

# 1 **Monolayer arrangement of fatty hydroxystearic** 2 **acids on graphite: Influence of hydroxyl groups**

3 *S. Medina<sup>1</sup>, J. J. Benítez<sup>2</sup>, M. A. Castro<sup>2</sup>, C. Cerrillos<sup>3</sup>,*

4 *C. Millán<sup>2</sup>, M. D. Alba<sup>2,\*</sup>*

5 <sup>1</sup>Laboratorio de Rayos-X, Centro de Investigación Tecnología e Innovación

6 de la Universidad de Sevilla (CITIUS), Universidad de Sevilla,

7 Avenida Reina Mercedes, 4B. 41012 Sevilla. Spain.

8 <sup>2</sup>Instituto de Ciencia de Materiales de Sevilla

9 Consejo Superior de Investigaciones Científicas-Universidad de Sevilla

10 Avenida Américo Vespucio, 49. 41092 Sevilla. Spain.

11 <sup>3</sup>Servicio de Microscopía, Centro de Investigación Tecnología e Innovación

12 de la Universidad de Sevilla (CITIUS), Universidad de Sevilla,

13 Avenida Reina Mercedes, 4B. 41012 Sevilla. Spain.

14

15

16 **ABSTRACT**

17 Previous studies have indicated that long-chain linear carboxylic acids form commensurate packed  
18 crystalline monolayers on graphite even at temperatures above their melting point. This study  
19 examines the effect on the monolayer formation and structure of adding one or more secondary  
20 hydroxyl, functional groups to the stearic acid skeleton (namely, 12-hydroxystearic and 9,10-  
21 dihydroxystearic acid). Moreover, a comparative study of the monolayer formation on recompressed  
22 and monocrystalline graphite has been performed through X-ray Diffraction (XRD) and Scanning  
23 Tunneling Microscopy (STM), respectively. The Differential Scanning Calorimetry (DSC) and XRD  
24 data were used to confirm the formation of solid monolayers and XRD data have provided a detailed  
25 structural analysis of the monolayers in good correspondence with obtained STM images. DSC and  
26 XRD have demonstrated that, in stearic acid and 12-hydroxystearic acid adsorbed onto graphite, the  
27 monolayer melted at a higher temperature than the bulk form of the carboxylic acid. However, no  
28 difference was observed between the melting point of the monolayer and the bulk form for 9,10-  
29 dihydroxystearic acid adsorbed onto graphite. STM results indicated that all acids on the surface  
30 have a rectangular  $p2$  monolayer structure, whose lattice parameters were uniaxially commensurate  
31 on the  $a$ -axis. This structure does not correlate with the initial structure of the pure compounds after  
32 dissolving, but it is conditioned to favor a) hydrogen bond formation between the carboxylic groups  
33 and b) formation of hydrogen bonds between secondary hydroxyl groups, if spatially permissible.  
34 Therefore, the presence of hydroxyl functional groups affects the secondary structure and behavior  
35 of stearic acid in the monolayer.

36

37 **Keywords.** Graphite, carboxylic acid, monolayer, solid-liquid interface, VT-XRD, DSC, STM.

38

39

40

41

42

## 43 1. Introduction

44

45 Physisorption of organic liquids to a solid surface has been widely investigated because the  
46 layers formed at the surface affects the surface properties and allows understanding the processes  
47 important in many areas, including wetting, detergents, lubricants and other surface agents [1].  
48 Among the substrates tested, graphite is a good substrate as it will not chemically bond or create  
49 stronger intermolecular interactions than van der Waals forces with the adsorbate molecules. The  
50 intermolecular interactions within the adsorbed monolayer are therefore the most significant forces  
51 present [2,3].

52 Nowadays, the literature continues to highlight the importance of detailed crystal structures  
53 in understanding the behavior of adsorbed monolayers in a wide variety of situations [4]. The  
54 formation of ordered, adsorbed monolayers of alkanes, alcohols and acids was demonstrated through  
55 delicate dilatometric and calorimetric studies [5,6]. Carboxylic acids have been reported to form  
56 close-packed crystalline monolayers on graphite even at temperatures above the bulk melting point  
57 of the acid, and those of 14 to 20 carbon atoms in length (C14 - C20) were reported as producing  
58 well-developed monolayers with strongly hydrogen-bonded dimers [7,8,9].

59 The majority of acids studied have even numbers of carbon atoms and were found to form  
60 slightly oblique unit cells with plane group  $p2$  and which exhibit a positional correlation with the  
61 underlying graphite [7,8,9]. The alkyl chains interdigitate and there is a superstructure in the  
62 direction perpendicular to the chains with a repeat distance of four or five molecules. The odd  
63 members that have been studied [7], such as heptadecanoic (C17) and nonadecanoic (C19) acid, were  
64 reported to exhibit a different  $pgg$  symmetry and rectangular unit cell. Rabe et al. [10], who studied  
65 stearic (C18), arachidic (C20), and tetracosanoic (C24) acids, concluded that the mismatch  
66 parameter between the side-by-side separation of alkyl chains and the graphite lattice is  
67 approximately 10%.

68 In most cases, the structure of carboxylic acid monolayers on graphite has been resolved by  
69 scanning tunneling microscopy. More recently, X-ray and neutron diffraction have been used to

70 study such solid crystalline monolayers [4]. However both techniques introduce some difficulties.  
71 Neutron diffraction can only be carried out at few specific facilities around the world. On the other  
72 side, X-ray diffraction of monolayers deposited on graphite substrates shows serious transmission  
73 problems that restrict its application only to sub-monolayer regimes. In this article we use an  
74 experimental setup that makes the monolayer diffraction studies accessible to relatively conventional  
75 XRD diffractometers. This methodology has been previously employed to study mixture  
76 undecanoic-dodecanoic acid adsorbed on graphite surface at submonolayer regime [11].

77 This present work is a comprehensive combination of calorimetry measurements (DSC),  
78 XRD and scanning tunnel microscopy (STM) forming an in depth study into the behavior and  
79 structure of stearic and hydroxystearic acids adsorbed onto graphite surfaces. Here, we study the  
80 adsorption structures of the pure acids from their liquids and compare the results with the structure  
81 of the bulk acids. X-ray diffraction data are used to confirm the formation of solid monolayers and  
82 provide a detailed structural analysis of the monolayers formed by each of the acids under  
83 investigation. STM is the main technique employed for characterizing surface structures of self-  
84 assembled monolayer and gives many insights into the structure and dynamics of such monolayers  
85 [12,13,14]. Visualization on the atomic scale of molecular structures in real space is extremely  
86 helpful for understanding self-assembly processes, plus the solid-liquid interface is an excellent  
87 environment in which to probe them [15,16,17].

88

## 89 **2. Experimental details**

90

### 91 *2.1 Materials*

92

93 Two substrates were used in these experiments. Recompressed exfoliated graphite Papyex (Le  
94 Carbone Lorraine, France) [18] for X-ray Diffraction and Differential Scanning Calorimetry (DSC)  
95 experiments and a highly ordered pyrolytic graphite (HOPG) (SPI Supplies, USA) for Scanning

96 Tunneling Microscopy (STM). The specific surface area determined by nitrogen adsorption is 31.6  
97  $\text{m}^2 \cdot \text{g}^{-1}$  for the graphite used in XRD and DSC and  $4.8 \cdot 10^{-4} \text{m}^2 \cdot \text{g}^{-1}$  for HOPG.

98 The adsorbates were stearic acid ( $\text{C}_{18}\text{H}_{36}\text{O}_2$ , CAS: 57-11-4), 12-hydroxystearic acid  
99 ( $\text{C}_{18}\text{H}_{36}\text{O}_3$ , CAS: 106-14-9) and erythro-9,10-dihydroxystearic acid ( $\text{C}_{18}\text{H}_{36}\text{O}_4$ , CAS: 3639-32-5)  
100 obtained from Sigma Aldrich at 99% purity and used without further purification.

101 The solvents used for STM sample preparations were phenyloctane ( $\text{C}_{14}\text{H}_{22}$ , CAS: 2189-60-  
102 8) for dissolving stearic acid and 12-hydroxystearic acid and octanol ( $\text{C}_8\text{H}_{18}\text{O}$ , CAS: 111-87-5) for  
103 dissolving 9,10-dihydroxystearic acid.

104

## 105 *2.2 Sample preparation*

106 The pure stearic acid, stearic, 12-hydroxystearic acid and 9,10-dihydroxystearic acid, without  
107 graphite were studied as provided, after heating at 150 °C and cooling down up to room temperature,  
108 after drying at 25 °C a  $5\text{-}10 \text{mg} \cdot \text{cm}^{-3}$  solution of the stearic and 9,10-dihydroxystearic acid in octanol  
109 and 12-hydroxystearic acid in phenyloctane, and, after drying at 25 °C a  $5\text{-}10 \text{mg} \cdot \text{cm}^{-3}$  solution of  
110 the 150 °C heated stearic acid and 9,10-dihydroxystearic acid in octanol and 150 °C heated 12-  
111 hydroxystearic acid in phenyloctane.

112 The graphite substrates used for diffraction and calorimetry were outgassed under vacuum in  
113 an oven at 350 °C. Subsequently, a known quantity of the adsorbates were added and annealed at a  
114 temperature of 150 °C, below the bulk boiling point.

115 Total coverage was maintained at 0.9 or 3 monolayers for XRD and 60 monolayers for DSC  
116 experiments. The volumes of adsorbate required to achieve the desired level of deposition were  
117 taken from the area per molecule values of  $122 \text{Å}^2/\text{molecule}$  for stearic acid,  $129 \text{Å}^2/\text{molecule}$  for  
118 12-hydroxystearic acid and  $135 \text{Å}^2/\text{molecule}$  for 9,10-dihydroxystearic acid, estimated using the  
119 Groszek model [19,20] and the specific surface area of the graphite.

120 The surface of HOPG used for STM measurements was cleaned by cleaving with an adhesive  
121 tape. A drop of the solution (approx.  $5\text{-}10 \text{mg} \cdot \text{cm}^{-3}$ ) was immediately deposited on the support and  
122 allowed for stabilizing at 25 °C for 5-10 minutes before STM analysis.

123

### 124 *2.3 Experimental procedures*

125

126 X-ray diffraction measurements of pure acids were carried out in a Bruker D8 Advance A25  
127 diffractometer (Bruker, Germany) in Bragg-Brentano configuration. The detector used was a  
128 Lynxeye PSD detector (Bruker, Germany), with a 0.5° fix slit in the incident beam and axial Soller  
129 slits of 2.5° in the incident and diffracted beams, for copper K $\alpha$  radiation, at Centro de Investigación  
130 Tecnología e Innovación de la Universidad de Sevilla (CITIUS). Measurements were taken with a  
131 2 $\theta$  range between 3° and 120°, a step of 0.015° and a time per step of 0.1 s.

132 Variable temperature X-ray diffraction (VTXRD) of the acids adsorbed onto graphite were  
133 carried out in an Anton Paar TTK 450 low-temperature chamber (Anton Paar, Austria) attached to a  
134 Bruker D8 Advance diffractometer (Bruker, Germany), modified for symmetrical transmission  
135 geometry [21] at CITIUS, University of Sevilla, Spain. Single rectangular sheets of graphite, with  
136 dimensions of approximately 15 × 30 × 2 mm and 1 g, were irradiated by copper K $\alpha$  radiation. The  
137 device uses  $\theta/\theta$  X-ray tube and detector movement to maintain the momentum transfer in the plane  
138 of the graphite sample. Experiments were performed with parallel Johansson geometry in the  
139 incident beam, using 60 mm Göbel mirrors (Bruker, Germany) for copper K $\alpha$  radiation. The  
140 experiments were carried out at a total coverage of 0.9 and 3 monolayers for the three adsorbed acids  
141 with a 2 $\theta$  range between 17° and 24°, a step of 0.015° and a time per step of 10 s. The detector used  
142 was a Vantec PSD detector (Bruker, Germany) with radial Soller slits. The temperature range for the  
143 measurements was between 25 and 125 °C, obtaining the patterns with a temperature step of 10 °C  
144 when far from the bulk melting point and in steps of 1 °C when near to the bulk and monolayer  
145 melting temperatures.

146 Both diffractometers were calibrated mechanically according to the manufacturer  
147 specifications and corundum and silicon standards were used to check the resolution in a wide range  
148 of angles.

149 The Differential Scanning Calorimetry (DSC) measurements were performed on a Thermal  
150 Analysis Instrument Q20P systems at the Instituto de Ciencia de los Materiales de Sevilla, Spain, as  
151 discussed previously [22]. The temperature range was from 30 to 200 °C. The rate of heating was 10  
152 °C·min<sup>-1</sup>.

153 The Scanning Tunneling Microscopy (STM) images of the liquid-solid interface were taken  
154 at room temperature, using Pt/Ir 80:20% mechanically cut tips. Two microscopes were used, a  
155 Topometrix Discoverer with a scanner of 1.5 x 1.5 μm<sup>2</sup> at the Instituto de Ciencia de los Materiales  
156 de Sevilla (CSIC-US) and a Molecular Imaging with a scanner of 1 x 1 μm<sup>2</sup> at CITIUS, University  
157 of Sevilla, Spain. They were operated in constant height mode, using sample negative bias voltages  
158 ranging from -0.12 to -1.2 V. For 1-octanol, a strong ionic background current was detected and the  
159 set point was modified accordingly. The molecular packing structure was only obtained at a very low  
160 and very narrow range of tunneling current. Higher set points meant the immediate observation of  
161 the underlying graphite pattern which was used for in-situ X and Y calibration.

162

#### 163 *2.4 Calculations for determining the structural parameters of the adsorbed materials*

164

165 Since not all of the structural parameters of the three materials adsorbed onto graphite have  
166 previously been reported, the parameters that are lacking have been determined using structural  
167 parameters published for other carboxylic acids [4] with shorter chains in other studies that explain  
168 how the carboxylic acids [7] and 12-hydroxystearic acid [23] adsorbed onto graphite. The position of  
169 the atoms was deduced from the data published for undecanoic acid [4]; stretching and filling the  
170 molecular structure up to an eighteen-carbon chain with the corresponding hydrogen and oxygen  
171 atoms in their positions, including the hydroxyl or dihydroxyl groups of the branched-chain acids.

172 Previously deduced structures were verified by superimposing the above schematic STM  
173 images obtained for the three acids adsorbed on graphite. The diagrams showing the molecular  
174 structures were produced using the ATOMS programme, by Shape Software (Eric Dowty, USA).

175

### 176 3. Results and discussion

177

#### 178 3.1 Polymorphic transitions of pure acids

179

180 This section characterizes the bulk of the material in absence of graphite. Fig. 1a shows the  
181 XRD patterns of the pure acids. They were completely different depending on the presence of one or  
182 more hydroxyl groups on the alkyl chain. The Le Bail [24] fits of the patterns were performed with  
183 the software TOPAS 4.2 from Bruker [25] (Tables 1-3) using the fundamental parameters method.  
184 The zero error ( $2\theta$ ), the sample displacement, the absorption (1/cm) and the lattice parameters of the  
185 phases were allowed to vary to provide the best fitting. The background was fitted by a fifth-order  
186 Chebychev polynomial. Lorentz and polarization geometric factors for the configuration of  
187 measurement were used. From the fits was revealed that the stearic acid XRD pattern (Fig. 1a,  
188 upper) matches with the polymorph B<sub>0</sub> (orthorhombic, *Pbca*) [26] whereas, 12-hydroxystearic and  
189 9,10-dihydroxystearic acids match with polymorphs A (triclinic, *P1*) [27] and E<sub>0</sub> (orthorhombic,  
190 *Pbca*) [28], respectively. Overall a good residue was obtained as the difference between the  
191 calculated value and the experimental value for all cases. For the fit to be as accurate as possible the  
192 GOF ("Goodness of fit") should be greater than 1, and as close as possible to it [29]. Another  
193 parameter to note is the experimental residue (Rwp), which must have a value as small as possible  
194 for the measurement configuration used [29]. Good fit values were obtained.

195 An in depth analysis of the polymorph's stability with respect to temperature and solvent  
196 type has been carried out using the Le Bail fit from each of the XRD patterns (Fig. 1 and Tables 1-  
197 3). The analysis reveals that, for stearic acid (Fig. 1, upper), the polymorph B<sub>0</sub> is transformed into  
198 polymorph C (monoclinic, *P2<sub>1</sub>/a*) [30] by heating to 150 °C or by dissolving in phenyloctane. After  
199 heating to 150 °C, the polymorph C in the phenyloctane solution evolved into a mixture of the  
200 polymorphs C and B<sub>0</sub>, accompanied by an amorphous phase that is probably due to incomplete  
201 recrystallization of the polymorphs. With time, the latter evolved to a polymorph C as the most  
202 stable phase and the amorphous phase decreased significantly (Fig. 2, upper).



203 The polymorph A of 12-hydroxystearic acid (Fig. 1, middle) was stable after heating to 150  
204 °C and dissolving in phenyloctane. However, heating the solution to 150 °C produced a small  
205 amount of amorphous phase that may indicate that the recrystallization of the polymorph A is a  
206 reconstructive process.

207 In the case of 9,10-dihydroxystearic acid (Fig. 1, bottom), the polymorph E<sub>o</sub> was transformed  
208 into polymorph C by heating to 150 °C. The polymorph E<sub>o</sub> was stable after dissolving in octanol but,  
209 after heating to 150 °C, it evolved into a mixture of the polymorphs C and E<sub>m</sub> (monoclinic,  $P2_1/a$ )  
210 [31] plus an amorphous phase due to the incomplete recrystallization of the polymorphs. The latter  
211 mixture evolved to a polymorph C with time as a more stable phase and the quantity of the  
212 amorphous phase decreased significantly (Fig. 2, bottom).

213

### 214 *3.2 Formation of acid monolayers on graphite surfaces*

215

216 Fig. 3 shows the DSC thermograms for approximately 60 monolayers of the acids adsorbed  
217 on graphite. Stearic acid and 12-hydroxystearic acid (Fig. 3a and 3b) exhibited a very large peak at a  
218 temperature of 65 - 80 °C corresponding to the bulk melting point of the acid. Note, at this high  
219 coverage the temperature of the bulk transitions are identical whether graphite is present or not  
220 (Table 4). Additionally, a small peak was observed at 95 - 100 °C that arises from the melting of the  
221 adsorbed monolayer that coexisted with the bulk liquid at this temperature. The DSC plot of 9,10-  
222 dihydroxystearic acid adsorbed on graphite (Fig. 3c) showed only a wide peak at a temperature of  
223 129.5 °C due to the complete melting of the acids. These results could imply that the monolayer, if it  
224 exists, melted at temperature quite similar to the bulk and could not be resolved.

225 The temperatures and enthalpies of the monolayer transitions are given in Table 4. The  
226 values for the adsorbed layer and bulk transition temperatures are peak maximum and on-set values,  
227 respectively. The bulk transition temperature was similar to those obtained in the literature for the  
228 pure liquid acids, similarly, as observed for alkanes and alcohols adsorbed onto graphite [6,32,33,  
229 34]. The monolayer transition temperature increases as the number of hydroxyl groups increases in

230 the alkyl chain, however, the temperature difference between bulk and monolayer decreases. This  
231 decrease (in the temperature difference between bulk and monolayer transition temperatures with  
232 increasing numbers of hydroxyl groups) is compatible with the observation of a single, wide DSC  
233 peak for the 9,10-dihydroxystearic acid system but it does not conclusively demonstrate that the  
234 melting transition temperature is independent of the bulk. The enthalpies of the monolayer  
235 transitions at higher coverage, which decrease with the number of hydroxyl groups, are much  
236 smaller than the bulk melting enthalpies for each acid investigated here.

237 Figs. 4 - 6 show the evolution of the XRD patterns with temperature for the acid systems  
238 adsorbed onto graphite at a coverage level of 0.9 monolayers (left) and 3 monolayers (right). In  
239 general, 2D adsorbed material shows a “saw shaped” peak [35], while for a higher dimensionality,  
240 i.e. bulk, a symmetrical 3D structure peak is observed. Therefore, the bulk melting point occurs  
241 when the peaks change from a 3D shape to a 2D shape and the monolayer melting point takes place  
242 when the 2D shape peak disappears. The disappearance of the 2D XRD peaks, the monolayer  
243 melting, for 0.9 monolayers of stearic acid adsorbed onto graphite (Fig. 4, left) occurred at  
244 approximately 68 °C. In the case of a 3-monolayer coverage, a change of 3D XRD peaks to a 2D  
245 XRD peaks was observed at 65 °C as consequence of the bulk melting point. The 2D XRD peaks  
246 disappeared at ca. 95 °C when the monolayer melts. For the 3-monolayer coverage, the value of the  
247 monolayer melting temperature was approximately 10% higher (in Kelvin) scale) than the bulk  
248 melting temperature and also higher than the monolayer melting for submonolayer coverage, as  
249 predicted by the literature [36].

250 A similar evolution of the XRD patterns was observed for the 12-hydroxystearic acid systems  
251 adsorbed onto graphite (Fig. 5), the only difference being the temperature transition as was observed  
252 with DSC. Following the explanations in the previous paragraph, from Fig. 5 we concluded that the  
253 melting point for 0.9 monolayers was approximately 58 °C. In the case of 3 monolayers, the bulk  
254 melting point was ca. 65 °C and the monolayer melting point was ca. 94 °C. Similar to stearic acid,  
255 in the sample with 3 monolayers, the value of the monolayer melting temperature is approximately

256 10% higher (in Kelvin) than the bulk melting temperature and higher than the monolayer melting for  
257 submonolayer coverage.

258 A different evolution for the XRD patterns of the 9,10-dihydroxystearic acid system was  
259 observed. In the case of 0.9 monolayers adsorbed onto graphite, Fig. 6 left, the 2D XRD peaks  
260 disappeared at approximately 68 °C and the monolayer melts. In the case of a 3 monolayer coverage  
261 (Fig. 6, right), the 3D XRD peaks disappeared at ca. 120 °C without observation of 2D XRD peaks,  
262 as previously observed by DSC, probably due to the complete melting of the system. However,  
263 before the melting point, at ca. 78° C, the whole set of 3D XRD peaks changed their 2θ position  
264 because of a change in the structure of the acid.

265

### 266 3.3 Structures of acid monolayers on graphite surfaces

267

268 The STM images (Fig. 7) allowed a detailed description of the monolayer structures and the  
269 structural parameters to be calculated (Table 5). The structural parameters showed that the  
270 monolayer structures are uniaxial commensurate on the graphite surface along the  $a$ -axis,  $6\sqrt{3}$  for  
271 stearic acid and 9,10-dihydroxystearic acid and  $12\sqrt{3}$  for 12-hydroxystearic acid. Referring to the  
272 XRD peaks found in the literature [4] and the structural parameters calculated by STM, the 2θ  
273 position of the diffraction peaks of the planes (0,2) and (-1,2) for stearic acid have been calculated  
274 and the values obtained, 18.825° and 19.029°, agreed with the values obtained from the  
275 deconvolution of the raw XRD pattern, 18.791° and 19.514°, recorded at 25 °C (see Fig. 4, left).

276 The schematic representations of the  $p2$  structure of stearic acid, 12-dihydroxystearic acid  
277 and 9,10-dihydroxystearic acid were drawn using starting data taken from the structural parameters  
278 of shorter chain carboxylic acids adsorbed onto graphite reported in the literature [4]. For this  
279 calculation, the atom positions for stearic acid were deduced from the fractional coordinates for a  
280 single repeating motif published for undecanoic acid [4] with a  $pgg$  structure. The molecular  
281 structure was stretched and filled-out up to an eighteen-carbon chain with the corresponding  
282 hydrogen and oxygen atom positions for stearic acid 12-hydroxystearic acid and 9,10-

283 dihydroxystearic acid. As was reported in the literature [23], 12-hydroxystearic acid presented a non-  
284 interdigitised dimer structure, however, stearic acid and 9,10-dihydroxystearic acid formed  
285 interdigitised dimer structures.

286 Finally, the superimposition of the schematic illustration of the  $p2$  structure for stearic acid  
287 (Fig. 7a), 12-hydroxystearic acid (Fig. 7b) and 9,10-dihydroxystearic acid (Fig. 7c) have confirmed  
288 that the calculated structures were correct. The self-assembly of the acids on the graphite surface  
289 implied that the monolayer structures did not maintain their correlation with the view of the structure  
290 from  $a$ -axis or  $b$ -axis of the carboxylic acids used for the preparation. The more related 3D structures  
291 with different monolayers were the polymorph A for the stearic acid and 9,10-dihydroxystearic acid  
292 and polymorph  $E_m$  for 12-hydroxystearic acid [37]. Analysis of the degree of molecular packing in  
293 the  $ac$  or  $bc$  plane of the 3D polymorph was performed, choosing the plane that exhibited a packing  
294 degree closest to the 2D structure and provided a rectangular cell (Table 6). The results highlighted  
295 that stearic acid and 9,10-dihydroxystearic acid exhibit the most compact structure; possibly due to  
296 the absence of -OH groups or, for the latter, due to geometrical factors caused by the proximity of  
297 both -OH groups complicating hydrogen bonding between neighboring molecules. Rabe and  
298 Buchholz [10] have already observed that carboxylic acids adsorbed in parallel monolayers onto the  
299 graphite surface provoke at least a 10% contraction of the monolayer unit cell in comparison with  
300 that of a bulk level.

301

## 302 **4. Conclusions**

303

304 The influence of hydroxyl groups on the formation and structure of carboxylic acid  
305 monolayers on graphite surfaces has been demonstrated. This influence has been explained as a  
306 consequence of the combination of the geometrical factors and hydrogen bonding between the -OH  
307 groups of neighboring molecules. Moreover, the combination of VTXRD and STM has allowed a  
308 comparison of the monolayer structure of hydroxystearic acids on recompressed and monocrystalline  
309 graphite.

310 The DSC and VTXRD results demonstrated the presence, in the case of stearic acid and 12-  
311 hydroxystearic acid, of a monolayer with a melting temperature 10% higher than the melting  
312 temperature of the bulk. The independent melting of a monolayer has not been demonstrated in the  
313 case of 9,10-dihydroxystearic acid, where the entire system melted at the same temperature, with a  
314 3D structure change before the melting point.

315 Independent of the presence of the hydroxyl groups, the monolayer structure of the acids  
316 showed rectangular  $p2$  unit cells that were uniaxially commensurate in the  $a$ -axis. However, the 2D  
317 structures were not structurally correlated to the polymorphs described in the starting materials.

318

## 319 **Acknowledgments**

320

321 We would like to thank the DGICYT and FEDER funds (project no. CTQ 2010-14874 and  
322 CTQ 2011-24299) for their financial support.

323

## 324 **References**

- 
- [1] D.C. Duffy, A. Friedmann, S.A. Boggis and D. Klenerman, *Langmuir* 14 (1998) 6518.
- [2] G.I. Andrews, N. Hairs, A.J. Groszek, *Am. Soc. Lubric. Trans.* 15 (1972) 184.
- [3] A.J. Groszek, *Proc. R. Soc. Lond. Ser. A* 314 (1970) 473.
- [4] A.K Bickerstaffe, N.P Cheah, S.M Clarke, J.E Parker, A. Perdigon, L Messe and A. Inaba, *J. Phys. Chem. B* 110 (2006) 5570.
- [5] G.H. Findenegg, *J. Chem. Soc., Faraday Trans. I* 68 (1972) 1799.
- [6] G.H. Findenegg, *J. Chem. Soc., Faraday Trans.1* 169 (1973) 1069.
- [7] M. Hibino, A. Sumi, H. Tsuchiya and I. Hatta, *J. Phys. Chem. B* 102 (1998) 4544.
- [8] M. Hibino, A. Sumi and I. Hatta, *Jpn. J. Appl. Phys.* 34 (1995) 3354.
- [9] I. Hatta, J. Nishino, A. Sumi, A. and M. Hibino, *Jpn. J. Appl. Phys.* 34 (1995) 3930.

- 
- [10] J.P Rabe and S. Buchholz, *Science* 253 (1991) 424.
- [11] M.D. Alba, A.K. Bickerstaffe, M.A. Castro, S.M. Clarke, S. Medina, C. Millán, M.M. Orta, E. Pavón and A.C. Perdigón, *Eur. Phys. J.* 167 (2009) 151.
- [12] (a) D.M. Cyr, B. Venkataraman, G.M. Flynn, *Chem. Mater.* 8 (1996) 1600. (b) F. Tao, J. Goswami, S.L. Bernasek, *J. Phys. Chem. B* 110 (2006) 4199.
- [13] S. De Feyter, A. Gesquiere, M.M. Abdel-Mottaleb, P.C.M. Grim, F.C. De Schryver, C. Meiners, M. Sieffert, S. Valiyaveetil, K. Mullen, *Acc. Chem. Res.* 33 (2000) 520.
- [14] F. Tao, J. Goswami, S.L. Bernasek, *J. Phys. Chem. B* 110 (2006) 4199.
- [15] S. De Feyter, F.C. De Schryver, *Chem. Soc. Rev.* 32 (2003) 139.
- [16] S. De Feyter, F.C. De Schryver, *J. Phys. Chem. B* 109 (2005) 4290.
- [17] D.B. Amabilino, S. De Feyter, R. Lazzaroni, E. Gomar-Nadal, J. Veciana, C. Rovira, M.M. Abdel-Mottaleb, W. Mamdouh, P. Iavicoli, K. Psychogyiopolou, M. Linares, A. Minoia, H. Xu, J. Puigmarti-Luis, *J. Phys.: Condens. Matter* 20 (2008) 184003.
- [18] E.P. Gilbert, P.A. Reynolds, J.A. White, *J. Chem. Soc., Faraday Trans.* 94 (1998) 1861.
- [19] A.J. Groszek, *Proc. R. Soc. London A* 413 (1970) 473.
- [20] K. Morishige, T.J. Kato, *Chem. Phys.* 111 (1999) 7095.
- [21] M.D. Alba, A.K. Bickerstaffe, M.A. Castro, S.M. Clarke, S. Medina, C. Millán, M.M. Orta, E. Pavón, A. Perdigón, *Eur. Phys. J. Special Topics* 167 (2009) 151.
- [22] D. Joseph, R. Menczel, B. Prime, *Thermal analysis of polymers: fundamentals and applications*, John Wiley & Sons: New Jersey, (2009).
- [23] P. Qian, H. Nanjo, T. Yokoyama, T.M. Suzuki, *Chem. Commun.* (1999) 1197.
- [24] A. Le Bail, *A. Powder Diffraction.* 20 (2005) 316.
- [25] TOPAS 4.2 User Manual. Bruker. (2009).
- [26] F. Kaneko, H. Sakashita, M. Kobayshi, *Acta Crystallogr., Sect. C: Cryst. Struct. Commun.* C 50 (1994) 245.
- [27] T. Kamijo, H. Nagase, T. Endo, H. Ueda M. Nakagaki, *Analytical Sciences.* 15 (1999) 1291.

- 
- [28] F. Kaneko, H. Sakashita, M. Kobayashi, *Acta Crystallogr., Sect. C: Cryst. Struct. Commun.* C 50 (1994) 247.
- [29] R.A. Young (Ed.): *The Rietveld method*; IUCr Monographs on Crystallography no 5; Oxford University Press; New York. (1993).
- [30] V. Malta, G. Celotti, R. Zannetti, A.F. Martelli, *J. Chem. Soc. B* (1971) 548.
- [31] F. Kaneko, M. Kobayashi, Y. Kitagawa, Y. Matsuura, *Acta Crystallogr., Sect. C: Cryst. Struct. Commun.* 46 (1990) 1490.
- [32] K.W. Herwig, B. Matthies, H. Taub, *Phys. Rev. Letters.* 75 (1995) 3154.
- [33] P. Espeau, P.A. Reynolds, T. Dowling, D. Cookson, J.W. White, *J. Chem. Soc., Faraday Trans.* 93 (1997) 3201.
- [34] G.S. Wang, S. Lei, S. De Feyter, R. Feldman, J.E. Parker, S.M. Clarke, *Langmuir* 24 (2008) 2501.
- [35] B.E. Warren, *Phys. Rev.* 59 (1941) 693.
- [36] M.A. Castro, S.M. Clarke and A. Inaba, *J. Phys. Chem. B* 1997, **101**, 8878-8882
- [37] S. Medina, PhD Thesis, University of Sevilla, (2012).

**Table 1.** Unit cell parameters of stearic acid after different treatments.

Sample	Polym.	Spatial group	a (Å)	b (Å)	c (Å)	$\beta$ (°)	Cell unit Volume (Å <sup>3</sup> )
raw	B <sub>0</sub>	<i>Pbca</i>	7.36339(60)	5.52351(56)	87.7820(39)	--	3570.25(49)
heated	C	<i>P2<sub>1</sub>/a</i>	9.32000(73)	4.97311(24)	50.3169(42)	127.5713(52)	1848.46(26)
solved	C	<i>P2<sub>1</sub>/a</i>	9.3805(22)	4.9873(10)	50.858(12)	128.1989(56)	1869.80(73)
heated / solved	B <sub>0</sub>	<i>Pbca</i>	7.43609(23)	5.58196(19)	88.6659(29)	--	3680.34(21)
	C	<i>P2<sub>1</sub>/a</i>	9.37962(90)	4.97941(52)	50.9474(48)	128.2427(65)	1868.85(36)

**Table 2.** Unit cell parameters of 12-hydroxystearic acid after different treatments.

Sample	Polym.	Spatial group	a (Å)	b (Å)	c (Å)
raw	A	<i>PI</i>	8.03054(65)	48.3912(44)	4.85065(36)
heated	A	<i>PI</i>	8.1768(35)	48.784(22)	4.9119(22)
solved	A	<i>PI</i>	8.1459(20)	49.540(12)	4.9601(11)
heated / solved	A	<i>PI</i>	8.0657(10)	48.4096(60)	4.89313(62)

Sample	$\alpha$ (°)	$\beta$ (°)	$\gamma$ (°)	Cell unit Volume (Å <sup>3</sup> )
raw	91.0581(70)	90.052(10)	104.9511(73)	1820.84(27)
heated	91.229(15)	90.616(24)	105.692(12)	1885.6(14)
solved	90.5349(84)	91.4609(90)	105.1133(89)	1931.50(81)
heated / solved	90.6138(38)	90.5938(47)	105.1308(41)	1844.04(40)



**Table 3.** Unit cell parameters of 9,10 dihydroxystearic acid after different treatments.

Sample	Polym.	Spatial group	a (Å)	b (Å)	c (Å)	$\beta$ (°)	Cell unit Volume (Å <sup>3</sup> )
raw	E <sub>o</sub>	<i>Pbca</i>	7.26799(53)	5.64513(38)	83.7057(34)	--	3434.34(37)
heated	C	<i>P2<sub>1</sub>/a</i>	9.36114(65)	4.96706(22)	50.6083(38)	127.8708(50)	1857.57(24)
solved	E <sub>o</sub>	<i>Pbca</i>	7.38007(71)	5.73232(56)	83.5647(63)	--	3535.20(55)
heated / solved	E <sub>m</sub>	<i>P2<sub>1</sub>/a</i>	5.60451(12)	7.40538(15)	49.8449(10)	117.2708(13)	1843.952(77)
	C	<i>P2<sub>1</sub>/a</i>	9.35621(22)	4.97341(12)	50.8273(12)	128.2263(13)	1857.963(83)

**Table 4.** Temperatures and enthalpies of transitions of the bulk and monolayer of pure acids at coverage of approximately 60 monolayers.<sup>a</sup>

Samples	T <sub>3D</sub> <sup>lit</sup> (°C)	T <sub>3D</sub> <sup>DSC</sup> (°C)	T <sub>2D</sub> <sup>DSC</sup> (°C)	$\Delta H_{2D}$ (J/g)
Stearic acid	67-72	67.0 69.4	95.8	0.682
12-hydroxystearic acid	74-76	79.6	100.8	0.598
9,10-dihydroxystearic acid	--	127.2 129.5	--	--

<sup>a</sup>T<sub>2D</sub><sup>DSC</sup> monolayer transition temperature, T<sub>3D</sub><sup>DSC</sup> “bulk” transition temperature from DSC, T<sub>3D</sub><sup>lit</sup>, literature value of bulk melting point, and  $\Delta H_{2D}$  monolayer transition enthalpy

**Table 5.** Structural parameters for the adsorbed monolayers of the stearic acid, the 12-hydroxystearic acid and the 9,10-dihydroxystearic acid.

Samples	a (nm)	b (nm)	$\nu$ (°)	tilt (°)	2D structure
Stearic acid	2.50	0.94	94	-6	p2
12-hydroxystearic acid	5.10	0.50	97	0	p2
9,10-dihydroxystearic acid	2.60	1.06	94	-6	p2

**Table 6.** Total area, molecule amount for unit cell and packing degree.

Sample	structure	Plane angle	Area ( $\text{\AA}^2$ )	Z	Molec. per area unit ( $\text{molec}/\text{\AA}^2$ ) $\times 10^2$
Stearic acid	Polymorph C	$\beta$	374.92	4	1.07
	2D p2	$\nu$	234.43	2	0.85
	Polymorph A	$\alpha$	245.71	2	0.81
12-hydroxystearic acid	Polymorph A	$\alpha$	245.71	2	0.81
	2D p2	$\nu$	278.41	4	1.44
	Polymorph E <sub>m</sub>	$\beta$	248.31	4	1.61
9,10-dihydroxystearic acid	Polymorph E <sub>o</sub>	$\beta$	616.71	8	1.30
	2D p2	$\nu$	311.24	2	0.64
	Polymorph A	$\alpha$	245.71	2	0.81

## FIGURE CAPTIONS

**Fig. 1.** XRD diffraction of pure stearic acid (a, upper), 12-hydroxystearic acid (b, middle) and 9,10-dihydroxystearic acid (c, bottom): 1) as provided, 2) after heating at 150 °C, 3) after drying at 25 °C a 5-10 mg·cm<sup>-3</sup> solution of the 1 component in octanol or phenyloctane, and, 4) after drying at 25 °C a 5-10 mg·cm<sup>-3</sup> solution of the 2 component in octanol or phenyloctane.

**Fig. 2.** XRD patterns of stearic acid solved in phenyloctane and heated at 150 °C (upper) and 9,10-dihydroxystearic acid solved in octanol and heated at 150 °C (bottom) as dried (a) and after 30 days at 25 °C (b).

**Fig. 3.** DSC thermograms for approximately 60 monolayers of stearic acid (a), 12-hydroxystearic acid (b) and 9,10-dihydroxystearic acid adsorbed on graphite.

**Fig. 4.** Experimental VT XRD patterns (points) and best fitting functions (lines) for 0.9 monolayers (left) and 3 monolayers (right) of stearic acid adsorbed on graphite. For 3 monolayers, bulk melting point is observed at *b*, and in both cases the melting point of the monolayer is observed at *a* and *c* respectively.

**Fig. 5.** Experimental VT XRD patterns (points) and best fitting functions (lines) for 0.9 monolayers (left) and 3 monolayers (right) of 12-hydroxystearic acid adsorbed on graphite. For 3 monolayers, bulk melting point is observed at *b*, and in both cases the melting point of the monolayer is observed at *a* and *c* respectively.

**Fig. 6.** Experimental VT XRD patterns (points) and best fitting functions (lines) for 0.9 monolayers (left) and 3 monolayers (right) of 9,10-dihydroxystearic acid adsorbed on graphite. For 3 monolayers, phase change is observed at *b* and bulk melting point at *c*. For 0.9 monolayers, melting point of the monolayer is observed at *a*.

**Fig. 7.** Obtained STM images and the superimposition of the schematic illustration of the *p2* structure for stearic acid (a), 12-hydroxystearic acid (b) and 9,10-dihydroxystearic acid (c).

Figure 1

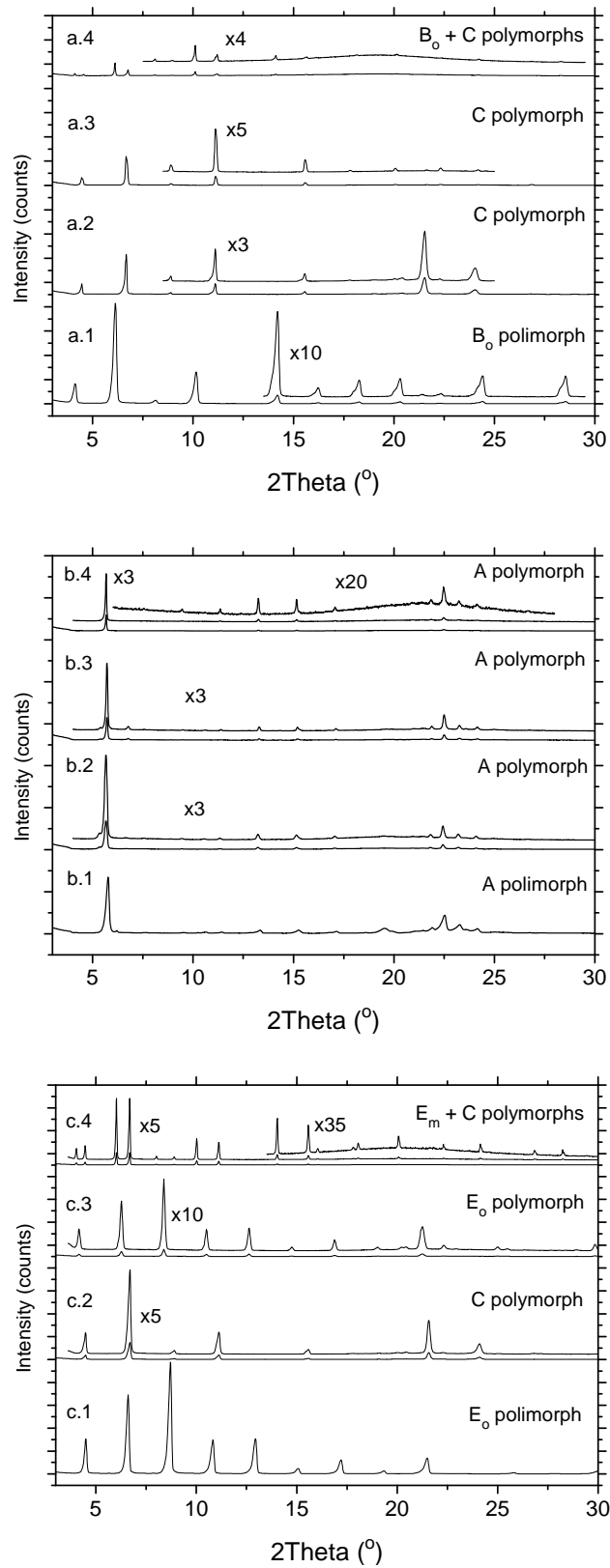


Figure 2

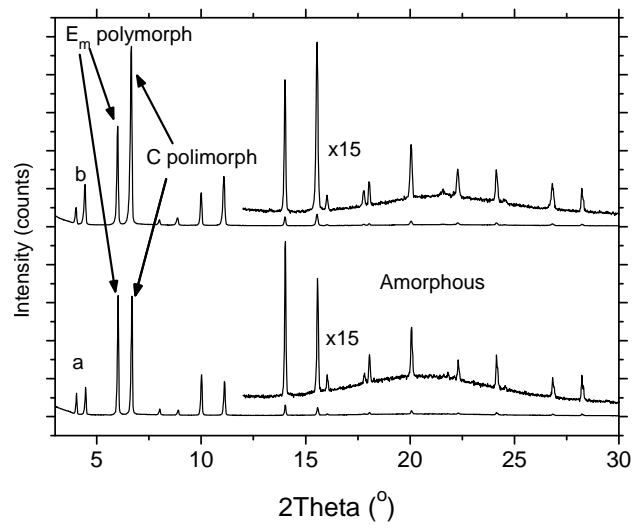
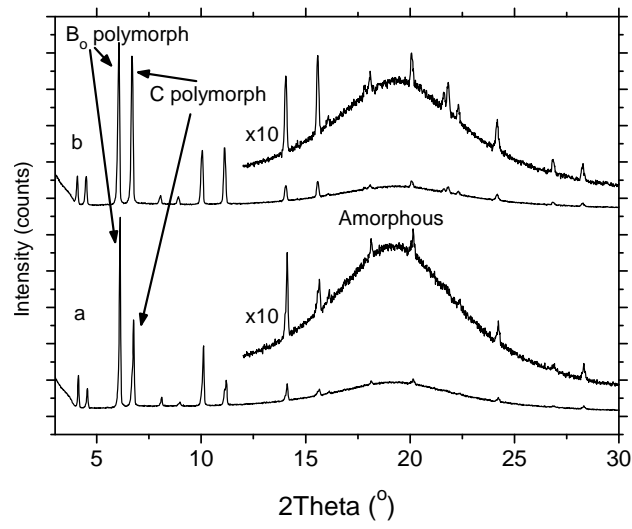


Figure 3

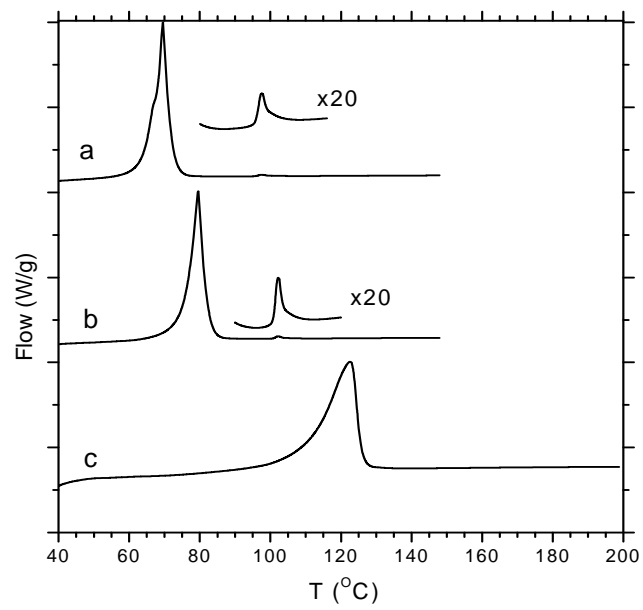


Figure 4

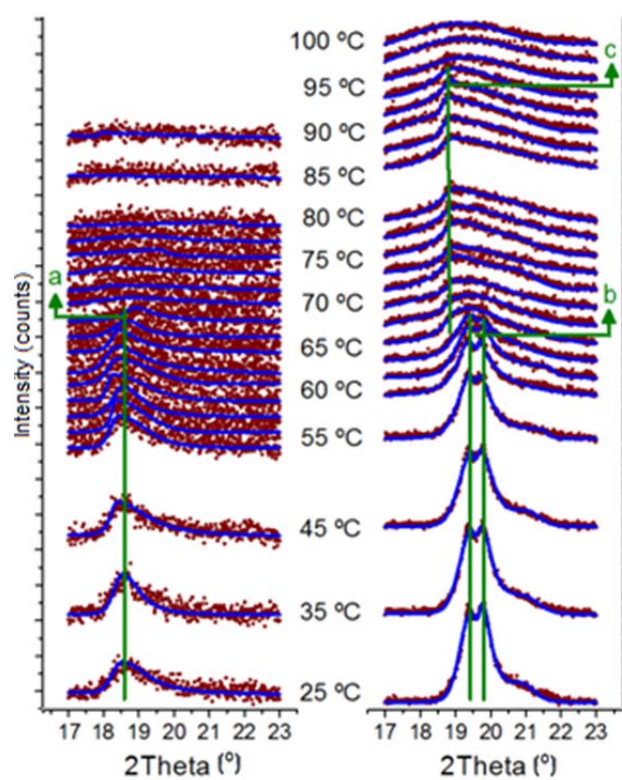




Figure 5

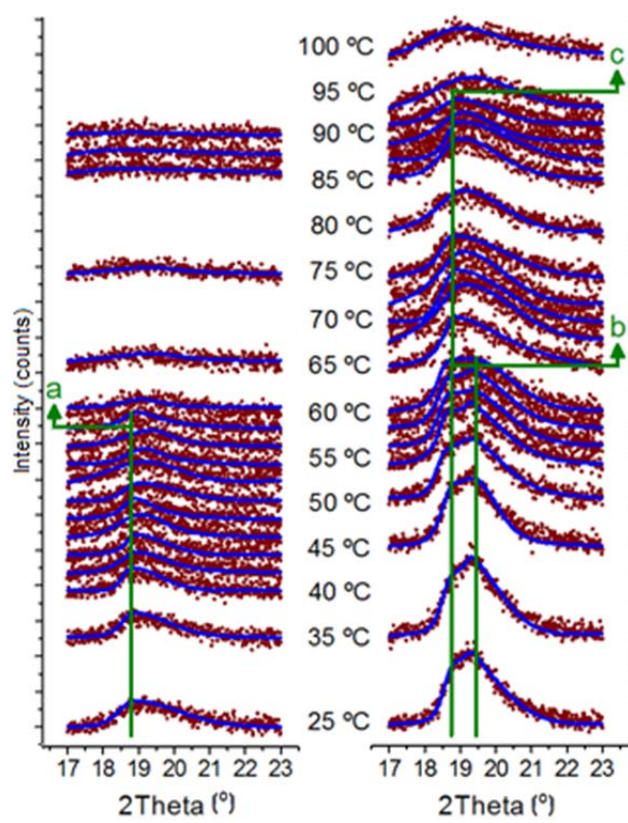


Figure 6

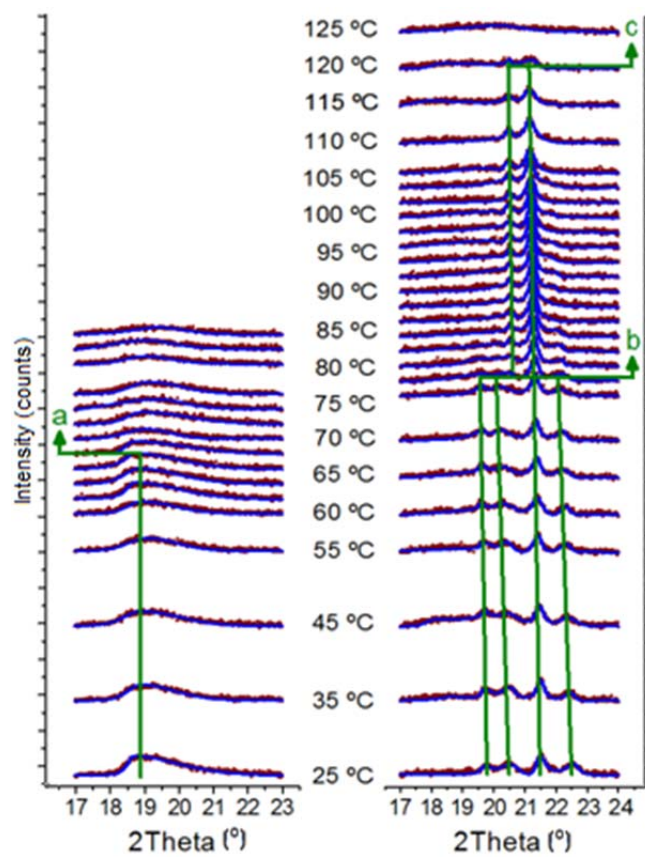


Figure 7

