




Review

# Effect of Life-History Traits and Habitat Condition on Genetic Diversity between Invasive and Native Plant Populations

Raquel Hernández-Espinosa <sup>1,\*</sup>, Jorge González-Astorga <sup>1,\*</sup> , Yessica Rico <sup>2</sup>  and Juan B. Gallego-Fernández <sup>3</sup> 

<sup>1</sup> Red de Biología Evolutiva, Instituto de Ecología A. C. Carretera Antigua a Coatepec, No. 351, El Haya, Xalapa 91073, Mexico

<sup>2</sup> Red de Diversidad Biológica del Occidente Mexicano, Instituto de Ecología A. C. Centro Regional del Bajío, Pátzcuaro 61600, Mexico

<sup>3</sup> Departamento de Biología Vegetal y Ecología, Universidad de Sevilla, 41012 Sevilla, Spain

\* Correspondence: raquel.hernandez@posgrado.ecologia.edu.mx (R.H.-E.); jorge.gonzalez@inecol.mx (J.G.-A.)

**Abstract:** Plant invasions have a huge impact on the health of ecosystems and human well-being. The invasion risk varies with the introduction pathway, the propagule pressure, and the genetic diversity of the founding population. We performed a systematic review and meta-analysis of 30 studies reporting the genetic diversity of 31 plant species in their invasive and native ranges. We evaluated if patterns of genetic diversity differ between ranges and whether these responses are influenced by life-history traits, hybridization, polyploidization, and habitat condition. We found that invasive populations had significantly lower genetic diversity and higher inbreeding than native populations. In fragmented and degraded habitats, the genetic diversity of invaders was lower, but inbreeding was not affected. Polyploid invaders with hybrid capacity also showed lower genetic diversity. Invasive herbs with vegetative propagation were more sensitive to the loss of genetic diversity and had higher levels of inbreeding. Our synthesis showed that the genetic response in the invaded range could result from historical processes, such as founder and bottleneck events. Traits such as selfing are more likely to preserve the signatures of founder events and influence the genetic diversity in invasive populations. Additionally, clonality seems to be the predominant reproduction system in the invaded range.

**Keywords:** meta-analysis; biological invasions; genetic diversity; microsatellites; life-history traits



**Citation:** Hernández-Espinosa, R.; González-Astorga, J.; Rico, Y.; Gallego-Fernández, J.B. Effect of Life-History Traits and Habitat Condition on Genetic Diversity between Invasive and Native Plant Populations. *Diversity* **2022**, *14*, 1025. <https://doi.org/10.3390/d14121025>

Academic Editor: Hong-Hu Meng

Received: 28 October 2022

Accepted: 21 November 2022

Published: 24 November 2022

**Publisher's Note:** MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



**Copyright:** © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

## 1. Introduction

One of the biggest threats to global biodiversity is biological invasions [1]. Several species of plants have become invasive in their non-native habitats due to human influence on the natural world [2]. Invasion begins with the transportation of plant propagules beyond their native range limits, their introduction into a new environment, and their establishment and spread [3]. However, only a fraction of the species introduced into a new habitat will become invasive due to the high probability of mortality at some stage of the invasion process [4]. The invasion risk also depends on the introduction pathway (i.e., intentionally or unintentionally introduced) [5], propagule pressure (i.e., the number of individuals introduced) [6], and genetic diversity of the founding population [7]. As for the last, it has been reported that small, founding, and expanding populations are more likely to lose genetic diversity by genetic drift and demographic bottlenecks [8,9]. However, while some species showed lower genetic diversity in the introduced range compared to their native range [10–15], others showed an opposite pattern, partially because of multiple introductions or the admixture of independent genetic stocks [16–20].

Plant life-history traits can influence levels of genetic diversity in the invasive range and can be crucial to the success or failure of an invasion [10,21,22]. Reproductive systems are the main determinant of plant genetic diversity [23]. For instance, there is evidence that genetic diversity may be reduced in invaded ranges because of a change in reproductive

strategies (e.g., clonality and selfing) [22]. Another factor determining genetic diversity in plants is the life form, with trees usually having traits that maintain high levels of genetic diversity compared to herbaceous plants [24]. Likewise, trees predominantly outcross, with a lower proportion of selfing than other life forms [25]. However, self-compatible trees sometimes exhibit high inbreeding, even if mixed, outcrossing, or predominantly selfing [26]. Low levels of inbreeding are also associated with perennial plants due to their longevity and overlapping generations [27]. In addition to life-history traits, interspecific hybridization and polyploidization (genome duplication) can influence genetic diversity [28,29]. Specifically, within a few generations, interspecific hybridization with native congeners may result in higher levels of standing genetic variation in the exotic population, thereby increasing its adaptive capacity in the new environment [19,30]. Additionally, polyploidization, which increases genome size, can prevent genetic erosion during the early stages of an invasion because polyploid genomes are less susceptible to drift [29,31].

Successful establishment in the invasive range might also depend on external factors, such as the interaction between the invader and the new environment [32]. Evidence suggests that human-disturbed habitats facilitate plant invasions [33]. Human-disturbed habitats can influence the distribution of genetic diversity by influencing propagule dispersal and reproduction [34–38]. For disturbed habitats, changes in habitat structure, resource availability, and community composition may reduce the diversity of pollinators and seed dispersers, resulting in changes in reproductive strategies (e.g., selfing and vegetative propagation), which, in turn, can reduce genetic diversity [39,40].

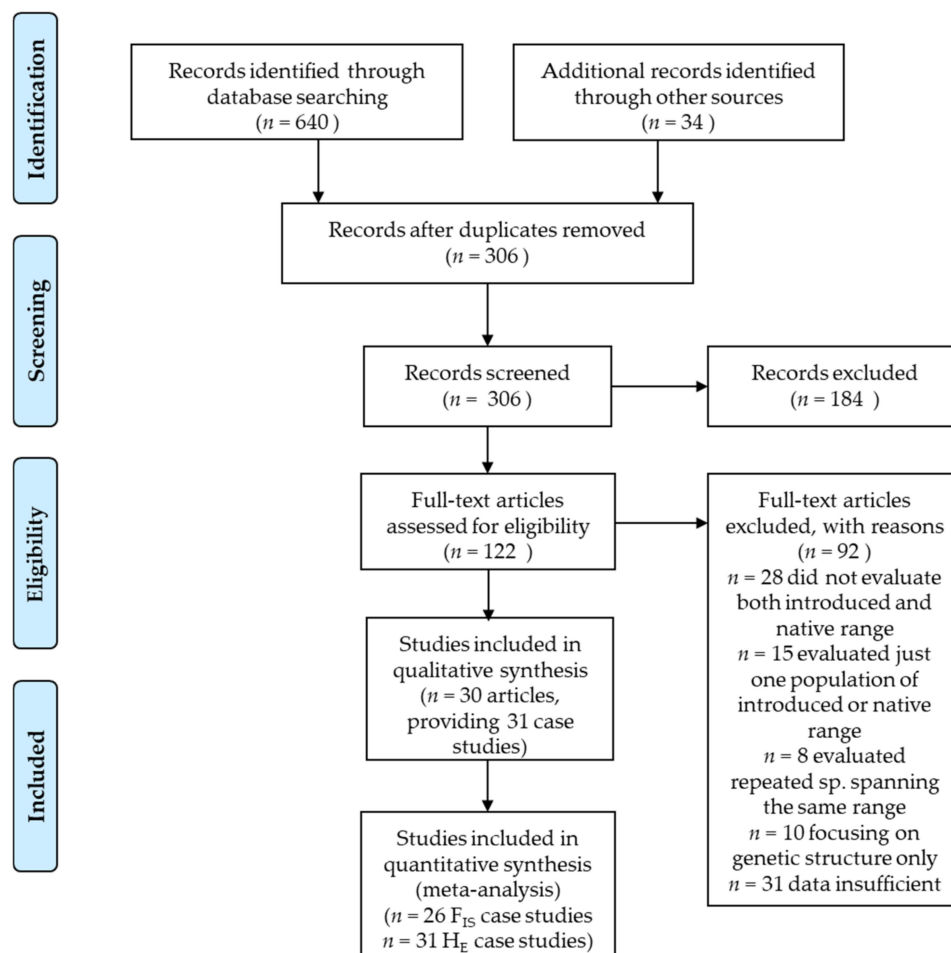
Systematic reviews comparing plant species in their invasive and native ranges have addressed the influence of fecundity-related traits on plant performance [41,42]. Meanwhile, meta-analytical approaches have evaluated performance-related traits, e.g., phenotypic plasticity, physiology, leaf-area allocation, shoot allocation, size, growth rate and fitness [43,44], life-history traits, and growth environment in species invasiveness [45]. So far, no previous studies have investigated the role of life-history traits, hybrid capacity, ploidy level, and habitat fragmentation and degradation in shaping genetic diversity in invaded ranges. In this study, we performed a systematic review and meta-analysis by comparing invasive and native ranges from empirical genetic studies in invasive plants to ask the following: (i) Is the genetic diversity of invasive populations lower relative to native populations? (ii) Are inbreeding levels higher in invasive populations than in native ones? (iii) Is genetic diversity and inbreeding within populations influenced by life-history traits in the invasive range? (iv) Does habitat fragmentation and degradation influence the invader's genetic diversity and inbreeding in the invasive range?

## 2. Materials and Methods

### 2.1. Study Selection and Coding

We searched for available scientific literature on invasive plant genetic diversity using a combination of the following keywords: *invas \* plants OR alien plants OR exotic plants OR introduced plants AND genetic diversity OR population genetic structure AND SSR OR microsatellite*, in Web of Science, Scopus, Google Scholar, JSTOR, Wiley, and Science Direct databases. We chose studies that used microsatellite markers because they have been the most popular markers in the last decades in population genetic studies and to avoid marker bias by including other types of genetic markers [46]. The search was conducted from 1980 to 2021 and followed the PRISMA statement [47], which provides a standardized framework for meta-analysis and systematic reviews (Figure 1). To ensure that the gathered information was of high quality and that our results were replicable, we limited our search to peer-reviewed articles written in English. We excluded grey literature and previous reviews (which may duplicate the information). More than one study focused on these three species: *Ambrosia artemisiifolia* L. (Asteraceae), *Spartina alterniflora* Loisel. (Poaceae), and *Acacia dealbata* A. Cunn. (Fabaceae) and because the study sites were the same, we

included only the articles reporting all information needed to fill out our inclusion criteria. Each evaluated species was considered a case study.



**Figure 1.** Flowchart of preferred reporting items for systematic reviews and meta-analysis (PRISMA), summarizing the sequence of information selected for this meta-analysis. *n* refers to the number of case studies used in each section.

We selected articles based on the following criteria: (i) included at least two invasive populations and two native populations of the same species; (ii) reported genetic diversity parameters of expected heterozygosity ( $H_E$ ) [48], allelic richness ( $A_r$ ), and inbreeding coefficients ( $F_{IS}$ ) [49] for at least five microsatellite markers (with their dispersion measure (SD or SE), or provide original data to calculate them); (iii) reported the sample size (as the proportion of individuals to the number of populations following González et al. (2020) [50]; and (iv) reported if the habitat was conserved, fragmented, or degraded (hereafter habitat condition, see below).

The information recorded for each species included: (i) species ID, (ii) a reference number for each article, (iii) population type (invasive or native), (iv) number of populations per type, (v) number of microsatellite loci, (vi) distribution range, (vii) family and genus. We categorized habitat conditions into three human habitat modification categories following the definitions of Lindenmayer and Fischer (2013) [51]: (i) undisturbed, (ii) degraded habitat, and (iii) fragmented habitat (Table 1). Invasiveness was determined according to the Invasive Species Compendium (CABI) [52] (Table S1). Information on life-history traits was obtained from the article when available. Otherwise, the information was extracted from peer-reviewed descriptions for the evaluated species: (i) life form, (ii) lifespan, (iii) reproductive system, (iv) vegetative propagation, (v) ploidy level, and (vi) hybrid capacity.

**Table 1.** Habitat condition categories.

Category	Description
Habitat fragmentation	Continuous areas subdivided into smaller fragments that increase habitat isolation and edge effects, usually accompanied by habitat loss.
Habitat degradation	Areas with a reduction in habitat quality due to changes in vegetation structure, resource availability, and microclimate conditions but without experiencing fragmentation or loss.
Undisturbed habitat	Areas unaltered by humans.

## 2.2. Effect Size and Moderators

All the evaluated studies included invasive and native populations; thus, we compared the effect of the moderators on genetic diversity and inbreeding for both distribution ranges. We calculated Hedge's *d* as the unbiased, standardized mean difference in genetic diversity between the invasive (treatment) and native (control) groups using the population as the unit of analysis [53,54]. Hedge's *d* compares the two groups' means, standard deviations, and sample sizes [55]. We used a correction for small samples (five to ten studies) [56] and conducted separate analyses for the genetic diversity and inbreeding data sets. We defined eight moderators: (i) habitat condition, (ii) invasiveness (highly invasive or moderate invasive), (iii) hybrid capacity (yes or no), (iv) vegetative propagation (yes or no), (v) ploidy level (diploid or polyploid), (vi) mating system (self-compatible or self-incompatible), (vii) lifespan (perennial or annual), and (viii) life form (herbs, shrubs, or trees).

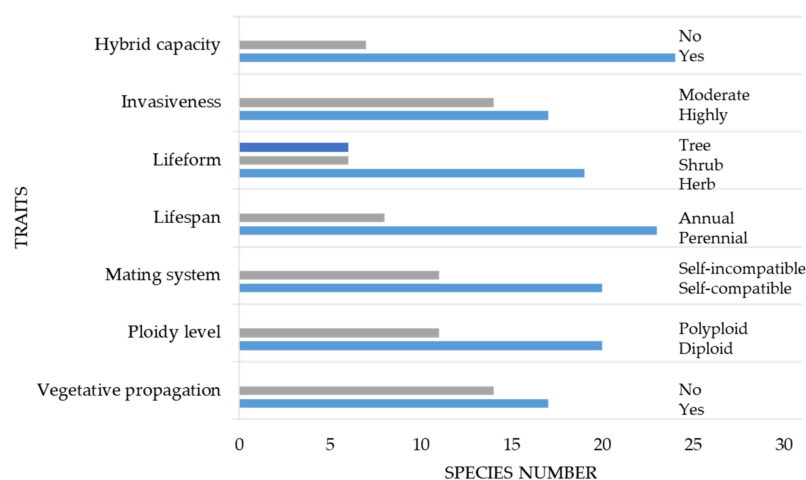
## 2.3. Effect Model and Publication Bias

The global effect sizes were estimated using mixed-effect models, considering each category in a moderator as a fixed subgroup and the variation between studies within categories as random [55]. To examine the heterogeneity among moderator levels, we estimated the between-group heterogeneity with the *Q*-between statistic and the  $\chi^2$ -distributed statistic, which compare the variation between and within moderators' levels and indicates a significant effect variation among moderator levels. *Q*-statistics are more appropriate for random-effect models than  $I^2$  or  $r^2$ , which assume a fixed-effects model structure [57]. We examined the relationship between the effect and sample size with funnel plots to assess the potential publication bias [57]. We used Egger's test to assess funnel-plot asymmetry [58]. We also conducted a trim-and-fill analysis, which recalculated the mean effect and confidence intervals to verify the robustness of the results [59]. All analyses were conducted using Comprehensive Meta-Analysis 3.0 software (Borenstein et al., 2005) [60].

## 3. Results

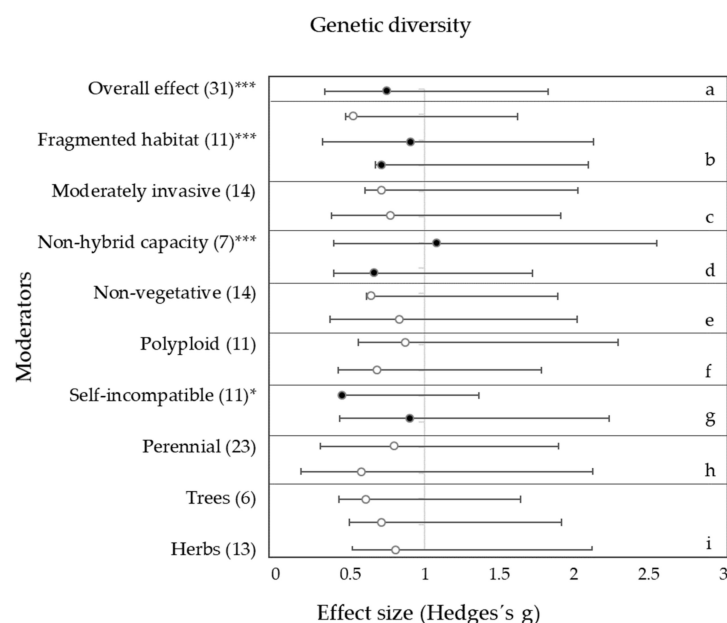
An initial search resulted in 674 articles, and after filtering out duplicates and irrelevant results, the total retained was 122. We excluded 92 articles that did not meet our inclusion criteria (28 did not evaluate both distribution ranges, 15 evaluated just one single population of each range, 8 evaluated the same species within the same range, 10 were focused on genetic structure only, and 31 lacked the data ( $H_E$ /Ar/ $F_{IS}$  or SD) to estimate diversity and inbreeding parameters, resulting in 30 articles that provided 31 case studies (Table S2). The 31 species evaluated were from 17 families and 29 genera. The family with the largest number of species was Asteraceae ( $n = 9$ ), followed by Fabaceae ( $n = 4$ ), Apiaceae ( $n = 4$ ), and then Rosaceae and Euphorbiaceae, with two species each. The remaining families had one representative. Only the genera *Senecio* L. (Asteraceae) and *Heracleum* L. (Apiaceae) had two species each. The number of populations included ranged from 4 to 50 in the invasive range and from 5 to 42 in the native range. The average number of individuals per population was 14, and the number of microsatellite loci ranged from 5 to 29. The dataset consisted of 19 herbs, 6 shrubs, 6 trees, 20 diploids, and 11 polyploids. Additionally, it

included 8 annual and 23 perennials, 20 self-compatible and 11 self-incompatible plants (Figure 2).

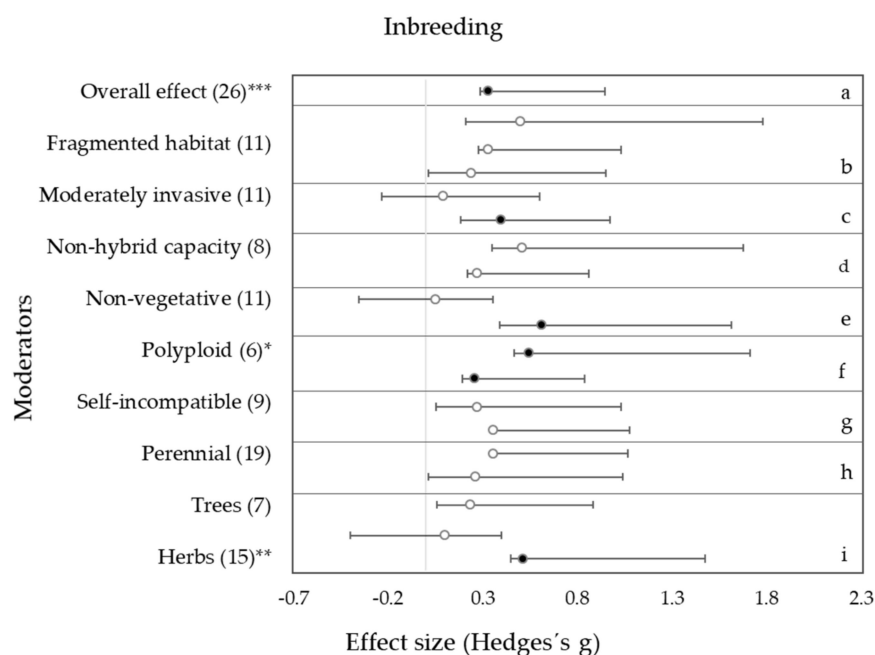


**Figure 2.** Plant species analyzed in the meta-analysis. The X-axis shows the number of species, and the Y-axis shows each of the seven moderators used. The bars show the categories within each moderator.

We could not include allelic richness (*Ar*) in the meta-analysis because the number of studies reporting it was insufficient to pass the publication bias tests (Figure S3). Thus, we only analyzed expected heterozygosity (hereafter genetic diversity), which provides an objective measurement of allelic richness and evenness in plants [50], for 31 case studies, whereas the inbreeding coefficient was obtained for 26 cases (hereafter inbreeding). Overall, the mixed effect models showed that invasive populations had a significantly lower genetic diversity ( $Z = -4.49, p = 0.0001$ ) and higher inbreeding ( $Z = -2.24, p = 0.025$ ) than the native populations. We found significant heterogeneity between study effect sizes for genetic diversity ( $QT = 179.356, df = 30, p = 0.0001$ ) (Figure 3a) and inbreeding ( $QT = 100.56, df = 25, p = 0.0001$ ) (Figure 4a).



**Figure 3.** Effect sizes on genetic diversity: (a) overall effect, (b) habitat condition, (c) invasiveness, (d) hybrid capacity, (e) vegetative propagation, (f) ploidy level, (g) mating systems, (h) lifespan, (i) life form (mean and 95% CI; numbers in parentheses represent sample size; \*  $p < 0.05$ , \*\*\*  $p < 0.001$ ; not significant,  $p \geq 0.05$ ). Black dots represent a significant effect on genetic diversity.



**Figure 4.** Effect sizes on inbreeding: (a) overall effect, (b) habitat condition, (c) invasiveness, (d) hybrid capacity, (e) vegetative propagation, (f) ploidy level, (g) mating systems, (h) lifespan, (i) life form (mean and 95% CI; numbers in parentheses represent sample size; \*  $p < 0.05$ , \*\*  $p < 0.01$ , \*\*\*  $p < 0.001$ ; not significant,  $p \geq 0.05$ ). Black dots represent a significant effect on inbreeding.

### 3.1. Habitat Condition

Habitat condition had a significant negative effect on genetic diversity ( $Z = -6.131$ ,  $p = 0.0001$ ) and inbreeding ( $Z = -2.271$ ,  $p = 0.023$ ) in the invasive populations. Specifically, habitat fragmentation and habitat degradation had statistically significant negative effects on the genetic diversity of invasive populations ( $Q$ -between = 6.345,  $df = 2$ ,  $p = 0.042$ ) (Figure 3b), whereas no effect was found on inbreeding ( $Q$ -between = 0.309,  $df = 2$ ,  $p = 0.857$ ) (Figure 4b).

### 3.2. Invasiveness

The overall effect size showed that species invasiveness significantly influences genetic diversity ( $Z = -4.370$ ,  $p = 0.0001$ ) and inbreeding ( $Z = -2.240$ ,  $p = 0.025$ ). Species recorded as highly invasive had no significant effect on genetic diversity ( $Q$ -between = 0.008,  $df = 1$ ,  $p = 0.930$ ) (Figure 3c), but did have an effect on inbreeding ( $Q$ -between = 3.838,  $df = 1$ ,  $p = 0.050$ ) (Figure 4c).

### 3.3. Hybrid Capacity and Ploidy Level

When considering hybrid capacity, both genetic diversity ( $Z = -4.465$ ,  $p = 0.0001$ ) and inbreeding ( $Z = -2.141$ ,  $p = 0.032$ ) were negatively affected in invasive populations. The genetic diversity of invasive populations was negatively affected by both hybrid and non-hybrid capacity ( $Q$ -between = 8.72,  $df = 1$ ,  $p = 0.003$ ) (Figure 3d), but not inbreeding ( $Q$ -between = 0.37,  $df = 1$ ,  $p = 0.539$ ) (Figure 4d). When we evaluated ploidy level, genetic diversity ( $Z = -4.393$ ,  $p = 0.0001$ ) was lower and inbreeding higher ( $Z = -2.252$ ,  $p = 0.024$ ) in invasive populations. No ploidy-level categories showed a significant effect on genetic diversity ( $Q$ -between = 2.34,  $df = 1$ ,  $p = 0.126$ ) (Figure 3f), but both diploids and polyploid species showed high inbreeding in invasive populations ( $Q$ -between = 2.453,  $df = 1$ ,  $p = 0.020$ ) (Figure 4f).



### 3.4. Life-History Traits

The mean effect sizes for life-history traits (except vegetative propagation), showed significant effects on genetic diversity and inbreeding. Based on vegetative propagation, genetic diversity of invasive populations was negatively affected ( $Z = -4.388$ ,  $p = 0.0001$ ), but the direct comparison between categories did not reveal significant differences ( $Q$ -between = 1.708,  $df = 1$ ,  $p = 0.191$ ) (Figure 3e). Moreover, vegetative propagation had no effect on inbreeding ( $Z = -1.845$ ,  $p = 0.065$ ). However, the direct comparison between inbreeding categories revealed that the “YES” category had a significant effect ( $Q$ -between = 14.22,  $df = 1$ ,  $p = 0.0001$ ) (Figure 4e). For mating systems, genetic diversity of invasive populations was significantly lower for self-compatible and self-incompatible plants ( $Q$ -between = 13.87,  $df = 1$ ,  $p = 0.0001$ ) (Figure 3g), but inbreeding was not influenced by mating system ( $Q$ -between = 2.21,  $df = 1$ ,  $p = 0.137$ ) (Figure 4g). Regarding lifespan, perennial plants of invasive populations showed significant lower values of genetic diversity ( $Z = -4.078$ ,  $p = 0.0001$ ) (Figure 3h) and inbreeding ( $Z = -1.980$ ,  $p = 0.048$ ) (Figure 4h). For life form, genetic diversity ( $Z = -4.289$ ,  $p = 0.0001$ ) and inbreeding ( $Z = -2.239$ ,  $p = 0.025$ ) showed lower values in the invasive populations. However, a direct comparison between life forms did not reveal significant differences for genetic diversity ( $Q$ -between = 0.332,  $df = 2$ ,  $p = 0.847$ ) (Figure 3i), but did for inbreeding. Inbreeding was significantly higher in invasive herbs ( $Q$ -between = 8.845,  $df = 2$ ,  $p = 0.012$ ) (Figure 4i).

### 3.5. Publication Bias

Funnel plots showed some cases with large variances within the total distribution for genetic diversity. However, Egger’s test showed no significant asymmetry (intercept =  $-2.68$ ,  $df = 29$ ,  $p = 0.181$ ). Similarly, for inbreeding, the funnel plot showed some study cases with a large variation, but Egger’s test showed no significant asymmetry (intercept =  $-2.20$ ,  $df = 24$ ,  $p = 0.211$ ). The direction and significance of inbreeding results did not change after the trim-and-fill procedure, indicating that they were robust and uninfluenced by asymmetric bias. For genetic diversity, the direction did not change but the effect size estimated for the trim-and-fill procedure diminished (Figure S3).

## 4. Discussion

Population genetic studies on invasive plants have aimed to determine which factors (i.e., mating systems, life form, habitat condition) play a significant role in species’ genetic diversity in the invaded range. However, there was no evidence of a general pattern. Our meta-analytical approach is the first to identify with statistical robustness the intrinsic (life-history traits) and extrinsic factors (human habitat modification) that influence the genetic response of invasive and native plant populations. Our global synthesis showed that invaders have lower genetic diversity and higher inbreeding relative to native populations. Empirical studies on invasive plants have reported low genetic diversity in the introduced range when populations experience genetic and demographic bottlenecks or colonization events [36,61]. Our results confirm that life-history traits, such as self-compatibility, vegetative propagation, and herbaceous life form, can significantly influence invaders’ genetic diversity and inbreeding. For invasive species, historical processes are important determinants of genetic diversity, such as founder events and frequent genetic bottlenecks. Traits such as selfing are more likely to preserve the signatures of founder events in invasive populations [62]. Likewise, we confirmed that habitat condition is a stronger driver of invasive plant genetic diversity. Both fragmented and degraded habitats had a negative effect on invaders’ genetic diversity.

### 4.1. Effects of Habitat Condition on Genetic Diversity and Inbreeding

We found a significant negative effect of fragmented and degraded habitats on invaders’ genetic diversity, but not on inbreeding. Human-modified habitats are more prone to invasion than natural or semi-natural areas; the flora of urban areas and surroundings is usually richer in invasive species than natural areas [63]. Fragmented habitats are associ-

ated with a reduction in genetic diversity by a decrease in population size due to habitat isolation and matrix impermeability [64]. Fragmentation is also associated with higher selfing rates [50,65–67], which is consistent with our findings. We detected a significant reduction in invaders' genetic diversity in self-compatible species. We also found a significant negative effect of habitat degradation on invaders' genetic diversity. When an exotic species partially or totally replaces the native vegetation in the new area, the resulting habitat is usually composed of a mosaic of native and transformed stands [35]. Degraded habitats lack spatial discontinuities, but may impose restrictions to dispersal and gene flow [68]. The interruption of gene flow in altered habitats can affect invaders' genetic diversity [69] by changes in reproductive strategies, such as selfing or clonality, facilitating propagation. Habitat degradation also leads to changes in vegetation structure, source availability, and microclimate conditions [70], which can alter the diversity and behavior of pollinators and seed dispersers.

#### 4.2. Effects of Interspecific Hybridization and Polyploidization on Genetic Diversity and Inbreeding

The formation of polyploids may be affected by environmental stress stimuli, as seen during an invasion. We found significantly low genetic diversity in polyploid invaders with hybrid capacity. Even if genome duplication may increase genetic variability, clonal propagation is often the strategy to spread, leading to extensive areas of genetic uniformity in the novel range [71]. Additionally, such invaders as *Tragopogon* L. (Asteraceae) [72] and *Spartina* L. (Poaceae) [73] have shown a marked reduction in the expression of duplicated genes with similar or redundant functions, which has led to genetic diversity loss in response to "genomic shock" by genomic incompatibilities and genetic redundancy during a polyploidization event [74,75]. Our results did not show a significant relationship between the mating system and inbreeding, suggesting that selfing does not favor inbreeding.

#### 4.3. Effects of Life-History Traits on Genetic Diversity and Inbreeding

We found that genetic diversity and inbreeding were associated with specific plant traits in the invasive populations. For mating systems, we found a significantly low genetic diversity in self-compatible invasive populations. Selfing ability could promote invasiveness because it ensures reproduction even in the absence of pollinators [76]. Nevertheless, selfing can be harmful to reproductive success when inbreeding depression is present [26]. When we evaluated life forms, we found that herbs with vegetative propagation (clonality) had a negative effect on inbreeding. This result confirms what is expected when the reproductive system is predominantly vegetative in an invasion scenery. Clonal invasions in which sexual reproduction is limited or absent involve striking bottlenecks [22,28]. In many instances, invasion processes cause transitions to uniparental reproduction in the introduced range [23]. Although vegetative propagation appears to have ecological advantages in novel environments [23], extensive clonality can promote complete loss of sexual reproduction, increasing inbreeding in the populations [28].

Genetic studies, including the native range of an invader's species, have shown that selfing and inbreeding depression varied between life forms [26]. Herbaceous plants are frequently associated with early successional series in disturbed areas, where selfing is probably selected as a colonizing strategy when pollinators are scarce or inefficient [77]. Here, we found significantly higher inbreeding in invasive self-compatible herbs. Other studies evaluating selfing in herbaceous species have found that inbreeding may occur under various conditions, more frequently for taxa growing in stressful environments, as expected for invasive species [77–79]. Additionally, high levels of inbreeding depression are reported in tree invasions that are generally self-incompatible [80], although long-distance dispersal can ensure that some selfed progeny escape competition with outcrossed progeny [81,82]. In our meta-analysis, we did not find a strong relationship between trees and inbreeding, but we found a strong effect of self-compatibility on genetic diversity. Thus, despite high levels of inbreeding reported in tree invasions, there is a possibility that selfing promotes invasion or natural long-distance colonization. Invasive plants experienced



reductions in genetic diversity and an increase in inbreeding resulting from founder effects, genetic bottlenecks, autogamy, and reduced gene flow among isolated populations owing to habitat fragmentation. Such a combination of factors may reduce the invader's fitness due to the fixation of mildly deleterious recessive alleles, indicating potential genetic weakness, as reported in *Alliaria petiolata* [83]. Like *A. petiolata*, other invasive species may restore genetic variation by multiple introductions and genetic exchange among locally isolated populations. Still, once again, the strength of processes like this is highly dependent on the invasion history of species.

## 5. Future Research Recommendations

Despite the increasing number of genetic studies that include populations in the native and invasive range, few include all the necessary information to conduct a meta-analysis. For instance, parameters such as  $H_E$  and  $SD$  and  $F_{IS}$  and  $SD$  were lacking in 31 articles, whereas allelic richness ( $A_r$ ) was reported only in a small number of studies.  $A_r$  is highly affected by demographic changes, being more sensitive to detect founder events. In a meta-analysis, the sample size is another constraint. We excluded 15 articles investigating only one invasive or native population and 8 that used less than five microsatellites, thereby drastically reducing our sample size.

Moreover, we found a lack of representativity of plant families, life forms, and lifespans. Perennial herbs were the most frequent, doubling the other categories. Increasing the number of empirical studies for diverse plant species is thus needed. Lastly, we evaluated studies based on microsatellites, which are codominant neutral markers, and, thereby, cannot directly inform selection and local adaptation. A future meta-analysis may consider including empirical studies based on functional genomic data, such as genome-wide association studies (GWAS) or next-generation sequencing (NGS). This information would shed light on traits under selection that influence the genetic response during a plant invasion.

**Supplementary Materials:** The following supporting information can be downloaded at the following: <https://www.mdpi.com/article/10.3390/d14121025/s1>, Table S1: Invasiveness categories; Table S2: Species evaluated in the meta-analysis; Figure S1: Publication bias results; Table S3: Duval & Tweedie's trim and fill procedure on genetic diversity. References [84–104] are cited in the Table S2.

**Author Contributions:** R.H.-E. and J.G.-A. conceived and designed the study. R.H.-E. conducted the formal analysis and wrote the original draft; J.G.-A., Y.R. and J.B.G.-F. conducted the review and editing. All authors have read and agreed to the published version of the manuscript.

**Funding:** This study was supported by the Consejo Nacional de Ciencia y Tecnología de México CONACYT by a scholarship (2019-000037-02NACF-29365) awarded to R.H.-E., as part of her doctoral thesis directed by J.G.-A. This study was also supported by the Ministerio de Economía y Competitividad, Spain (MINECO Project CGL2015-65058-R co-funded by FEDER).

**Institutional Review Board Statement:** Not applicable.

**Data Availability Statement:** Not applicable.

**Conflicts of Interest:** The authors declare no conflict of interest. The funders had no role in the design of the study; in the collection, analyses, or interpretation of data; in the writing of the manuscript, or in the decision to publish the results.

## References

1. Paini, D.R.; Sheppard, A.W.; Cook, D.C.; De Barro, P.J.; Worner, S.P.; Thomas, M.B. Global threat to agriculture from invasive species. *Proc. Natl. Acad. Sci. USA* **2016**, *113*, 7575–7579. [[CrossRef](#)] [[PubMed](#)]
2. Castro-Díez, P.; Godoy, O.; Alonso, A.; Gallardo, A.; Saldaña, A. What explains variation in the impacts of exotic plant invasions on the nitrogen cycle? A meta-analysis. *Ecol. Lett.* **2014**, *17*, 1–12. [[CrossRef](#)] [[PubMed](#)]
3. Blackburn, T.M.; Pyšek, P.; Bacher, S.; Carlton, J.T.; Duncan, R.P.; Jarošík, V.; Wilson, J.R.U.; Richardson, D.M. A proposed unified framework for biological invasions. *Trends Ecol. Evol.* **2011**, *26*, 333–339. [[CrossRef](#)] [[PubMed](#)]
4. Young, S.L.; Kimball, S.; Novak, S.J. Invasion of Plant Communities. In *Global Plant Invasions*; 2022; pp. 29–51, ISBN 9783030896843.
5. Riera, M.; Pino, J.; Melero, Y. Impact of introduction pathways on the spread and geographical distribution of alien species: Implications for preventive management in Mediterranean ecosystems. *Divers. Distrib.* **2021**, *27*, 1019–1034. [[CrossRef](#)]

6. Blackburn, T.M.; Lockwood, J.L.; Cassey, P. The influence of numbers on invasion success. *Mol. Ecol.* **2015**, *24*, 1942–1953. [[CrossRef](#)]
7. Estoup, A.; Ravigné, V.; Hufbauer, R.; Vitalis, R.; Gautier, M.; Facon, B. Is There a Genetic Paradox of Biological Invasion? *Annu. Rev. Ecol. Evol. Syst.* **2016**, *47*, 51–72. [[CrossRef](#)]
8. Dlugosch, K.M.; Parker, I.M. Founding events in species invasions: Genetic variation, adaptive evolution, and the role of multiple introductions. *Mol. Ecol.* **2008**, *17*, 431–449. [[CrossRef](#)]
9. Allendorf, F.W.; Lundquist, L.L. Society for Conservation Biology Introduction: Population Biology, Evolution, and Control of Invasive Species. *Biology* **2003**, *17*, 24–30.
10. Amsellem, L.; Noyer, J.L.; Hossaert-Mckey, M. Evidence for a switch in the reproductive biology of *Rubus alceifolius* (Rosaceae) towards apomixis, between its native range and its area of introduction. *Am. J. Bot.* **2001**, *88*, 2243–2251. [[CrossRef](#)]
11. Husband, B.C.; Barrett, S.C.H. Colonization history and population genetic structure of *Eichornia paniculata* in Jamaica. *Heredity* **1991**, *66*, 287–296. [[CrossRef](#)]
12. Frankham, R. Resolving the genetic paradox in invasive species. *Heredity* **2005**, *94*, 385. [[CrossRef](#)]
13. Zimmermann, H.; Ritz, C.M.; Hirsch, H.; Renison, D.; Wesche, K.; Hensen, I. Highly Reduced Genetic Diversity of *Rosa rubiginosa* L. Populations in the Invasive Range. *Int. J. Plant Sci.* **2010**, *171*, 435–446. [[CrossRef](#)]
14. Castillo, J.M.; Gallego-Tévar, B.; Figueroa, E.; Grewell, B.J.; Vallet, D.; Rousseau, H.; Keller, J.; Lima, O.; Dréano, S.; Salmon, A.; et al. Low genetic diversity contrasts with high phenotypic variability in heptaploid *Spartina densiflora* populations invading the Pacific coast of North America. *Ecol. Evol.* **2018**, *8*, 4992–5007. [[CrossRef](#)]
15. Williams, D.; Harms, N.; Grodowitz, M.; Purcell, M. Genetic structure of *Hydrilla verticillata* L.f. Royle in eastern China and the Republic of Korea: Implications for surveys of biological control agents for the invasive monoecious biotype. *Aquat. Bot.* **2018**, *149*, 17–27. [[CrossRef](#)]
16. Bossdorf, O.; Auge, H.; Lafuma, L.; Rogers, W.E.; Siemann, E.; Prati, D. Phenotypic and genetic differentiation between native and introduced plant populations. *Oecologia* **2005**, *144*, 1–11. [[CrossRef](#)]
17. Rollins, L.A.; Moles, A.T.; Lam, S.; Buitenwerf, R.; Buswell, J.M.; Brandenburger, C.R.; Flores-Moreno, H.; Nielsen, K.B.; Couchman, E.; Brown, G.S.; et al. High genetic diversity is not essential for successful introduction. *Ecol. Evol.* **2013**, *3*, 4501–4517. [[CrossRef](#)]
18. Helsen, K.; Hagenblad, J.; Acharya, K.P.; Brunet, J.; Cousins, S.A.O.; Decocq, G.; De Frenne, P.; Kimberley, A.; Kolb, A.; Michaelis, J.; et al. No genetic erosion after five generations for *Impatiens glandulifera* populations across the invaded range in Europe. *BMC Genet.* **2019**, *20*, 20. [[CrossRef](#)]
19. Shang, L.; Li, L.-F.; Song, Z.-P.; Wang, Y.; Yang, J.; Wang, C.-C.; Qiu, S.-Y.; Huang, J.-X.; Nie, M.; Wolfe, L.M.; et al. High Genetic Diversity with Weak Phylogeographic Structure of the Invasive *Spartina alterniflora* (Poaceae) in China *Front. Plant Sci.* **2019**, *10*, 1467.
20. Li, F.; van Kleunen, M.; Li, J.; Liu, X.; Gao, K.; Zhu, J.; Zhao, X.; Zhao, C.; Li, J. Patterns of genetic variation reflect multiple introductions and pre-admixture sources of common ragweed (*Ambrosia artemisiifolia*) in China. *Biol. Invasions* **2019**, *21*, 2191–2209. [[CrossRef](#)]
21. Kirk, H.; Paul, J.; Straka, J.; Freeland, J.R. Long-distance dispersal and high genetic diversity are implicated in the invasive spread of the common reed, *Phragmites australis* (Poaceae), in northeastern North America. *Am. J. Bot.* **2011**, *98*, 1180–1190. [[CrossRef](#)]
22. Bentley, K.E.; Mauricio, R. High degree of clonal reproduction and lack of large-scale geographic patterning mark the introduced range of the invasive vine, kudzu (*Pueraria montana* var. *lobata*), in North America. *Am. J. Bot.* **2016**, *103*, 1499–1507. [[CrossRef](#)]
23. Barrett, S.C.H. Why reproductive systems matter for the invasion biology of plants. In *Fifty Years of Invasion Ecology: The Legacy of Charles Elton*; Richardson, D.M., Ed.; Wiley-Blackwell: Oxford, UK, 2011; pp. 195–210.
24. Chung, M.Y.; Son, S.; Herrando-Moraira, S.; Tang, C.Q.; Maki, M.; Kim, Y.-D.; López-Pujol, J.; Hamrick, J.L.; Chung, M.G. Incorporating differences between genetic diversity of trees and herbaceous plants in conservation strategies. *Conserv. Biol. J. Soc. Conserv. Biol.* **2020**, *34*, 1142–1151. [[CrossRef](#)]
25. Hamrick, J.; Godt, M. Effects of life history traits on genetic diversity in plant species. *Philos. Trans. R. Soc. Lond. Ser. B Biol. Sci.* **1996**, *351*, 1291–1298. [[CrossRef](#)]
26. Rodger, J.G.; Johnson, S.D. Self-pollination and inbreeding depression in *Acacia dealbata*: Can selfing promote invasion in trees? *S. Afr. J. Bot.* **2013**, *88*, 252–259. [[CrossRef](#)]
27. Rosche, C.; Hensen, I.; Schaar, A.; Zehra, U.; Jasieniuk, M.; Callaway, R.M.; Khasa, D.P.; Al-Gharaibeh, M.M.; Lekberg, Y.; Nagy, D.U.; et al. Climate outweighs native vs. nonnative range-effects for genetics and common garden performance of a cosmopolitan weed. *Ecol. Monogr.* **2019**, *89*, e01386. [[CrossRef](#)]
28. Ferrero, V.; Barrett, S.C.H.; Castro, S.; Caldeirinha, P.; Navarro, L.; Loureiro, J.; Rodríguez-Echeverría, S. Invasion genetics of the Bermuda buttercup (*Oxalis pes-caprae*): Complex intercontinental patterns of genetic diversity, polyploidy and heterostyly characterize both native and introduced populations. *Mol. Ecol.* **2015**, *24*, 2143–2155. [[CrossRef](#)] [[PubMed](#)]
29. Te Beest, M.; Le Roux, J.J.; Richardson, D.M.; Brysting, A.K.; Suda, J.; Kubesoová, M.; Pysek, P. The more the better? The role of polyploidy in facilitating plant invasions. *Ann. Bot.* **2012**, *109*, 19–45. [[CrossRef](#)]
30. Ellstrand, N.C.; Schierenbeck, K.A. Hybridization as a stimulus for the evolution of invasiveness in plants? *Euphytica* **2006**, *148*, 35–46. [[CrossRef](#)]

31. Rosche, C.; Durka, W.; Hensen, I.; Mráz, P.; Hartmann, M.; Müller-Schärer, H.; Lachmuth, S. The population genetics of the fundamental cytotype-shift in invasive *Centaurea stoebe* s.l.: Genetic diversity, genetic differentiation and small-scale genetic structure differ between cytotypes but not between ranges. *Biol. Invasions* **2016**, *18*, 1895–1910. [[CrossRef](#)]
32. Sans, F.X.; Garcia-Serrano, H.; Afán, I. Life-history traits of alien and native *Senecio* species in the Mediterranean region. *Acta Oecologica-Int. J. Ecol.* **2004**, *26*, 167–178. [[CrossRef](#)]
33. Lee, C.E.; Gelembiuk, G.W. Evolutionary origins of invasive populations. *Evol. Appl.* **2008**, *1*, 427–448. [[CrossRef](#)]
34. Williams, D.A.; Overholt, W.A.; Cuda, J.P.; Hughes, C.R. Chloroplast and microsatellite DNA diversities reveal the introduction history of Brazilian peppertree (*Schinus terebinthifolius*) in Florida. *Mol. Ecol.* **2005**, *14*, 3643–3656. [[CrossRef](#)]
35. Chabrierie, O.; Verheyen, K.; Saguez, R.; Decocq, G. Disentangling relationships between habitat conditions, disturbance history, plant diversity, and American black cherry (*Prunus serotina* Ehrh.) invasion in a European temperate forest. *Divers. Distrib.* **2008**, *14*, 204–212. [[CrossRef](#)]
36. Henry, P.; Le Lay, G.; Goudet, J.; Guisan, A.; Jahodová, S.; Besnard, G. Reduced genetic diversity, increased isolation and multiple introductions of invasive giant hogweed in the western Swiss Alps. *Mol. Ecol.* **2009**, *18*, 2819–2831. [[CrossRef](#)]
37. DeWalt, S.J.; Siemann, E.; Rogers, W.E. Geographic distribution of genetic variation among native and introduced populations of Chinese tallow tree, *Triadica sebifera* (Euphorbiaceae). *Am. J. Bot.* **2011**, *98*, 1128–1138. [[CrossRef](#)]
38. Hernández, F.; Presotto, A.; Poverene, M.; Mandel, J.R. Genetic Diversity and Population Structure of Wild Sunflower (*Helianthus annuus* L.) in Argentina: Reconstructing Its Invasion History. *J. Hered.* **2019**, *110*, 746–759. [[CrossRef](#)]
39. Banks, S.C.; Cary, G.J.; Smith, A.L.; Davies, I.D.; Driscoll, D.A.; Gill, A.M.; Lindenmayer, D.B.; Peakall, R. How does ecological disturbance influence genetic diversity? *Trends Ecol. Evol.* **2013**, *28*, 670–679. [[CrossRef](#)]
40. Devictor, V.; Robert, A. Measuring community responses to large-scale disturbance in conservation biogeography. *Divers. Distrib.* **2009**, *15*, 122–130. [[CrossRef](#)]
41. Daehler, C. Performance comparisons of co-occurring native and alien invasive plants: Implications for conservation and restoration. *Annu. Rev. Ecol. Syst.* **2003**, *34*, 183–211. [[CrossRef](#)]
42. Pysek, P.; Richardson, D. Traits associated with invasiveness in alien plants: Where do we stand? In *Biological Invasions*; Nentwig, W., Ed.; Springer: New York, NY, USA, 2007; pp. 97–125.
43. Van Kleunen, M.; Weber, E.; Fischer, M. A meta-analysis of trait differences between invasive and non-invasive plant species. *Ecol. Lett.* **2010**, *13*, 235–245. [[CrossRef](#)]
44. Palacio-López, K.; Gianoli, E. Invasive plants do not display greater phenotypic plasticity than their native or non-invasive counterparts: A meta-analysis. *Oikos* **2011**, *120*, 1393–1401. [[CrossRef](#)]
45. Oduor, A.M.O.; Leimu, R.; van Kleunen, M. Invasive plant species are locally adapted just as frequently and at least as strongly as native plant species. *J. Ecol.* **2016**, *104*, 957–968. [[CrossRef](#)]
46. Tarin, D.; Pepper, A.E.; Goolsby, J.A.; Moran, P.J.; Arqueta, A.C.; Kirk, A.E.; Manhart, J.R. Microsatellites Uncover Multiple Introductions of Clonal Giant Reed (*Arundo donax*). *Invasive Plant Sci. Manag.* **2013**, *6*, 328–338. [[CrossRef](#)]
47. Moher, D.; Liberati, A.; Tetzlaff, J.; Altman, D.G.; Altman, D.; Antes, G.; Atkins, D.; Barbour, V.; Barrowman, N.; Berlin, J.A.; et al. Preferred reporting items for systematic reviews and meta-analyses: The PRISMA statement. *PLoS Med.* **2009**, *6*, e1000097. [[CrossRef](#)] [[PubMed](#)]
48. Nei, M. Estimation of average heterozygosity and genetic distance from a small number of individuals. *Genetics* **1978**, *89*, 583–590. [[CrossRef](#)] [[PubMed](#)]
49. Wright, S. The Genetical Structure of Populations. *Ann. Hum. Genet.* **1950**, *15*, 323–354. [[CrossRef](#)]
50. González, A.V.; Gómez-Silva, V.; Ramírez, M.J.; Fontúrbel, F.E. Meta-analysis of the differential effects of habitat fragmentation and degradation on plant genetic diversity. *Conserv. Biol. J. Soc. Conserv. Biol.* **2020**, *34*, 711–720. [[CrossRef](#)]
51. Lindenmayer, D.B.; Fischer, J. *Habitat Fragmentation and Landscape Change: An Ecological and Conservation Synthesis*; Island Press: Washington, DC, USA, 2013; ISBN 159726606X.
52. CABI Invasive Species Compendium. Available online: [www.cabi.org/isc](http://www.cabi.org/isc) (accessed on 12 December 2021).
53. Bonett, D.G. Confidence intervals for standardized linear contrasts of means. *Psychol. Methods* **2008**, *13*, 99–109. [[CrossRef](#)]
54. Bonett, D.G. Meta-analytic interval estimation for standardized and unstandardized mean differences. *Psychol. Methods* **2009**, *14*, 225–238. [[CrossRef](#)]
55. Koricheva, J.; Gurevitch, J.; Mengersen, K. *Handbook of Meta-Analysis in Ecology and Evolution*; Koricheva, J., Gurevitch, J., Mengersen, K., Eds.; Princeton University Press: Princeton, NJ, USA, 2013; ISBN 9780691137292.
56. Hedges, L. V Distribution theory for Glass's estimator of effect size and related estimators. *J. Educ. Stat.* **1981**, *6*, 107–128. [[CrossRef](#)]
57. Borenstein, M.; Hedges, L.V.; Higgins, J.P.T.; Rothstein, H.R. *Introduction to Meta-Analysis*; John Wiley & Sons: Chichester, UK, 2009.
58. Egger, M.; Smith, G.D.; Minder, C. Bias in meta-analysis detected by a simple, graphical test. *Br. Med. J.* **1997**, *315*, 629–634. [[CrossRef](#)]
59. Jennions MD, M.A. Publication bias in ecology and evolution: An empirical assessment using the 'trim and fill' method. *Biol. Rev.* **2002**, *77*, 211–222. [[CrossRef](#)]
60. Borenstein, M.; Hedges, L.; Higgins, J.; Rothstein, H. *Comprehensive Meta-Analysis*; Biostat: Englewood, NJ, USA, 2005.

61. Alexander, J.M.; Poll, M.; Dietz, H.; Edwards, P.J. Contrasting patterns of genetic variation and structure in plant invasions of mountains. *Divers. Distrib.* **2009**, *15*, 502–512. [[CrossRef](#)]
62. Zhu, B.R.; Barrett, S.C.H.; Zhang, D.Y.; Liao, W.J. Invasion genetics of *Senecio vulgaris*: Loss of genetic diversity characterizes the invasion of a selfing annual, despite multiple introductions. *Biol. Invasions* **2017**, *19*, 255–267. [[CrossRef](#)]
63. Vilá, M.; Valladares, F.; Traveset, A.; Santamaría, L.; Castro, P. *Invasiones Biológicas*; Cyan, S.A., Ed.; Consejo Superior de Investigaciones Científicas: Madrid, Spain, 2008; ISBN 9788400085452.
64. Honnay, O.; Jacquemyn, H. Susceptibility of common and rare plant species to the genetic consequences of habitat fragmentation. *Conserv. Biol. J. Soc. Conserv. Biol.* **2007**, *21*, 823–831. [[CrossRef](#)]
65. Aguilar, R.; Quesada, M.; Ashworth, L.; Herrerias-Diego, Y.; Lobo, J. Genetic consequences of habitat fragmentation in plant populations: Susceptible signals in plant traits and methodological approaches. *Mol. Ecol.* **2008**, *17*, 5177–5188. [[CrossRef](#)]
66. Breed, M.F.; Ottewell, K.M.; Gardner, M.G.; Marklund, M.H.K.; Dormontt, E.E.; Lowe, A.J. Mating patterns and pollinator mobility are critical traits in forest fragmentation genetics. *Heredity* **2015**, *115*, 108–114. [[CrossRef](#)]
67. DiBattista, J.D. Patterns of genetic variation in anthropogenically impacted populations. *Conserv. Genet.* **2008**, *9*, 141–156. [[CrossRef](#)]
68. Fontúrbel, F.E.; Bruford, M.W.; Salazar, D.A.; Cortés-Miranda, J.; Vega-Retter, C. The hidden costs of living in a transformed habitat: Ecological and evolutionary consequences in a tripartite mutualistic system with a keystone mistletoe. *Sci. Total Environ.* **2019**, *651*, 2740–2748. [[CrossRef](#)]
69. González-Varo, J.P.; Carvalho, C.S.; Arroyo, J.M.; Jordano, P. Unravelling seed dispersal through fragmented landscapes: Frugivore species operate unevenly as mobile links. *Mol. Ecol.* **2017**, *26*, 4309–4321. [[CrossRef](#)]
70. Lu, S.; Luo, X.; Han, L.; Yang, J.; Jin, J.; Yang, J. Genetic patterns reveal differences between the invasion processes of common ragweed in urban and non-urban ecosystems. *Glob. Ecol. Conserv.* **2022**, *38*, e02214. [[CrossRef](#)]
71. García-Verdugo, C.; Calleja, J.A.; Vargas, P.; Silva, L.; Moreira, O.; Pulido, F. Polyploidy and microsatellite variation in the relict tree *Prunus lusitanica* L.: How effective are refugia in preserving genotypic diversity of clonal taxa? *Mol. Ecol.* **2013**, *22*, 1546–1557. [[CrossRef](#)] [[PubMed](#)]
72. Tate, J.A.; Ni, Z.; Scheen, A.-C.; Koh, J.; Gilbert, C.A.; Lefkowitz, D.; Chen, Z.J.; Soltis, P.S.; Soltis, D.E. Evolution and expression of homeologous loci in *Tragopogon miscellus* (Asteraceae), a recent and reciprocally formed allopolyploid. *Genetics* **2006**, *173*, 1599–1611. [[CrossRef](#)] [[PubMed](#)]
73. Salmon, A.; Ainouche, M.L.; Wendel, J.F. Genetic and epigenetic consequences of recent hybridization and polyploidy in *Spartina* (Poaceae). *Mol. Ecol.* **2005**, *14*, 1163–1175. [[CrossRef](#)] [[PubMed](#)]
74. Parisod, C.; Salmon, A.; Zerjal, T.; Tenaillon, M.; Grandbastien, M.-A.; Ainouche, M. Rapid structural and epigenetic reorganization near transposable elements in hybrid and allopolyploid genomes in *Spartina*. *New Phytol.* **2009**, *184*, 1003–1015. [[CrossRef](#)]
75. Chen, Z.J. Genetic and epigenetic mechanisms for gene expression and phenotypic variation in plant polyploids. *Annu. Rev. Plant Biol.* **2007**, *58*, 377–406. [[CrossRef](#)]
76. Baker, H.G. Characteristics and modes of origin of weeds. In *Characteristics and Modes of Origin of Weeds*; Academic Press Inc.: New York, NY, USA, 1965; pp. 147–172.
77. Levin, D.A. Mating system shifts on the trailing edge. *Ann. Bot.* **2012**, *109*, 613–620. [[CrossRef](#)]
78. Herrera, I.; Nassar, J.M. Reproductive and recruitment traits as indicators of the invasive potential of *Kalanchoe daigremontiana* (Crassulaceae) and *Stapelia gigantea* (Apocynaceae) in a Neotropical arid zone. *J. Arid Environ.* **2009**, *73*, 978–986. [[CrossRef](#)]
79. Lemus-Jiménez, L.J.; Ramírez, N. Breeding systems of the Paraguaná coastal plain plants, Venezuela. *Rev. Biol. Trop.* **2005**, *53*, 415–430. [[CrossRef](#)]
80. Duminil, J.; Hardy, O.J.; Petit, R.J. Plant traits correlated with generation time directly affect inbreeding depression and mating system and indirectly genetic structure. *BMC Evol. Biol.* **2009**, *9*, 177. [[CrossRef](#)]
81. Dick, C.W.; Bermingham, E.; Lemes, M.R.; Gribel, R. Extreme long-distance dispersal of the lowland tropical rainforest tree *Ceiba pentandra* L. (Malvaceae) in Africa and the Neotropics. *Mol. Ecol.* **2007**, *16*, 3039–3049. [[CrossRef](#)]
82. Lobo, J.A.; Quesada, M.; Stoner, K.E. Effects of pollination by bats on the mating system of *Ceiba pentandra* (Bombacaceae) populations in two tropical life zones in Costa Rica. *Am. J. Bot.* **2005**, *92*, 370–376. [[CrossRef](#)]
83. Mullarkey, A.A.; Byers, D.L.; Anderson, R.C. Inbreeding depression and partitioning of genetic load in the invasive biennial *Alliaria petiolata* (Brassicaceae). *Am. J. Bot.* **2013**, *100*, 509–518. [[CrossRef](#)]
84. Durka, W.; Bossdorf, O.; Prati, D.; Auge, H. Molecular evidence for multiple introductions of garlic mustard (*Alliaria petiolata*, Brassicaceae) to North America. *Mol. Ecol.* **2005**, *14*, 1697–1706. [[CrossRef](#)]
85. Ross, C.A.; Auge, H.; Durka, W. Genetic relationships among three native North-American *Mahonia* species, invasive *Mahonia* populations from Europe, and commercial cultivars. *Plant Syst. Evol.* **2008**, *275*, 219–229. [[CrossRef](#)]
86. Lachmuth, S.; Durka, W.; Schurr, F.M. The making of a rapid plant invader: Genetic diversity and differentiation in the native and invaded range of *Senecio inaequidens*. *Mol. Ecol.* **2010**, *19*, 3952–3967. [[CrossRef](#)]
87. Pairon, M.; Petitpierre, B.; Campbell, M.; Guisan, A.; Broennimann, O.; Baret, P.V.; Jacquemart, A.L.; Besnard, G. Multiple introductions boosted genetic diversity in the invasive range of black cherry (*Prunus serotina*; Rosaceae). *Ann. Bot.* **2010**, *105*, 881–890. [[CrossRef](#)]
88. Niu, H.-Y.; Hong, L.; Wang, Z.-F.; Shen, H.; Ye, W.-H.; Mu, H.-P.; Cao, H.-L.; Wang, Z.-M.; Bradshaw, C.J.A. Inferring the invasion history of coral berry *Ardisia crenata* from China to the USA using molecular markers. *Ecol. Res.* **2012**, *27*, 809–818. [[CrossRef](#)]



89. Kelager, A.; Pedersen, J.S.; Bruun, H.H. Multiple introductions and no loss of genetic diversity: Invasion history of Japanese Rose, *Rosa rugosa*, in Europe. *Biol. Invasions* **2013**, *15*, 1125–1141. [[CrossRef](#)]
90. Molins, M.P.; Corral, J.M.; Aliyu, O.M.; Koch, M.A.; Betzin, A.; Maron, J.L.; Sharbel, T.F. Biogeographic variation in genetic variability, apomixis expression and ploidy of St. John's wort (*Hypericum perforatum*) across its native and introduced range. *Ann. Bot.* **2014**, *113*, 417–427. [[CrossRef](#)]
91. Williams, J.L.; Fishman, L. Genetic evidence for founder effects in the introduced range of houndstongue (*Cynoglossum officinale*). *Biol. Invasions* **2014**, *16*, 205–216. [[CrossRef](#)]
92. Kang, M.; Buckley, Y.M.; Lowe, A.J. Testing the role of genetic factors across multiple independent invasions of the shrub Scotch broom (*Cytisus scoparius*). *Mol. Ecol.* **2007**, *16*, 4662–4673. [[CrossRef](#)] [[PubMed](#)]
93. Prentis, P.J.; Sigg, D.P.; Raghu, S.; Dhileepan, K.; Pavasovic, A.; Lowe, A.J. Understanding invasion history: Genetic structure and diversity of two globally invasive plants and implications for their management. *Divers. Distrib.* **2009**, *15*, 822–830. [[CrossRef](#)]
94. Gaudeul, M.; Giraud, T.; Kiss, L.; Shykoff, J.A. Nuclear and chloroplast microsatellites show multiple introductions in the worldwide invasion history of common ragweed, *Ambrosia artemisiifolia*. *PLoS ONE* **2011**, *6*, e17658. [[CrossRef](#)] [[PubMed](#)]
95. Hornoy, B.; Atlan, A.; Roussel, V.; Buckley, Y.M.; Tarayre, M. Two colonisation stages generate two different patterns of genetic diversity within native and invasive ranges of *Ulex europaeus*. *Heredity* **2013**, *111*, 355–363. [[CrossRef](#)]
96. Mandak, B.; Hadincová, V.; Mahelka, V.; Wildova, R. European invasion of North American *Pinus strobus* at large and fine scales: High genetic diversity and fine-scale genetic clustering over time in the adventive range. *PLoS ONE* **2013**, *8*, e68514. [[CrossRef](#)]
97. Zhao, J.; Solís-Montero, L.; Lou, A.; Vallejo-Marín, M. Population structure and genetic diversity of native and invasive populations of *Solanum rostratum* (Solanaceae). *PLoS ONE* **2013**, *8*, e79807. [[CrossRef](#)]
98. Rijal, D.P.; Alm, T.; Jahodová, Š.; Stenøien, H.K.; Alsos, I.G. Reconstructing the invasion history of *Heracleum persicum* (Apiaceae) into Europe. *Mol. Ecol.* **2015**, *24*, 5522–5543. [[CrossRef](#)]
99. Xu, C.Y.; Tang, S.; Fatemi, M.; Gross, C.L.; Julien, M.H.; Curtis, C.; Van Klinken, R.D. Population structure and genetic diversity of invasive *Phyla canescens*: Implications for the evolutionary potential. *Ecosphere* **2015**, *6*, 1–21. [[CrossRef](#)]
100. Mukherjee, A.; Williams, D.; Gitzendanner, M.A.; Overholt, W.A.; Cuda, J.P. Microsatellite and chloroplast DNA diversity of the invasive aquatic weed *Hygrophila polysperma* in native and invasive ranges. *Aquat. Bot.* **2016**, *129*, 55–61. [[CrossRef](#)]
101. Thompson, G.D.; Richardson, D.M.; Wilson, J.R.U.; Bellstedt, D.U.; Le Roux, J.J. Genetic diversity and structure of the globally invasive tree, *Paraserianthes lophantha* subspecies *lophantha*, suggest an introduction history characterised by varying propagule pressure. *Tree Genet. Genomes* **2016**, *12*, 1–9. [[CrossRef](#)]
102. Závada, T.; Malik, R.J.; Kesseli, R. V Population structure in chicory (*Cichorium intybus*): A successful US weed since the American revolutionary war. *Ecol. Evol.* **2017**, *7*, 4209–4219. [[CrossRef](#)]
103. Xia, L.; Geng, Q.; An, S. Rapid genetic divergence of an invasive species, *Spartina alterniflora*, in China. *Front. Genet.* **2020**, *11*, 284. [[CrossRef](#)]
104. Hirsch, H.; Richardson, D.M.; Pauchard, A.; Le Roux, J.J. Genetic analyses reveal complex introduction histories for the invasive tree *Acacia dealbata* Link around the world. *Divers. Distrib.* **2021**, *27*, 360–376. [[CrossRef](#)]