smith, A.M. Whitaker, Alcohol abuse: Critical
  601 pathophysiological processes and contribution to disease burden, Physiology. 29 (2014) 203–215.
  602 https://doi.org/10.1152/physiol.00055.2013.
- [44] D.F. Wilson, F.M. Matschinsky, Ethanol metabolism: The good, the bad, and the ugly, Medical
  Hypotheses. 140 (2020). https://doi.org/10.1016/j.mehy.2020.109638.
- 605 [45] Y. Jiang, T. Zhang, P. Kusumanchi, S. Han, Z. Yang, S. Liangpunsakul, Alcohol Metabolizing Enzymes, Microsomal Ethanol Oxidizing System, Cytochrome P450 2E1, Catalase, 606 and Aldehyde 607 Dehydrogenase Alcohol-Associated Liver Disease, Biomedicines. 8 (2020)in 50. https://doi.org/10.3390/biomedicines8030050. 608
- 609 [46] C. Ceron, K. Marchi, J. Muniz, C. Tirapelli, Vascular Oxidative Stress: A Key Factor in the
  610 Development of Hypertension Associated with Ethanol Consumption, Current Hypertension
  611 Reviews. 10 (2015) 213–222. https://doi.org/10.2174/157340211004150319122736.
- 612 [47] A.W. Jones, Alcohol, its absorption, distribution, metabolism, and excretion in the body and
  613 pharmacokinetic calculations, Wiley Interdisciplinary Reviews: Forensic Science. 1 (2019) e1340.
  614 https://doi.org/10.1002/wfs2.1340.
- 615 [48] C.S. Lieber, The discovery of the microsomal ethanol oxidizing system and its physiologic and
  616 pathologic role, Drug Metabolism Reviews. 36 (2004) 511–529.
  617 https://doi.org/10.1081/DMR-200033441.
- [49] R. Teschke, Microsomal Ethanol-Oxidizing System: Success Over 50 Years and an Encouraging Future,
  Alcoholism: Clinical and Experimental Research. 43 (2019) 386–400.
  https://doi.org/10.1111/acer.13961.
- [50] B.J. Song, M. Akbar, I. Jo, J.P. Hardwick, M.A. Abdelmegeed, Translational Implications of the
  Alcohol-Metabolizing Enzymes, Including Cytochrome P450-2E1, in Alcoholic and Nonalcoholic
  Liver Disease, in: Advances in Pharmacology, Academic Press Inc., 2015: pp. 303–372.
  https://doi.org/10.1016/bs.apha.2015.04.002.

- E. Tanaka, M. Terada, S. Misawa, Cytochrome P450 2E1: Its clinical and toxicological role, Journal of
  Clinical Pharmacy and Therapeutics. 25 (2000) 165–175.
  https://doi.org/10.1046/j.1365-2710.2000.00282.x.
- A.J. Kowaltowski, A.E. Vercesi, Mitochondrial damage induced by conditions of oxidative stress, Free
   Radical Biology and Medicine. 26 (1999) 463–471. https://doi.org/10.1016/S0891-5849(98)00216-0.
- [53] D. Zhong, H. Wang, M. Liu, X. Li, M. Huang, H. Zhou, S. Lin, Z. Lin, B. Yang, Ganoderma lucidum
  polysaccharide peptide prevents renal ischemia reperfusion injury via counteracting oxidative stress,
  Scientific Reports. 5 (2015). https://doi.org/10.1038/srep16910.
- [54] Z. Zhou, X. Sun, Y. James Kang, Metallothionein protection against alcoholic liver injury through
  inhibition of oxidative stress, Experimental Biology and Medicine. 227 (2002) 214–222.
  https://doi.org/10.1177/153537020222700310.
- M.L. Ojeda, P. Sobrino, R.M. Rua, M.C. Gallego-Lopez, F. Nogales, O. Carreras, Selenium, a
  dietary-antioxidant with cardioprotective effects, prevents the impairments in heart rate and systolic
  blood pressure in adolescent rats exposed to binge drinking, The American Journal of Drug and
  Alcohol Abuse. (2021).
- E.C. Schlorff, K. Husain, S.M. Somani, Dose- and time-dependent effects of ethanol on plasma antioxidant system in rat, Alcohol. 17 (1999) 97–105. https://doi.org/10.1016/S0741-8329(98)00039-1.
- 642 M. del C. Gallego-Lopez, M.L. Ojeda, I. Romero-Herrera, F. Nogales, O. Carreras, Folic Acid [57] Homeostasis and Its Pathways Related to Hepatic Oxidation in Adolescent Rats Exposed to Binge 643 Drinking, Antioxidants 2022, Vol. 362. 11 (2022)644 11, Page 362. 645 https://doi.org/10.3390/ANTIOX11020362.
- 646 [58] M.P. Rayman, The importance of selenium to human health, Lancet. 356 (2000) 233–241.
   647 https://doi.org/10.1016/S0140-6736(00)02490-9.
- 648 [59] F. Hiller, K. Besselt, S. Deubel, R. Brigelius-Flohé, A.P. Kipp, GPx2 induction is mediated through
  649 STAT transcription factors during acute colitis, Inflammatory Bowel Diseases. 21 (2015) 2078–2089.
  650 https://doi.org/10.1097/MIB.0000000000464.
- [60] A.P. Kipp, D. Strohm, R. Brigelius-Flohé, L. Schomburg, A. Bechthold, E. Leschik-Bonnet, H. Heseker,
  Revised reference values for selenium intake, Journal of Trace Elements in Medicine and Biology. 32
  (2015) 195–199. https://doi.org/10.1016/j.jtemb.2015.07.005.
- [61] M.P. Rayman, Selenium intake, status, and health: a complex relationship, Hormones. 19 (2020) 9–14.
  https://doi.org/10.1007/s42000-019-00125-5.
- 656 [62] J. Brozmanová, D. Mániková, V. Vlčková, M. Chovanec, Selenium: A double-edged sword for defense 657 and offence Archives of Toxicology. 84 (2010)919-938. in cancer. https://doi.org/10.1007/s00204-010-0595-8. 658
- [63] K.A. See, P.S. Lavercombe, J. Dillon, R. Ginsberg, Accidental death from acute selenium poisoning,
  Medical Journal of Australia. 185 (2006) 388–389. https://doi.org/10.5694/j.1326-5377.2006.tb00616.x.
- [64] R. Williams, A. Ansford, Acute selenium toxicity: Australia's second fatality [12], Pathology. 39 (2007)
  289–290. https://doi.org/10.1080/00313020701230682.
- N. Hadrup, G. Ravn-Haren, Acute human toxicity and mortality after selenium ingestion: A review, 663 [65] of Trace in Medicine Biology. 664 Journal Elements and 58 (2020). 665 https://doi.org/10.1016/j.jtemb.2019.126435.

- [66] L.F.C. Pedrosa, A.K. Motley, T.D. Stevenson, K.E. Hill, R.F. Burk, Fecal selenium excretion is
  regulated by dietary selenium intake, Biological Trace Element Research. 149 (2012) 377–381.
  https://doi.org/10.1007/s12011-012-9430-9.
- 669 [67] F.J. López-Bellido Garrido, L. López Bellido, Selenio y salud; valores de referencia y situación actual
  670 de la población Española, Nutricion Hospitalaria. 28 (2013) 1396–1406.
  671 https://doi.org/10.3305/nh.2013.28.5.6634.
- 672 [68] O.K. Chun, A. Floegel, S.J. Chung, C.E. Chung, W.O. Song, S.I. Koo, Estimation of antioxidant intakes
  673 from diet and supplements in U.S. adults, Journal of Nutrition. 140 (2010) 317–324.
  674 https://doi.org/10.3945/jn.109.114413.
- [69] A.P. Kipp, D. Strohm, R. Brigelius-Flohé, L. Schomburg, A. Bechthold, E. Leschik-Bonnet, H. Heseker,
  Revised reference values for selenium intake, Journal of Trace Elements in Medicine and Biology. 32
  (2015) 195–199. https://doi.org/10.1016/j.jtemb.2015.07.005.
- A.C. Pappas, E. Zoidis, P.F. Surai, G. Zervas, Selenoproteins and maternal nutrition, Comparative
  Biochemistry and Physiology B Biochemistry and Molecular Biology. 151 (2008) 361–372.
  https://doi.org/10.1016/j.cbpb.2008.08.009.
- [71] O.M. Guillin, C. Vindry, T. Ohlmann, L. Chavatte, Selenium, selenoproteins and viral infection,
   Nutrients. 11 (2019). https://doi.org/10.3390/nu11092101.
- 683 [72] M.P. Rayman, The argument for increasing selenium intake, Proceedings of the Nutrition Society. 61
  684 (2002) 203–215. https://doi.org/10.1079/pns2002153.
- [73] V.N. Gladyshev, Identity, evolution and function of selenoproteins and selenoprotein genes, in:
  Selenium, Springer US, 2001: pp. 99–114. https://doi.org/10.1007/978-1-4615-1609-5\_9.
- 687 [74] C. Vindry, T. Ohlmann, L. Chavatte, Translation regulation of mammalian selenoproteins, Biochimica
  688 et Biophysica Acta General Subjects. 1862 (2018) 2480–2492.
  689 https://doi.org/10.1016/j.bbagen.2018.05.010.
- [75] J. Joseph, J. Loscalzo, Selenistasis: Epistatic effects of selenium on cardiovascular phenotype,
   Nutrients. 5 (2013) 340–358. https://doi.org/10.3390/nu5020340.
- [76] V.M. Labunskyy, D.L. Hatfield, V.N. Gladyshev, Selenoproteins: Molecular pathways and
  physiological roles, Physiological Reviews. 94 (2014) 739–777.
  https://doi.org/10.1152/physrev.00039.2013.
- 695 [77] R. Brigelius-Flohé, M. Maiorino, Glutathione peroxidases, Biochimica et Biophysica Acta (BBA) 696 General Subjects. 1830 (2013) 3289–3303. https://doi.org/10.1016/j.bbagen.2012.11.020.
- 697 [78] S. Gromer, J.K. Eubel, B.L. Lee, J. Jacob, Human selenoproteins at a glance, Cellular and Molecular
  698 Life Sciences. 62 (2005) 2414–2437. https://doi.org/10.1007/s00018-005-5143-y.
- R.A. Sunde, K.M. Thompson, Dietary selenium requirements based on tissue selenium concentration
  and glutathione peroxidase activities in old female rats, Journal of Trace Elements in Medicine and
  Biology. 23 (2009) 132–137. https://doi.org/10.1016/j.jtemb.2009.02.002.
- [80] L. v. Papp, A. Holmgren, K.K. Khanna, Selenium and selenoproteins in health and disease,
   Antioxidants and Redox Signaling. 12 (2010) 793–795. https://doi.org/10.1089/ars.2009.2973.

[81] M.A. Reeves, P.R. Hoffmann, The human selenoproteome: Recent insights into functions and
regulation, Cellular and Molecular Life Sciences. 66 (2009) 2457–2478.
https://doi.org/10.1007/s00018-009-0032-4.

R. Brigelius-Flohé, L. Flohé, Regulatory Phenomena in the Glutathione Peroxidase Superfamily,
 Antioxidants and Redox Signaling. 33 (2020) 498–516. https://doi.org/10.1089/ars.2019.7905.

- [83] R.F. Burk, K.E. Hill, Regulation of Selenium Metabolism and Transport, Annual Review of Nutrition.
  35 (2015) 109–134. https://doi.org/10.1146/annurev-nutr-071714-034250.
- [84] R. Brigelius-Flohé, Glutathione peroxidases and redox-regulated transcription factors, in: Biological
   Chemistry, 2006: pp. 1329–1335. https://doi.org/10.1515/BC.2006.166.
- [85] K. Brown, J. Arthur, Selenium, selenoproteins and human health: a review, Public Health Nutrition. 4
  (2001) 593–599. https://doi.org/10.1079/phn2001143.
- [86] G.E. Olson, J.C. Whitin, K.E. Hill, V.P. Winfrey, A.K. Motley, L.M. Austin, J. Deal, H.J. Cohen, R.F.
  Burk, Extracellular glutathione peroxidase (Gpx3) binds specifically to basement membranes of
  mouse renal cortex tubule cells., Am J Physiol Renal Physiol. 298 (2010) F1244-53.
  https://doi.org/10.1152/ajprenal.00662.2009.
- [87] M. Adachi, D.A. Brenner, Clinical syndromes of alcoholic liver disease, Digestive Diseases. 23 (2006)
  255–263. https://doi.org/10.1159/000090173.
- K.M. Lisdahl, R. Thayer, L.M. Squeglia, T.M. McQueeny, S.F. Tapert, Recent binge drinking predicts 721 [88] 722 smaller cerebellar volumes in adolescents, Psychiatry Res. 211 (2013)17. 723 https://doi.org/10.1016/J.PSCYCHRESNS.2012.07.009.
- [89] X. Tian, Y. Hu, M. Li, K. Xia, J. Yin, J. Chen, Z. Liu, Carnosic acid attenuates acute ethanol-induced
  liver injury via a SIRT1/p66Shc-mediated mitochondrial pathway, Can J Physiol Pharmacol. 94 (2016)
  416–425. https://doi.org/10.1139/CJPP-2015-0276.
- [90] D. Sorbi, J. Boynton, K.D. Lindor, The ratio of aspartate aminotransferase to alanine aminotransferase:
   potential value in differentiating nonalcoholic steatohepatitis from alcoholic liver disease, The
   American Journal of Gastroenterology. 94 (1999) 1018–1022.
   https://doi.org/10.1016/S0002-9270(99)00061-1.
- [91] N.B. Ruderman, X.J. Xu, L. Nelson, J.M. Cacicedo, A.K. Saha, F. Lan, Y. Ido, AMPK and SIRT1: A
  long-standing partnership?, American Journal of Physiology Endocrinology and Metabolism. 298
  (2010). https://doi.org/10.1152/ajpendo.00745.2009.
- P. Ramadori, F.J. Cubero, C. Liedtke, C. Trautwein, Y.A. Nevzorova, Alcohol and Hepatocellular 734 [92] (2017). 735 Carcinoma: Adding Fuel to the Flame., Cancers (Basel). 9 736 https://doi.org/10.3390/cancers9100130.
- Z. Jiang, J. Zhou, D. Zhou, Z. Zhu, L. Sun, A.A. Nanji, The adiponectin-SIRT1-AMPK pathway in alcoholic fatty liver disease in the rat, Alcoholism: Clinical and Experimental Research. 39 (2015) 424–433. https://doi.org/10.1111/acer.12641.
- 740 [94] A. Wullaert, G. van Loo, K. Heyninck, R. Beyaert, Hepatic tumor necrosis factor signaling and nuclear
  741 factor-κB: Effects on liver homeostasis and beyond, Endocrine Reviews. 28 (2007) 365–386.
  742 https://doi.org/10.1210/er.2006-0031.
- R. Griffin, A.M. Poe, J.M. Cross, L.W. Rue, G. McGwin, The association between blood alcohol level
  and infectious complications among burn patients, Journal of Burn Care and Research. 30 (2009) 395–
  399. https://doi.org/10.1097/BCR.0b013e3181a28966.
- [96] S.J. Mancini, I.P. Salt, Investigating the role of AMPK in inflammation, in: Methods in Molecular
  Biology, Humana Press Inc., 2018: pp. 307–319. https://doi.org/10.1007/978-1-4939-7598-3\_20.
- 748 Ashutosh, M. Garg, S. Sundar, R. Duncan, H.L. Nakhasi, N. Goyal, Downregulation of [97] 749 mitogen-activated protein kinase 1 of Leishmania donovani field isolates is associated with antimony 750 resistance, Antimicrobial Agents and Chemotherapy. 56 (2012)518-525. 751 https://doi.org/10.1128/AAC.00736-11.

- [98] S.K. Das, D.M. Vasudevan, Alcohol induced effects on kidney, Indian Journal of Clinical Biochemistry.
  23 (2008) 4–9. https://doi.org/10.1007/s12291-008-0003-9.
- A. Matsumoto, Y. Nagasawa, R. Yamamoto, M. Shinzawa, Y. Hasuike, T. Kuragano, Y. Isaka, T.
  Nakanishi, K. Iseki, K. Yamagata, K. Tsuruya, H. Yoshida, S. Fujimoto, K. Asahi, T. Moriyama, T.
  Watanabe, The association of alcohol and smoking with CKD in a Japanese nationwide cross-sectional
  survey, Hypertension Research. 40 (2017) 771–778. https://doi.org/10.1038/hr.2017.25.
- [100] J.E. Hall, The Kidney, Hypertension, and Obesity, Hypertension. 41 (2003) 625–633.
   https://doi.org/10.1161/01.HYP.0000052314.95497.78.
- [101] J. González, N. Valls, R. Brito, R. Rodrigo, Essential hypertension and oxidative stress: New insights.,
   World J Cardiol. 6 (2014) 353–66. https://doi.org/10.4330/wjc.v6.i6.353.
- 762 R. Rodrigo, G. Rivera, Renal damage mediated by oxidative stress: A hypothesis of protective effects [102] 763 of red wine, Free Radical Biology and Medicine. 33 (2002)409-422. https://doi.org/10.1016/S0891-5849(02)00908-5. 764
- [103] H.-N. Kim, S.-H. Kim, S.-W. Song, Is Alcohol Drinking Associated with Renal Impairment in the
  General Population of South Korea?, Kidney and Blood Pressure Research. 39 (2014) 40–49.
  https://doi.org/10.1159/000355775.
- [104] R. Rodrigo, L. Thielemann, M. Olea, P. Muñoz, M. Cereceda, M. Orellana, Effect of ethanol ingestion
  on renal regulation of water and electrolytes., Arch Med Res. 29 (1998) 209–18.
  http://www.ncbi.nlm.nih.gov/pubmed/9775453 (accessed June 1, 2018).
- 771 T. Yang, C. Xu, Physiology and pathophysiology of the intrarenal renin-angiotensin system: An [105] 772 Journal the American Society of Nephrology. 28 (2017)1040-1049. update, of 773 https://doi.org/10.1681/ASN.2016070734.
- [106] M.R. Piano, A. Mazzuco, M. Kang, S.A. Phillips, Cardiovascular Consequences of Binge Drinking: An
   Integrative Review with Implications for Advocacy, Policy, and Research, Alcoholism: Clinical and
   Experimental Research. 41 (2017) 487–496. https://doi.org/10.1111/acer.13329.
- [107] C. Latchoumycandane, L.E. Nagy, T.M. McIntyre, Chronic ethanol ingestion induces oxidative kidney
   injury through taurine-inhibitable inflammation, Free Radical Biology and Medicine. 69 (2014) 403–
   416. https://doi.org/10.1016/j.freeradbiomed.2014.01.001.
- [108] S. Li, Q. Zhao, K. Zhang, W. Sun, X. Jia, Y. Yang, J. Yin, C. Tang, J. Zhang, Se deficiency induces renal
   pathological changes by regulating selenoprotein expression, disrupting redox balance, and
   activating inflammation, Metallomics. 12 (2020) 1576–1584. https://doi.org/10.1039/d0mt00165a.
- [109] K.M. Barnes, J.K. Evenson, A.M. Raines, R.A. Sunde, Transcript analysis of the selenoproteome indicates that dietary selenium requirements of rats based on selenium-regulated selenoprotein mRNA levels are uniformly less than those based on glutathione peroxidase activity., J Nutr. 139 (2009) 199–206. https://doi.org/10.3945/jn.108.098624.
- [110] R.F. Burk, G.E. Olson, V.P. Winfrey, K.E. Hill, D. Yin, Glutathione peroxidase-3 produced by the kidney binds to a population of basement membranes in the gastrointestinal tract and in other tissues, American Journal of Physiology-Gastrointestinal and Liver Physiology. 301 (2011) G32–G38. https://doi.org/10.1152/ajpgi.00064.2011.
- 791 A.R. Willey, R.I. Anderson, M. Morales, R.L. Ramirez, L.P. Spear, Effects of ethanol administration on [111] 792 corticosterone adolescent and adult Alcohol. (2012)29-36. levels in rats, 46 793 https://doi.org/10.1016/j.alcohol.2011.08.005.

- 794 [112] R. Rodrigo, S. Trujillo, C. Bosco, M. Orellana, L. Thielemann, J. Araya, Changes in (Na + K)-adenosine triphosphatase activity and ultrastructure of lung and kidney associated with oxidative stress 795 796 induced by acute ethanol intoxication., Chest. 121 (2002)589-96. 797 http://www.ncbi.nlm.nih.gov/pubmed/11834676 (accessed June 1, 2018).
- [113] L. Ojeda, F. Nogales, L. Murillo, O. Carreras, The role of folic acid and selenium against oxidative
  damage from ethanol in early life programming: A review, Biochemistry and Cell Biology. 96 (2018).
  https://doi.org/10.1139/bcb-2017-0069.
- [114] C. Latchoumycandane, L.E. Nagy, T.M. McIntyre, Myeloperoxidase formation of PAF receptor
   ligands induces PAF receptor-dependent kidney injury during ethanol consumption, Free Radical
   Biology and Medicine. 86 (2015) 179–190. https://doi.org/10.1016/j.freeradbiomed.2015.05.020.
- R. Cífková, A. Krajčoviechová, Alcohol and cardiovascular disease: Position paper of the Czech
  society of cardiology, Central European Journal of Public Health. 27 (2019) S6–S9.
  https://doi.org/10.21101/cejph.a5998.
- [116] K. Husain, R.A. Ansari, L. Ferder, Alcohol-induced hypertension: Mechanism and prevention, World
   Journal of Cardiology. 6 (2014) 245. https://doi.org/10.4330/wjc.v6.i5.245.
- [117] Y. Wang, J. Zhao, W. Yang, Y. Bi, J. Chi, J. Tian, W. Li, High-dose alcohol induces reactive oxygen
   species-mediated apoptosis via PKC-β/p66Shc in mouse primary cardiomyocytes, Biochemical and
   Biophysical Research Communications. 456 (2015) 656–661. https://doi.org/10.1016/j.bbrc.2014.12.012.
- [118] L. Ai, E. Perez, A. Asimes, T. Kampaengsri, M. Heroux, A. Zlobin, M.A. Hiske, C.S. Chung, T.R. Pak,
  J.A. Kirk, Binge alcohol exposure in adolescence impairs normal heart growth, J Am Heart Assoc. 9
  (2020) 15611. https://doi.org/10.1161/JAHA.119.015611.
- [119] H.D. Fahimi, M. Kino, L. Hicks, K.A. Thorp, W.H. Abelman, Increased myocardial catalase in rats fed
  ethanol, American Journal of Pathology. 96 (1979) 373–390.
- [120] R.H. Zhang, J.Y. Gao, H.T. Guo, G.I. Scott, A.R. Eason, X.M. Wang, J. Ren, Inhibition of CYP2E1
  attenuates chronic alcohol intake-induced myocardial contractile dysfunction and apoptosis,
  Biochimica et Biophysica Acta Molecular Basis of Disease. 1832 (2013) 128–141.
  https://doi.org/10.1016/j.bbadis.2012.08.014.
- [121] S. Tanguy, S. Grauzam, J. de Leiris, F. Boucher, Impact of dietary selenium intake on cardiac health:
  Experimental approaches and human studies, Molecular Nutrition & Food Research. 56 (2012) 1106–
  1121. https://doi.org/10.1002/mnfr.201100766.
- P.R. Hoffmann, S.C. Hoge, P.-A. Li, F.W. Hoffmann, A.C. Hashimoto, M.J. Berry, The selenoproteome
  exhibits widely varying, tissue-specific dependence on selenoprotein P for selenium supply, Nucleic
  Acids Research. 35 (2007) 3963–3973. https://doi.org/10.1093/nar/gkm355.
- [123] E.A. Knopp, T.L. Arndt, K.L. Eng, M. Caldwell, R.C. LeBoeuf, S.S. Deeb, K.D. O'Brien, Murine
  phospholipid hydroperoxide glutathione peroxidase: cDNA sequence, tissue expression, and
  mapping., Mamm Genome. 10 (1999) 601–5. https://doi.org/10.1007/s003359901053.
- [124] H. Liang, Q. Ran, Y.C. Jang, D. Holstein, J. Lechleiter, T. McDonald-Marsh, A. Musatov, W. Song, H.
  van Remmen, A. Richardson, Glutathione peroxidase 4 differentially regulates the release of
  apoptogenic proteins from mitochondria., Free Radic Biol Med. 47 (2009) 312–20.
  https://doi.org/10.1016/j.freeradbiomed.2009.05.012.
- [125] T.J. Park, J.H. Park, G.S. Lee, J.Y. Lee, J.H. Shin, M.W. Kim, Y.S. Kim, J.Y. Kim, K.J. Oh, B.S. Han, W.K.
  Kim, Y. Ahn, J.H. Moon, J. Song, K.H. Bae, D.H. Kim, E.W. Lee, S.C. Lee, Quantitative proteomic

- analyses reveal that GPX4 downregulation during myocardial infarction contributes to ferroptosis in
   cardiomyocytes, Cell Death and Disease. 10 (2019) 835. https://doi.org/10.1038/s41419-019-2061-8.
- [126] N.A. Umoh, R.K. Walker, R.M. Millis, M. Al-Rubaiee, P.R. Gangula, G.E. Haddad, Calcitonin
  Gene-Related Peptide Regulates Cardiomyocyte Survival through Regulation of Oxidative Stress by
  PI3K/Akt and MAPK Signaling Pathways., Ann Clin Exp Hypertens. 2 (2014) 1007.
  http://www.ncbi.nlm.nih.gov/pubmed/25478604 (accessed April 25, 2020).
- [127] M. Katary, A.A. Abdel-Rahman, Alcohol suppresses cardiovascular diurnal variations in male
  normotensive rats: Role of reduced PER2 expression and CYP2E1 hyperactivity in the heart, Alcohol.
  844 89 (2020) 27–36. https://doi.org/10.1016/j.alcohol.2020.08.001.
- 845 M. Kannan, L. Wang, Y.J. Kang, Myocardial oxidative stress and toxicity induced by acute ethanol [128] in Experimental Biology and Medicine. 229 (2004)553-559. 846 exposure mice, 847 https://doi.org/10.1177/153537020422900614.
- [129] A.H. Rose, P.R. Hoffmann, Selenoproteins and cardiovascular stress, Thrombosis and Haemostasis.
  113 (2015) 494–504. https://doi.org/10.1160/TH14-07-0603.
- [130] J. Joseph, J. Loscalzo, Selenistasis: epistatic effects of selenium on cardiovascular phenotype.,
   Nutrients. 5 (2013) 340–58. https://doi.org/10.3390/nu5020340.
- [131] B. Buijsse, D.-H. Lee, L. Steffen, R.R. Erickson, R. v Luepker, D.R. Jacobs, J.L. Holtzman, Low serum
  glutathione peroxidase activity is associated with increased cardiovascular mortality in individuals
  with low HDLc's., PLoS One. 7 (2012) e38901. https://doi.org/10.1371/journal.pone.0038901.
- [132] L. Guo, J. Xiao, H. Liu, H. Liu, Selenium nanoparticles alleviate hyperlipidemia and vascular injury in
   ApoE-deficient mice by regulating cholesterol metabolism and reducing oxidative stress, Metallomics.
   12 (2020) 204–217. https://doi.org/10.1039/c9mt00215d.
- [133] J.Q. Huang, J.C. Zhou, Y.Y. Wu, F.Z. Ren, X.G. Lei, Role of glutathione peroxidase 1 in glucose and
  lipid metabolism-related diseases, Free Radical Biology and Medicine. 127 (2018) 108–115.
  https://doi.org/10.1016/j.freeradbiomed.2018.05.077.
- [134] C. Banfi, P. Risé, L. Mussoni, C. Galli, E. Tremoli, Linoleic acid enhances the secretion of plasminogen
   activator inhibitor type 1 by HepG2 cells, Journal of Lipid Research. 38 (1997) 860–869.
- [135] T.M. Williams, M.P. Lisanti, The Caveolin genes: From cell biology to medicine, Annals of Medicine.
  36 (2004) 584–595. https://doi.org/10.1080/07853890410018899.
- [136] M. Bosch, M. Marí, S.P. Gross, J.C. Fernández-Checa, A. Pol, Mitochondrial cholesterol: A connection
  between caveolin, metabolism, and disease, Traffic. 12 (2011) 1483–1489.
  https://doi.org/10.1111/j.1600-0854.2011.01259.x.
- 868 [137] B. Lees, L.R. Meredith, A.E. Kirkland, B.E. Bryant, L.M. Squeglia, Effect of alcohol use on the
  adolescent brain and behavior, Pharmacology Biochemistry and Behavior. 192 (2020) 172906.
  870 https://doi.org/10.1016/J.PBB.2020.172906.
- [138] J. Jeanblanc, B. Rolland, F. Gierski, M.P. Martinetti, M. Naassila, Animal models of binge drinking,
  current challenges to improve face validity, Neuroscience & Biobehavioral Reviews. 106 (2019) 112–
  121. https://doi.org/10.1016/J.NEUBIOREV.2018.05.002.
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# 879 FIGURE CAPTIONS.

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881 Fig. 1 EtOH metabolism in liver: oxidative implications. Effects of sodium selenite supplementation (A) EtOH is 882 oxidized in hepatocytes, mostly through the alcohol dehydrogenase (ADH) enzyme, which in turn produces an increase in 883 cytoplasmic NADH/NAD<sup>+</sup> ratio. In BD exposure, ADH is saturated (KM < 5 mM) and CYP2E1 increases its activity, 884 generating a large amount of ROS, that together with an imbalance of the antioxidant enzymes Superoxide Dismutase 885 (SOD), Catalase (CAT), Glutathione Peroxidase 1 (GPx1) and Glutathione Reductase (GR), leads to cytosolic lipid and 886 protein oxidation (OS). This large amount of ADH- and CYP2E1-generated acetaldehyde enters the mitochondria and is 887 oxidized into acetate by acetaldehyde dehydrogenase (ALDH), increasing intramitochondrial NADH/NAD+ ratio. The 888 acetate formed is converted into Acetyl CoA which enters the Krebs cycle (KC) and via oxidative phosphorylation 889 (Ox-Phos) produces ATP and ROS. At blood alcohol levels greater than 0.01 g/dL, hepatic KC is markedly suppressed, 890 and the remaining acetate is exported into the blood. The increase in ATP and ROS production in mitochondria decreases 891 AMP-dependent protein kinase (AMPK) activity. This decrease leads to higher Acetyl CoA carboxylase (ACC) activity and higher Malonyl CoA levels which increase lipogenesis and prevents lipolysis. High mitochondrial ROS production, 892 together with lower GPx4 expression, leads to mitochondrial OS. This, in turn, is related to higher caspase-3 activation 893 894 and apoptosis, as well as to a lower NFkB activation. Despite the lower Se deposits in liver caused by BD, expression of 895 the main hepatic selenoprotein SelP is not affected by BD exposure, and, therefore, its action delivering Se to serum and 896 other tissues seems to be unaffected; (B) Sodium selenite supplementation in adolescent BD rats by increasing hepatic Se 897 deposits, increases GPx1 expression and activity, preventing cytosolic lipid and protein oxidation. It also increases GPx4 expression and decreases mitochondrial OS. This latter reduces apoptosis, increases NFkB activation and increases 898 AMPK activity, while also improving hepatic lipid profile. Solid lines and hatched lines indicate stimulatory and 899 900 inhibitory actions, respectively.

901 Fig. 2 EtOH metabolism in kidney: oxidative implications. Effects of sodium selenite supplementation (A) EtOH is 902 mostly oxidized in hepatocytes, but when EtOH arrives in large amounts to blood, as occurs in BD exposure, other tissues 903 collaborate with liver. This is the case of kidney; kidney cells contain the enzyme alcohol dehydrogenase (ADH), which 904 in turn produces an increase in cytoplasmic NADH/NAD<sup>+</sup> ratio. If this is not sufficient, it could increase CYP2E1 activity 905 generating large amount of ROS, which together with antioxidant enzyme imbalance leads to cytosolic lipids and protein 906 oxidation. The amount of acetaldehyde generated by ADH and CYP2E1 enters the mitochondria and is oxidized to acetate 907 by acetaldehyde dehydrogenase (ALDH), this acetate production, together with the acetate taken up from blood increases 908 intramitochondrial NADH/NAD<sup>+</sup> ratio. Acetate is converted to Acetyl CoA which enters the Krebs cycle (KC) and via 909 oxidative phosphorylation (Ox-Phos) produces ATP and ROS. Mitochondrial ROS production, together with a lower 910 GPx4 expression, leads to mitochondrial OS. This, in turn, is related to higher caspase-3 activation and apoptosis, and to a 911 lower NFkB activation. The lower Se deposits in kidney caused by BD also affects expression of the main kidney 912 selenoprotein, GPx3, which decreases. Therefore, its action delivering Se to serum decreases; (B) Sodium selenite 913 supplementation in adolescent BD rats by increasing renal Se deposits increases GPx1 expression and activity, preventing 914 cytosolic lipid oxidation. Supplementation also increases GPx4 expression and decreases mitochondrial OS, decreasing 915 apoptosis and increasing NFkB activation. It also increases GPx3 expression which probably will shunt Se to serum. Solid 916 lines and hatched lines indicate stimulatory and inhibitory actions, respectively.

917 Fig. 3 EtOH metabolism in heart: oxidative implications. Effects of sodium selenite supplementation (A) Heart, due 918 to the lack of ADH, may oxidize some ethanol mainly through CAT and/or CYP2E1, but these rates are low, however, it 919 could generate ROS that joint to an imbalanced antioxidant system leads to cytosolic oxidation of lipids and proteins. The 920 acetaldehyde generated by CAT and CYP2E1 enters the mitochondria and is oxidized to acetate by acetaldehyde 921 dehydrogenase (ALDH). This acetate, together with the acetate taken up from blood in an uncontrolled manner, increases 922 the intramitochondrial NADH/NAD<sup>+</sup> ratio. Acetate is converted to Acetyl CoA which then enters the Krebs cycle (KC) 923 and via oxidative phosphorylation (Ox-Phos) produces ATP and ROS. Mitochondrial ROS production, together with the 924 extremely low GPx4 expression, leads to mitochondrial OS. This, in turn, is related to higher caspase-3 activation and 925 apoptosis, and to a higher NFkB activation (fibrosis). The lower heart Se deposits caused by BD do not affect the 926 extracellular expression of GPx3 in heart, a protein is related to serum glucose alteration; (B) Sodium selenite 927 supplementation in adolescent BD rats increases Se heart deposits; increases GPx1 expression and activity preventing 928 cytosolic lipid and protein oxidation. It also increases GPx4 expression and decreases mitochondrial OS, thus decreasing 929 apoptosis and NFkB activation. Solid lines and hatched lines indicate stimulatory and inhibitory actions, respectively.

Fig. 4 EtOH metabolism in plasma. Effects of sodium selenite supplementation High EtOH consumption leads to large amount of acetate and ROS in plasma, but it also provokes an unbalanced antioxidant system in which serum GPx (GPx3) decreases, generating lipid and protein oxidation in serum. BD exposure also generates profound oxidation in lymphocytes DNA; therefore, systemic OS appears. This OS could be related to the increase in serum endothelial markers such as Vascular Endothelial Growth Factor (VEGF), Connective Tissue Growth Factor (CTGF), Caveolin 1 (Cav-1) and Plasminogen Activator Inhibitor-1 (tPAI-1). By increasing serum Se levels, sodium selenite supplementation in adolescent BD rats increases GPx3 activity preventing serum lipid oxidation and lymphocyte DNA oxidation, leading to 938 lower systemic OS. This supplementation also decreases the levels of proangiogenic growth factors VEGF and CTGF, something that is considered beneficial; however, the increase in serum Cav-1 and tPAI-1 was even greater.