Supplemental Digital Content 1. Sample Collection and Biomarker Analysis Methods

Whole blood was collected, plasma or serum was generated at the study site, and samples were shipped frozen on dry ice to the Q2 Solutions central laboratory (Valencia, CA, or Edinburgh, UK), stored frozen at −70°C, and transferred or shipped frozen to the Q2 Solutions biomarker laboratory (Valencia, CA; all biomarkers except for D-dimer) or an affiliated laboratory (Quest Diagnostics Nichols Institute, San Juan Capistrano, CA; D-dimer analysis) for short-term storage (−70°C) and analysis.

Data were acquired within the stability period of storage for each biomarker: 1 month for D-dimer and fatty acid binding protein-2, 6 months for C-reactive protein and soluble CD163, 10 months for soluble vascular cell adhesion molecule-1, 12 months for interleukin-6, and 24 months for soluble CD14. Highly sensitive C-reactive protein in serum was assessed by turbidimetric detection using an Olympus AU chemistry analyzer (Beckman Coulter, Brea, CA). Soluble CD14, soluble CD163, and fatty acid binding protein-2 in serum were assessed by sandwich enzyme-linked immunosorbent assays (ELISA) using a GENios™ Pro microplate reader (Tecan, Männedorf, Switzerland). Highly sensitive interleukin-6 in serum was assessed by sandwich ELISA using a Synergy™ 2 microplate reader (BioTek, Winooski, VT). D-dimer in plasma was assessed by immunoturbidity using a BCS® coagulation analyzer (Siemens, Munich, Germany). Soluble vascular cell adhesion molecule-1 in serum was assessed by sandwich ELISA using a CiraScan™ microplate reader (Aushon Biosystems, Billerica, MA). Calibration curves and quality-control processes were implemented by the central laboratory for the various biomarker assays to ensure data integrity and eliminate any possible batch effects or inter-assay variability.